MAHATMA GANDHI UNIVERSITY

School of Biosciences

Priyadarsini Hills P. O.,Kottayam–686560

Learning Outcomes based Curriculum Framework (LOCF) for PostGraduateProgramme

MSc Biophysics

Under the CSS scheme for University

(EFFECTIVE FROM 2021 ADMISSIONS)

Preface

Mahatma Gandhi University

Mahatma Gandhi University is an Indian collegiate public University based in Kerala, established in 1983, approved by UGC, and accredited with NAAC "A" Grade, 3.24 CGPA. With its academic excellence, the University has bagged Chancellor's Award twice for the best University (2015-16 and 2017-18) within the state of Kerala. It has also secured 30th position in NIRF ranking (April 2019) and 11th position in India Today-MDRA ranking, 2018. CSIR has ranked the University 13th for its intellectual productivity and NISTADS has rated it as 19th in terms of h-index.

At present, Mahatma Gandhi University offers research programs in forty disciplines through its own Schools and approved Research Centers. It has close collaboration for academic, research and extension programs with a number of national agencies and institutions including the UGC, DST-FIST, DRS, ISRO, COSIT, DIT, DST (Nano Mission), CSIR, DAAD, STEC, ICMR, BARC and MOEF. The University is also involved in active collaboration with research institutions of international reputation such as the Max Planck Institute of Technology, Germany; Brown University, USA; University of Nantes, France; California Institute of Technology, USA; University of Toronto, Canada; Catholic University, Belgium; Heidelberg University, Germany; the Institute of Political Studies, Rennes, France; Trent University, Canada; IPF Dresden, Germany; University of Paris and University of Strasbourg.

Mahatma Gandhi University has made immense strides in the fields of inter disciplinary teaching and research. The faculty comprises of outstanding scholars, many of whom have made original contributions in their respective fields of specialization. The faculty andresearch scholars of several departments have gained widespread recognition for the commendable quality of their research publications. The web enabled University library has large collection of books, journals, e-journals and online theses. The digital library provides open access to its enviable collection of digitized Ph.D dissertations. All these work in tandem with the academic business transacted by the University, making the whole experience a holistic one. The University has a well established instrumentation facility with many sophisticated equipments functioning at the various departments and also at the platform provided by the common Inter University Instrumentation Centre (IUIC).

The University has well established and internationally reputed facility and academic expertise in various areas like Nanoscience, Environmental science, Bioscience, Chemical science, Physics, Arts and Humanities.The Centre for Nanoscience and Nanotechnology focus on the enhancement of research and higher studies in the cutting edge areas of Nanoscience and Nanotechnology. The Centre is motivated to thrust its research and development focusing on developing novel materials and devices prospering the outrage of Nanoscience. With a vision to consolidate the existing and to pay focus attention to the frontier areas of Environmental Science, the University has established the School of Environmental Sciences as a Centre of learning for advanced studies in different branches of environmental science. The major mandate of the School is to develop appropriate technologies and skilled human resource for sustainable utilization, management and conservation of natural resources. The school has established a Centralized Remote Sensing and GIS facility, the first ofits kind in a University in the state, with the support of Indian Space Research Organization (ISRO). It has also established a regional center, the Highrange Environmental Research center (HERC) at Nedumkandam, Idukki district. The School has a live laboratory named as “Jeevaka” which consists of areas with rich biodiversity within the Mahatma Gandhi University Campus.

Vision and Mission of MGU

Vision of Mahatma Gandhi University

"Mahatma Gandhi University envisions to excel in the field of higher education and cater to the scholastic and developmental needs of the individual, through continuous creation of critical knowledge base for the society's sustained and inclusive growth."

Mission of Mahatma Gandhi University

- **To conduct and support undergraduate, postgraduate and research-level programmes of quality in different disciplines**
- **To foster teaching, research and extension activities for the creation of new knowledge for the development of society**
- **To help in the creation and development of manpower that would provide intellectual leadership to the community**
- **To provide skilled manpower to the professional, industrial and service sectors in the country so as to meet global demands**
- **To help promote the cultural heritage of the nation and preserve the environmental sustainability and quality of life**
- **To cater to the holistic development of the region through academic leadership**

Preamble

OUTCOME BASED EDUCATION (OBE)FROM THE ACADEMIC YEAR 2020-21
MAHATMA GANDHI UNIVERSITY SCHOOL OF BIOSCIENCES

Introduction

A high priority task in the context of education in India is improvement of quality of higher education for equipping young people with skills relevant for global and national standards and enhancing the opportunities for social mobility. Mahatma Gandhi University has initiated an Outcome Based Education (OBE) for enhancing employability of graduates through curriculum reforms based on a learning outcomes-based curriculum framework, upgrading academic resources and learning environment.

Learning outcomes specify what graduates completing a particular programme of study are expected to know, understand and be able to do at the end of their programme of study. The fundamental premise underlying the learning outcomes-based approach to curriculum development is that higher education qualifications are awarded on the basis of demonstrated achievement of outcomes, expressed in terms of knowledge, understanding, skills, attitudes and values. Outcomes provide the basis for an effective interaction among the various stakeholders. It is the results-oriented thinking and is the opposite of input-based education where the emphasis is on the educational process.

Benefits of OBE

1. The OBE Framework is a paradigm shift from traditional education system into OBE system where there is greater focus on programme and course outcomes. It guarantees that curriculum, teaching and learning strategies and assessment tools are continuously enhancedthrough a continuous improvement process. All decisions including those related to curriculum,delivery of instruction and assessment are based on the best way to achieve the predeterminedoutcomes. Traditionally, educators have measured learning in terms of standardised tests. In contrast, outcome-based education defines learning as what studentscan demonstrate that they know.

Benefits of OBE:

*More directed & coherent curriculum.

*Graduates will be more "relevant" to industry & other stakeholders (more well-rounded Graduates)

*Continuous Quality Improvement is in place.

*OBE shifts from measuring input and process to include measuring the output (outcome)

Outcome Based Education (OBE) process

OBE is a comprehensive approach to organise and operate a curriculum that is focused onand defined by the successful demonstrations of learning sought from each learner. The term clearly means focusing and organising everything in an education system around "what is essential for all learners to be able to do successfully at the end of their learning experiences".

OBE is an approach to education in which decisions about the curriculum and instruction are driven by the exit learning outcomes that the students should display at the end of a programme or a course. By the end of educational experience, each student should have achievedthe outcomes.

Learning Outcomes based Curriculum Framework (LOCF) for Post Graduate Programmes-

IQAC MG University

One of the main objectives of OBE is to ensure continuous improvement of programmes in terms of maintaining the relevance in curriculum as well as responding to the requirements of the stakeholders. In other words, it ensures that Post graduate programme next year is better than Post graduate programme this year, offered by a department.

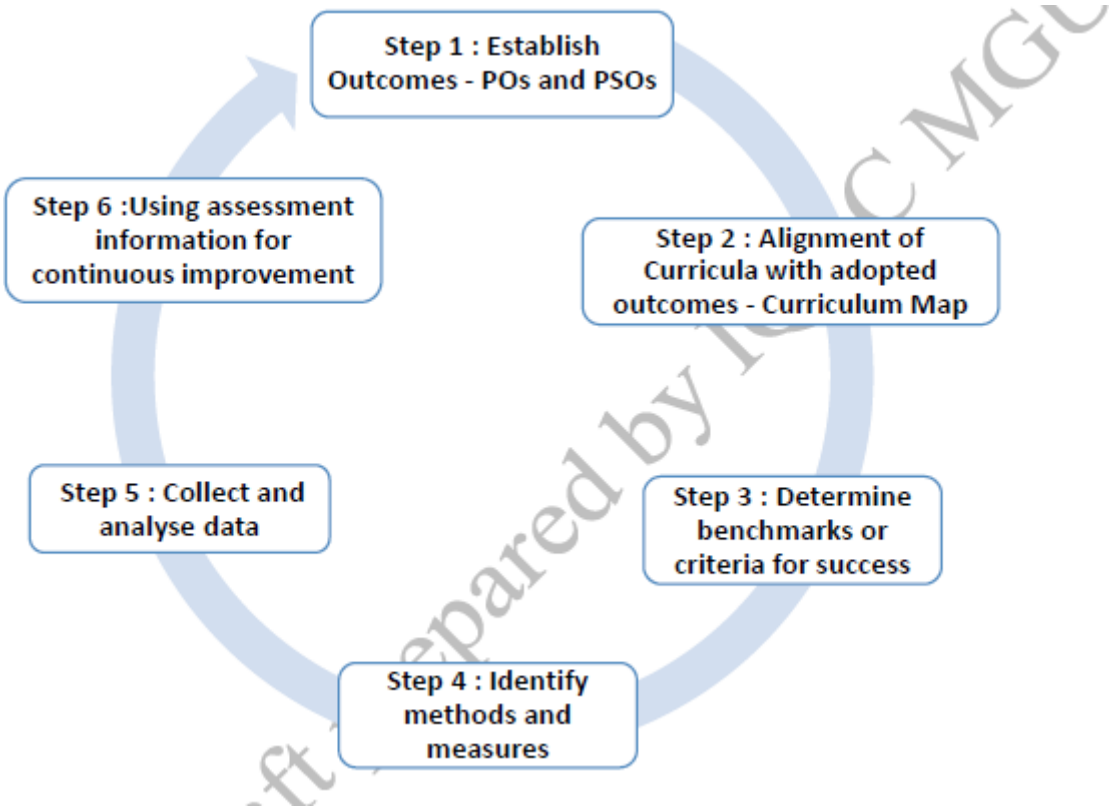
An OBE system has been proposed and to be implemented at various Departments of Mahatma Gandhi University, as a quality-assurance approach to improve teaching and learning

outcomes and processes. This OBE plan incorporates the "outcomes assessment" process to be followed in the departments. OBE should be a key driver of the curriculum management in allthe departments of the university.

The OBE is a 6 step process as shown in the figure

Figure: OBE Process

The process is presented as a cycle or a loop. The cycle represents the continuous nature of assessing learning outcomes.



As envisaged by the IQAC of Mahatma Gandhi university, an OBE based curricular framework has been proposed for the School of Biosciences from the academic year 2020-2021 which is presented hereafter.

School of Biosciences

The Life Science research of the University is carried out under the School of Biosciences, which is another prestigious department of the University and it provides academic expertise to students in advanced areas of Biochemistry, Microbiology, Biotechnology and Biophysics. The established research areas at School of Biosciences specifically include the Bioprocess technology, toxicology, ethnopharmacology, inflammation, ecology, ecotechnology, agricultural microbiology, immunobiology, medicinal plant research, probiotic development, microbial and natural product research, molecular microbiology etc. The department harbours a state -of-the-art instrumentation facility, animal maintenance facility and animal cell culture facility as well. The institute has been a successful aspirant in producing a large number of PhDs,and has completed several funded projects with significant number of publications.

Our Vision

* An Institution of excellence developing professional competence, ambition and determination in students to face new challenges and find new opportunities in the field of Biological Sciences and facilitating the wellbeing and prosperity of mankind especially our Mother Land by utilising the opportunities in advanced Biological research.

Key points

1. Institution of excellence
2. Professional competence, ambition and determination
3. New challenges and new oppurtunities
4. Well being and prosperity of nation and humanity
5. Utilise opportunities in research

Our Mission

* To provide advanced knowledge and technological knowhow to the students in the field ofBiological sciences.
* To utilise the expertise of faculty in diverse areas of biology for benefitting the students inachieving their career goals.
* To conduct cutting-edge research in areas of life Sciences and to extend the knowledge gainedfrom lab to land and benchtop to bedside.

Key points

1. provide advanced knowledge and technological knowhow
2. To utilise the expertise of faculty
3. benefitting the students in achieving their career goals.
4. conduct cutting-edge research
5. extend the knowledge gained from lab to land and benchtop to bedside.

	Mahatma Gandhi University **Graduate attributes**

	**Critical thinking and analytical reasoning**	Capability to analyze, evaluate and interpret evidence, arguments, claims, beliefs on the basis of empirical evidence; reflect relevant implications to the reality; formulate logical arguments; critically evaluate practices, policies and theories to develop knowledge and understanding; able to envisage the reflective thought to the implication on the society.
	**Scientific reasoning and Problem solving**	Ability to analyze, discuss, interpret and draw conclusions from quantitative/qualitative data and experimental evidences; and critically evaluate ideas, evidence and experiences from an unprejudiced and reasoned perspective; capacity to extrapolate from what one has learned and apply their competencies to solve problems and contextualize into researchand apply one's learning to real life situations.
	**Multidisciplinary/ Interdisciplinary/ Transdisciplinary approach**	Acquire interdisciplinary /multidisciplinary/ transdisciplinary knowledge base as a consequence of the learning they engage with their programme of study; develop a collaborative-multidisciplinary/interdisciplinary/transdisciplinary-approach for formulate constructive arguments and rational analysis for achieving common goals and objectives.
	**Intra and Interpersonal skills**	Ability to work effectively and respectfully with diverse teams;facilitate collaborative and coordinated effort on the part of a group,and act together as a group or a team in the interests of a common cause and work efficiently as a member of a team; lead the team to guide people to the right destination, in a smooth and efficient way.
	**Digital literacy**	Capability to use ICT in a variety of learning situations, demonstrate ability to access, choose, collect and evaluate, and use a variety of relevant information sources; structure and evaluate those data for decision making.

	**Global Citizenship**	Building a sense of belonging to a common humanity and to become responsible and active global citizens. Appreciation and adaptation of different sociocultural setting and embrace and promote equity.
	**Social competency**	Possess knowledge of the values and beliefs of multiple cultures, appreciate and adapt to a global perspective; and capability to effectively engage in a multicultural society and interact respectfully, manage and lead with diverse groups.
	Equity, Inclusiveness and Sustainability	Appreciate and embrace equity, inclusiveness and sustainability and diversity; acquire ethical and moral reasoning and values of unity, secularism and national integration to enable to act as dignified citizens; able to understand and appreciate diversity
	**Lifelonglearning**	Continuous acquisition of knowledge and skills. Learn, unlearn and re-learn based on changing ecosystem. “Learning how to learn”, that are necessary for participating in learning activities throughout life, through self-paced and self-directed learning aimed at personal development, meeting economic, social and cultural objectives, and adapting to changing trades and demands of work place through knowledge/skill development/reskilling.

	Mahatma Gandhi University **Programme Outcome**

Programme Outcomes (PO)

PO 1: Critical Thinking and Analytical Reasoning
Capability to analyse, evaluate and interpret evidence, arguments, claims, beliefs on the basis of empirical evidence; reflect relevant implications to the reality; formulate logical arguments; critically evaluate practices, policies and theories to develop knowledge and understanding; able to envisage the reflective thought to the implication on the society.

PO 2 : Scientific Reasoning and Problem Solving
Ability to analyse, discuss, interpret and draw conclusions from quantitative/qualitative data and experimental evidences; and critically evaluate ideas, evidence and experiences from an unprejudiced and reasoned perspective; capacity to extrapolate from what one has learned and apply their competencies to solve problems and contextualise into researchand apply one's learning to real life situations.

PO 3: Multidisciplinary/Interdisciplinary/Transdisciplinary Approach
Acquire interdisciplinary /multidisciplinary/transdisciplinary knowledge base as a consequence of the learning they engage with their programme of study; develop a collaborative-multidisciplinary/interdisciplinary/transdisciplinary-approach for formulate constructive arguments and rational analysis for achieving common goals and objectives.

PO 4: Communication Skills
Ability to reflect and express thoughts and ideas effectively in verbal and nonverbal way; Communicate with others using appropriate channel; confidently share one's views and express herself/himself; demonstrate the ability to listen carefully, read and write analytically, and present complex information in a clear and concise manner and articulate in a specific context of communication.

PO 5: Leadership Skills
Ability to work effectively and lead respectfully with diverse teams; setting direction, formulating goal, building a team who can help achieve the goal, motivating and inspiring team members to engage with that goal, and using management skills to guide people to the right destination, in a smooth and efficient way.

PO 6: Social Consciousness and Responsibility
Ability to contemplate of the impact of research findings on conventional practices, and a clear understanding of responsibility towards societal needs and reaching the targets for attaining inclusive and sustainable development.

PO 7: Equity, Inclusiveness and Sustainability
Appreciate equity, inclusiveness and sustainability and diversity; acquire ethical and moral reasoning and values of unity, secularism and national integration to enable to act as dignified

citizens; able to understand and appreciate diversity, managing diversity and use of an inclusive approach to the extent possible.

PO 8: Moral and Ethical Reasoning
Ability to embrace moral/ethical values in conducting one's life, formulate a position/argument about an ethical issue from multiple perspectives, and use ethical practices in all work. Capable of demonstrating the ability to identify ethical issues related to one's work and living as a dignified person in the society.

PO 9: Networking and Collaboration
Acquire skills to be able to collaborate and network with scholars in an educational institutions, professional organizations, research organizations and individuals in India and abroad.

PO 10: Lifelong Learning
Ability to acquire knowledge and skills, including "learning how to learn", that are necessary for participating in learning activities throughout life, through self-paced and self-directed learning aimed atpersonal development, meeting economic, social and cultural objectives, and adapting to changing trades and demands of work place through knowledge/skill development/reskilling.

Programme outcome of MSc courses in School of Biosciences (PO)

To develop competent personnel in applied branches of life sciences with good academic standards, skill, technical knowhow, research aptitude, scientific ethics and societal consciousness.

Programme specific outcomes of M.Sc Biophysics

Programme specific Outcomes (PSOs) of Msc Biophysics

PSO1: Establish high academic standards by theoretical understanding and practical proficiency in the molecular and functional aspects, as well as the physiological, cellular, and biochemical functions and organization of biological systems.

PSO2: Develop proficiency in utilizing instrumentation, biophysical techniques, and analyzing biomolecules to enhance comprehension of dynamics within biological systems and processes.

PSO3: Recognise the importance of various biological databases, algorithms and molecular docking software used in *Insilco* drug discovery.

PSO4: Foster a high-level research aptitude that allows us to skillfully design, execute, and analyze research problems using statistical tools. Ensure the derivation of meaningful scientific conclusions while upholding the principles of scientific ethics.

PSO5: Acquire the capacity to effectively communicate and present a chosen subject or research problem, both in written form and verbally, while maintaining awareness of societal implications.

Programme Specific Outcomes (PSO) to Programme Outcomes (PO) Mapping - MSc Biophysics

PSO	Programme Specific Outcomes (PSO)	MGU PO
1	**Programme specific Outcomes (PSOs) of MSc Biophysics** **PSO1**: Establish high academic standards by theoretical understanding and practical proficiency in the molecular and functional aspects, as well as the physiological, cellular, and biochemical functions and organization of biological systems.	**PO 1, PO2 PO9, PO10**
2	**PSO2**: Develop proficiency in utilizing instrumentation, biophysical techniques, and analyzing biomolecules to enhance comprehension of dynamics within biological systems and processes.	**PO1, PO2 PO3, PO9 PO10**
3	**PSO3**: Recognise the importance of various biological databases, algorithms and molecular docking software's used in *Insilco* drug discovery	**PO1, PO2 PO3, PO9 PO10**
4	**PSO4:** Foster a high-level research aptitude that allows us to skillfully design, execute, and analyze research problems using statistical tools. Ensure the derivation of meaningful scientific conclusions while upholding the principles of scientific ethics.	**PO1, PO2 PO3, PO6 PO7, PO8 PO9, PO10**
5	**PSO5**: Acquire the capacity to effectively communicate and present a chosen subject or research problem, both in written form and verbally, while maintaining awareness of societal implications.	**PO3, PO4 PO5, PO6 PO7, PO9**

SCHEME OF MSc BIOPHYSICS PROGRAMME

FIRST SEMESTER SCHEME

Course Code	Course Title	Credits
BSM 21C 01	Biochemistry	3
BSM 21C 02	Microbiology	3
BSM 21C 03	Cell Biology, Genetics &Evolution	3
BSM 21C 04	Biophysics &Biostatistics	3
BSM 21C 05	Physiology	3
	Entry level orientation programme in applied life sciences	
BSM 21C 06	Laboratory Course – 1	2
BSM 21C 07	Laboratory Course – 2	2

SECOND SEMESTER SCHEME

BSM 21 C 08	Immunology	3
BSM 21 C 09	MolecularBiologyand GeneticEngineering	3
BSM 21 C 10	MetabolismandBioenergetics	3
BSM 21 C 11	BiophysicalTechniquesandBioinstrumentation	3
BSM 21 C 12	LaboratoryCourse–3	3
BSM 21 C 13	LaboratoryCourse–4	3
Elective 1	To be selected from among the elective courses offered	3
Elective 2	To be selected from among the elective courses offered	3
Total Credits for the Second Semester Programme in M.Sc. Biophysics		**22**
List of elective courses for the Second Semester		

BSM21E 14	Microbial Technology	3
BSM21E 15	Ecology and Environment	3
BSM21E 16	Neurobiology	3
BSM21E 17	Environment Science	3
BSM21E 18	Molecular Microbiology	3
BSM21E 19	Developmental Biology	3

SCHEME OF THIRD SEMESTER BIOPHYSICS

Course No	**Subject of the Course**	Credit
BSM 21C 20	Enzymology	4
BSM 21C 36	Molecular Biophysics	4
BSM 21C 37	Electrophysiology	4
BSM 21C 38	Laboratory Course 5 Biophysics	2
BSM 21C 39	Laboratory Course 6 Biophysics	2
Course taken by the student from other department	Open course	4
Total Credits of the 3rd Semester Programme in M Sc Biophysics		20

OPEN Courses

OFFERED BY SCHOOL OF BIOSCIENCES

FOR STUDENTS OF OTHER SCHOOLS

SCHEME OF THIRD SEMESTER OPEN ELECTIVE COURSES

Students need to select one open elective course offered by other departments

	Course No.	**Subject of the Course**	**Credits**
	BSM 21O 40	Biotechnology and Society	4
	BSM 21O 41	Microbiology in Everyday Life	4
	BSM 21O 42	Environment Lead Auditor Course	4
	BSM 21O 43	System Biology	4

	BSM 21O 44	Ecology of Soil Fertility	4
	BSM 21O 45	Infectious Disease Management	4
	BSM 21O 46	Probiotics and Nutraceuticals	4
	BSM 21O 47	History and Philosophy of Science	4
	BSM 21O 48	Organic Farming For sustainability	4

SCHEME OF FOURTH SEMESTER BIOPHYSICS		
Course No	**Subject of the Course**	Credit
BSM 21C 58	Biophysical chemistry	3
BSM 21C 59	Laboratory Course 7 Biophysics	3
BSM 21C 60	Major Research Project	7
Elective 1	To be selected from among the elective courses offered	3
Elective 2	To be selected from among the elective courses offered	3
	Internship Programme of 1-2 weeks	
Total Credits of the 4th Semester Programme in M Sc Biophysics		19

SCHEME OF FOURTH SEMESTER ELECTIVE COURSES **Students need to select any two of the following elective courses**		
BSM 21E 61	Quality Control in Herbal Drugs	3
BSM 21E 62	Environment Biotechnology	3
BSM 21E 63	IPR and Patenting	3
BSM 21E 64	Omics in Biotechnology	3
BSM 21E 65	Molecular Phylogeny	3
BSM 21E 66	Human Virology	3
BSM 21E 67	Advanced Techniques in Diagnostic Microbiology	3
BSM 21E 68	Radiation Biophysics	3
BSM 21E 69	Good Laboratory Practices	3
BSM 21E 70	Health and Nutrition	3
BSM 21E 71	Neutrophil Biology	3
BSM 21E 72	Plant Microbe Interactions	3

BSM 21E 73	Sustainable Agriculture	3

MAHATMA GANDHI UNIVERSITY

BSM 21C 01: BIOCHEMISTRY

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**BIOCHEMISTRY**					
Type of Course	Core					
Course Code	**BSM 21C 01**					
Names of Academic Staff & Qualifications	1. **Prof. M S Latha – M. Sc (Biochemistry), PhD** 2. **Dr. Anie Y – M. Sc (Biochemistry), PhD** 3. **Guest faculty – M. Sc (Biochemistry), PhD**					
Course Summary & Justification	The course is designed to get a clear idea on the basic biomolecules and their importance in the various biochemical processes in life so that the course builds a base for the students to comprehend and articulate the advanced concepts in life sciences.					
Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg. Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding of chemical groups and bonding; basics of cell biology and physiology					

O No.	Expected Course Outcome	Learning Domains	PSO No.
1	To identify the different types of biomolecules such as lipids, carbohydrates, proteins and nucleic acids	U	1

2	To differentiate the structural and functional characters of different biomolecules	A	2
3	To narrate the coordinated functions of different biomolecules in a complex living system	A/Ap	1
4	To compare the structure and functions of biomolecules in plants, animals and microbes	A	2
5	To describe the structure and functions of vitamins and hormones	U	2
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hours
1	**Carbohydrates:** Classification of Carbohydrates with examples- monosaccharides, disaccharides and oligosaccharides; their structure and functions; Polysaccharides - occurrence, structure, isolation, properties and functions of homoglycans- starch, glycogen, cellulose, dextrin, inulin, chitins, xylans, arabinans, galactans. Occurrence, structure, properties, and functions of heteroglycans – bacterial cell wall polysaccharides, glycoaminoglycans, agar, alginic acid, pectins, amino sugars and deoxy sugars, blood group substances and sialic acids. Glycolipids and Glycoproteins and their biological applications. Lectin- structure and functions.	1,2,3,4	10
2	**Lipids:** Classification of lipids with examples; their structure and functions Complex lipids- phospholipids -classification, structure and functions. Ceramides and sphingomyelins. Eicosanoids, structure and functions of prostaglandins, thromboxanes, leukotrienes Types and functions of plasma lipoproteins. Amphipathic lipids -membranes, micelles, emulsions and liposomes. Steroids -cholesterol structure and biological role - bile acids, bile salts. Sterols in Plant system: Phytohormones: Brassinosterroids (functions); Sterols in microbial system: mycosterols.	1,2,3,4	10
3.	**Proteins:** Amino acids- Structure amd properties**,** Classification of proteins on the basis of solubility and shape, structure, and biological functions. Isolation, fractionation and purification of proteins. Denaturation and renaturation of proteins. Primary structure -determination of amino acid sequence of proteins. Ramachandran plot, Secondary, tertiary and quartenary structures of proteins. Detailed study on structure and function with an example: Fibrous Protein (Collagen) Globular protein (Hemoglobin)., Enzymes- Different classes and functions.	1,2,3,4	10

4	**Nucleic Acids: Components of nucleic acids,** Watson -Crick model of DNA structure. A, B and Z DNA Cruciform structure in DNA, miscellaneous alternative conformation of DNA. Higher order organization of DNA. Methods for nucleic acid sequence determination, isolation and purification of DNA, molecular hybridization, Cot value curve, Reassociation kinetics, RNA Structure: Types of RNA; structure of mRNA, tRNA and rRNA ,Si RNA, micro RNA with emphasis on importance of structure to its function	1,2,3,4	10
5	**Vitamins and Hormones:** Vitamins -water soluble -thiamine, riboflavin, niacin, pyridoxine, folic acid, ascorbic acid-source, structure, biochemical functions, deficiency diseases, daily requirements; fat soluble -vitamin A, vitamin D2, vitamin E and vitamin K -sources, structure, biochemical functions, deficiency diseases, daily requirements. Hormones: different types, structures, their biological role and disorders. Mechanism of action of peptide and steroid hormones.	5	20
	Total Credits of the Course	3	

Books for Reference

Compulsory Reading:

1. Principles Of Biochemistry, 4/e (2006) by Robert Horton H , Laurence A Moran, Gray Scrimgeour K **Publisher:** Pearsarson **ISBN:** 0131977369, **ISBN-13:**9780131977365, 978-0131977365
2. Biochemistry 6th Edition (2007) by Jeremy M.berg John L.tymoczko Lubert Stryer **Publisher:** B.i.publicationsPvt.Ltd **ISBN:**071676766X **ISBN-13:** 9780716767664, 978-716767664
3. Lehninger Principles of Biochemistry, Fourth Edition by David L. Nelson Michael M. Cox Publisher: W. H. Freeman; Fourth Edition edition (April 23, 2004) ISBN-10: 0716743396 ISBN-13: 978-0716743392

Further Reading:

- Biochemistry: A Students survival Guide by Hiram. F. Gilbert (2002) Publishers: McGraw-Hill ISBN 0-07-135657-6
- Introduction to Biophysics by Pranab Kumar Banerjee (2008) Publishers: S. Chand & Company ltd ISBN: 81-219-3016-2
- **E.S. West**, **W.R. Todd**, **H.S. Mason and J.T. van Bruggen**, A **Text Book of Biochemistry**, Oxford and IBH Publishing Co., New Delhi, 1974
- Biochemistry [with Cdrom] (2004) by Donald Voet, Judith G. Voet **Publisher:** John Wiley & Sons Inc **ISBN:** 047119350X **ISBN-13:** 9780471193500, 978-0471193500
- Principles Of Biochemistry (1995) by Geoffrey L Zubay, William W Parson, Dennis E Vance **Publisher:** Mcgraw-hill Book Company – Koga **ISBN:**0697142752 **ISBN-13:** 9780697142757, 978-0697142757
- Molecular Biology of the Cell by Bruce Alberts, Alexander Johnson, Julian Lewis, Martin Raff, Keith Roberts, Peter Walter Publisher: Garland Science; 5 edition ISBN-10: 0815341059 ISBN-13: 978-0815341055
- Genes IX by Benjamin Lewin (2008) Publisher: J&b ISBN:0763752223 ISBN-13:

9780763752224, 978-0763752224

- Molecular Biology Of The Gene 5/e (s) by James D Watson, Tania A Baker, Stephen P Bell (2008) Publisher: Dorling Kindersley (India) Pvt Ltd ISBN: 8177581813 ISBN-13: 9788177581812, 978-8177581812
- Cell and Molecular Biology, 3e (2003) by Karp Publisher: Jw ISBN: 0471268909 ISBN-13: 9780471268901, 978-0471268901

Molecular Cell Biology (2002) by H.S. Bhamrah Publisher: Anmol Publications ISBN: 8126111429 ISBN-13: 9788126111428, 978-8126111428

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** **A. Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks **B. Semester End examination – 60 marks**

Approval Date	
Version	
Approval by	
Implementation Date	

MAHATMA GANDHI UNIVERSITY

BSM 21C 02: MICROBIOLOGY

SchoolName	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**MICROBIOLOGY**					
Type of Course	Core					
Course Code	**BSM 21C 02**					
Names of Academic Staff & Qualifications	**Dr.JISHA.M.S**					
Course Summary & Justification	This course on Microbiology introduces the milestones of Microbiology key components and their functions. The objective of the course content is to impart Knowledge on Landmark discoveries in Microbiology and different domains classification of living organisms. To develop a very good understanding of the characteristics of different types of microorganisms, methods to organize/classify these into and basic tools to study these in the laboratory.					
Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of General microbiology**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Summarize the contributions made by prominent scientists in microbiology and bacterial taxonomy	E	1
2	Understanding of basic microbial structure and similarities and differences among various groups of microorganisms	U/ An	2
3	Exemplify basic tools to study these in the laboratory	S	2
4	Explain various factors affecting the microbial growth and nutritional requirements and will be acquainted with methods of measuring microbial growth	U/R	2
5	Analyse various methods for identification and sterilization of isolated microorganisms.	An	1
6	Create an insight to the interactions and characteristics of microorganisms	An/ C	2
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**History and scope of microbiology:** The historical foundations and development of microbiology. An overview of microbial world. The bacteria and the archae. Classification of bacteria and Archaea according to the Bergey's Manual of Systematic Bacteriology. Tools for Systematics: Numerical taxonomy, Phylogenetic analysis, Polyphasic approach; Modern methods of studying microbial diversity; Microbial culture collections.	1,2	10
2	**Microbial Diversity:** Prokaryotic and eukaryotic microbial diversity.General characteristics of various groups of prokaryotes: bacteria including, Rickettsiae, Chlamydiae and Actinomycetes, Cyanobacteria and Mycoplasmas. Morphology and structure of bacteria. Viruses unique properties, morphology, structure and cultivation; Viroids and Prions.Viral replication.Viral diversity– bacterial, plant and animal viruses; Fungi - properties and classification. Microorganism in extreme environments..	1,2,3	20
3.	**Microbial physiology:** Factors influencing microbial growth. Environmental and nutritional factors. Nutritional types of bacteria. Microbial growth curve. Mathematical expression of growth-continuous and batch cultures. Diauxic and synchronous growth. Measurement of bacterial growth.Cultivation of bacteria- culture media and methods. Aerobic and Anaerobic culture methods. Culture preservation techniques. Microbial locomotion – flagellar motility, gliding motility and amoeboid motion. Chemotaxis,	4	20

	Phototaxis and other taxes. Microbial photosynthesis.		
4	**Identification of bacteria and Sterilization methods:** Identification of bacteria. Staining reactions. Cultural, physiological and biochemical properties. Molecular methods for identification. Sterilisation – Principles and methods, physical and chemical methods. Disinfectants – modes of action. Testing of disinfectants. Antibiotics – mechanism of action. Drug resistance in bacteria. Antibiotic sensitivity tests	5	10
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction: Active co-operative learning, Seminar, Group Assignments Authentic learning: Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** C. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks D. **Semester End examination – 60 marks**

References

Compulsory Reading:

1. Prescott, L. M., Harley, J. P. and Klein, D. A.2014. *Microbiology*. 9th Edition. Edition, McGraw Hill Higher Education.
2. Pelczar, M. J. Jr., Chan, E. C. S. and Krieg, N. R. 1993. *Microbiology*, 5th Edition, Tata MacGraw Hill Press.

Further Reading:

1. Jeffrey C.Pommerville.2016.Alcamos fundamentals of microbiology. Tenth Edition. Jones and Bartlett Learning.
2. Tortora G. J., Funke B. R. and Case C. L. 2015. *Microbiology: An Introduction*. 12th Edition. Pearson Education Inc.
3. Madigan, M. T. and Martinko, J. M. 2015. *Brock's Biology of Microorganisms*. 14th Edition. Pearson Education Inc.
4. .Willey, J. M., Sherwood, L. M. and Woolverton, C. J. 2013. *Prescott's Microbiology*.

8th Edition, McGraw-Hill Higher Education.

5. Stanier, R. Y., Adelberg, E. A. and Ingraham, J. L. 1987. *General Microbiology*, 5th Edition. Macmillan Press Ltd.

6. Russell, A. D., Hugo, W. B., and Ayliffe, G. A. J. 2013. *Principles and practice of disinfection, preservation and sterilization*, 5thEdition. Blackwell Science, Oxford.

7. Black, J. G. 2013. *Microbiology: Principles and Explorations*. 6th Edition, John Wiley and Sons, Inc.

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MAHATMA GANDHI UNIVERSITY

BSM 21C 03: CELL BIOLOGY, GENETICS & EVOLUTION

SchoolName	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**CELL BIOLOGY, GENETICS & EVOLUTION**					
Type of Course	Core					
Course Code	**BSM 21C 03**					
Names of Academic Staff & Qualifications	**Dr J G Ray, Dr Keerthi T R, Dr Jayachandran K, Dr. Linu Mathew**					
Course Summary & Justification	Th The course on Cell Biology and Genetics deals with the frontier areas of basic biology Th The objective of the course content is to create a sound awareness about the current developments taking place in different fields of cell biology and genetics The course content is designed with a view to augment CSIR/UGC syllabus					
Semester	First					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours

	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of cell biology and genetics**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**Build a perspective on current developments in the fields of cell biology, genetics and evolution and the cellular level organization of organisms**	E	1
2	**Compare and analyze the processes of cell cycle, cell division, cell differentiation and cell death and analyze the relationship between cell cycle, ageing, cell death and cancer**	U/ An	2
3	**Explain the processes, laws, and theories related to inheritance and evolution**	R	2
4	**Perform genetic mapping based on data supplied**	S	1
5	**Evaluate the behavior of genotypes and alleles in natural populations**	E	2
6	**Communicate effectively about a given topic in cellbiology/ genetics/ evolution both verbally and in writing**	An/ C	1
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**Cell and its constituents: Cell constituents -** Mitochondria,Chloroplast, Endoplasmic Reticulum Golgi complex, Peroxisomes, Lysosome, Ribosome, Nucleus, Nucleolus, Chromosomes, Nucleosomes, Histones, Genome, Genomics, Proteomics. **Cell cycle and Cancer: Cell cycle-** Different stages, variations, checkpoints, regulations of cell cycle, maturation Promoting factor, cells, cyclins , ubiquitin, protein ligases, Anaphase Promoting complex, inhibitors of CdK, growth factors and D cyclins. Rb protein and E2F transcription factors. **Cancer -** Stages in cancer development, causes, properties of cancerous cells, tumor Viruses, oncogenes, functions of oncogene products, oncogene and signal Transduction , oncogene and G	1,2,6	10

	proteins, oncogene and cell survival, Tumor Suppressor gene, functions of tumor suppressor gene products, Diagnosis,prevention and treatment of cancer		
2	**Cell Differentiation**-Stages of development, regulation of development, cascade control/ Differentiation in Drosophila, maternal, Segmentation and homeotic Genes, Genetic control of embryonic development, Bi thorax mutant, Antennapediac mutant ,Hemeobox **Aging** Process of aging, theories of aging, Arking's contribution Oxidative stress, Telomere problem, DNA repair defects. **Cell Death** Necrosis and Apoptosis, Differences between necrosis and Apoptosis, stages in Apoptosis, mitochondrial damage DNA ladders, transglutaminase activity, programmed cell death in *Ceanorhabdtis elegans* CED 3, CED 4, CED 9 and their roles in Apoptosis Bax, Bid, Bcl2 protein	1,2,6	10
3.	**Classical Genetics:** Genetics, the evolution of the subject through pre mendelian, Mendelian and post Mendelian Peroids. Mendelism – the basis principles of inheritance, gene interactions – allelic and no allelic. Environment and gene expression, penetrance and expressivity. Multiple alleles and polygenic inheritance, Heritability and genetic advance **Evolution:** Origin of the universe and origin of life; concept of Oparin, Miller-Urey Experiments; Evolution of Prokaryotes - origin of eukaryotic cells - Margulis Endosymbiotic theory; **Geological Timescale:** Tools and techniques in estimating evolutionary time scale; **Theories of evolution of life**: Pre-Darwinian concepts – Lamarkism, Darwinism – major concepts - variation, adaptation, struggle, fitness and natural selection, Neo-Darwinian theories – theories of speciation – allopatric and sympatric speciation - Rose Mary and Peter Grant (Molecular evolution in Darwinian finches) - Neutral Theory of Molecular Evolution.	3,6	20
4	**Chromasome genetic mapping ,Organelle Genetics and Population Genetics:** Linkage and linked genes with special reference to inheritance, Chromosome mapping with three - point test crosses. Organelle Genetics and cytoplasmic inheritance. Population Genetics – types of gene variations, Measuring genetic variations, Hardy Weinberg principle and its deviations. Medical genetics - an introduction	4,5,6	10
5	**Genetic System in Microbe, Yeast and Neurospora:** Plasmids & bacterial sex. Types of plasmids. Plasmids copy number and incompatibility, Replication of plasmid. Plasmid a cloning vector. Episomes. Transposable element-IS element and transposon, Integrons and Antibiotic resistance cassettes, Multiple antibiotic resistant bacteria, Mu-virus. Gene mapping in Bacteria. Bacteriophage genetics-Plaque formation & phage mutants, genetic recombination in lytic cycle. Genetic system in Yeast & Neurospora.	4,5,6	10
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Jonathan B (2016) Principles of Evolution, Garland Science, Taylor and Francis
2. Strickberger M W (2015) Genetics 3rd Edition, Pearson
3. Genetics a conceptual approach. 6th edition. Benjamin Pierce, Macmillan Learning, New York
4. The Cell-A Molecular approach, Fifth edition, Geoffrey M Cooper and Robert E .Hausman. , ASM Press ,Washington DC

Further Reading:

1. Principles of Genetics, Snustad, Simmons and Jenkins, John Wiley And Sons Inc
2. Genetics, Robert Weaver and Philip Hendricks, WH.C. Brown Publishers, Iowa
3. Introduction to Genetic Analysis, Griffiths, Wessler, Lewontin, Gelbart,Suzuki and Miller, Freeman's and Co, New York
4. REA's Problem Solvers in Genetics, Research Education Association,61, Ethel Roadwest, New Jersey
5. Cell and Molecular Biology by Gerald Karp,7th Edition,
6. Cell and Molecular Biology by De Robertis E.D.P, 8th Edition

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BSM 21C 04: BIOPHYSICS AND BIOSTATISTICS

SchoolName	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**BIOPHYSICS AND BIOSTATISTICS**					
Type of Course	Core					
Course Code	**BSM 21C 04**					
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S**					
Course Summary & Justification	This course is to introduce interdisciplinary Biophysics area, its scope and its importance The objective of the course is to give an insight into the basic concepts of thermodynamics, importance of basic biophysical phenomena, conformation and conformational changes, interaction of protein with other molecules and basic knowledge about radiation, its interaction with matter and its applications. The course content is to familiarize the basic concepts of biostatistics and its importance in research area of Life sciences The course content is designed with a view to augment CSIR/UGC syllabus					
Semester	First					
Total StudentLearning	Learning Approach	Lecture	Tutorial	Practical	Others	Total

Time (SLT)						Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of** Biophysics and Biostatistics					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Explain the scope and importance of biophysics	E	1
2	Describe the concepts of thermodynamics and applications of basic biophysical phenomena.	U/ An	2
3	Narrate the conformation and interaction of proteins and nucleic acids	R	2
4	Explain the electromagnetic radiation, its interaction with matter and applications.	S	1
5	Perform the retrieval of biological information by using structural and sequence databases	E	2
6	Explain the basic concept of biostatistics and analyze, interpret statistical softwares and to do statistical design for their research	An/ C	2
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**Biophysical phenomena and Thermodynamics of biomolecular interactions**: Scope and definition of Biophysics, Principle and biological importance of Osmosis, Electroosmosis, osmotic pressure, osmotic equilibrium, Donnan equilibrium, Diffusion, Sedimentation, Filtration, Surface tension, Dialysis, Adsorption and Colloids. Laws of thermodynamics, Enthalpy, Entropy, Free energy, Redox reactions, Redox potential and its calculation by Nernst equation, examples of redox reactions in biological system.	2	10

2	**Structural Biophysics and computational biology**: The molecular interactions between proteins and nucleic acids: DNA- protein interaction and RNA- protein interactions, DNA-binding motifs: Helix-turn-Helix motif, Zn fingers, Helix-loop helix motifs and Leucine zippers. Molecular forces: Hydrogen bonding, hydrophobic interactions, Dipole interactions: charge-dipole interactions, induced dipoles, steric repulsion, Vander waals force in biomolecules, Structural and Sequence databases, Alignment algorithms; Retrieval of biological information from widely used resources: NCBI and PDB, Molecular modelling and Structure based drug designing.	1,3,5	10
3.	**Radiation Biophysics**: Electromagnetic spectrum, Ionizing and non ionizing radiation. Properties and biological effects of ultraviolet radiation, infrared and microwave radiations. Radioactivity, Interaction of radiation with matter. Units of Radiation. Biological effects of radiation. Applications of ionizing and non-ionising radiations in industry, agriculture and research. Radiation hazards.	1,4	20
4	**Introduction to Biostatistics:** Scope of Biostatistics, probability and probability distribution analysis. Variables in biology- collection, classification and tabulation of data- graphical and diagrammatic representation- scatter diagrams, histograms- frequency polygon- frequency curve-logarithmic curves. Descriptive statistics- measures of central tendency, Arithmetic mean, median, mode, geometric mean, harmonic mean. Measures of dispersion, standard deviation, standard error, variance, coefficient of variation. Correlation and Regression	5	10
5	**Test of significance:** Basic idea of significance test- hypothesis testing, levels of significance. Testing of single mean, double mean, single proportion, double proportion in large sample. Testing of single mean, double mean and Paired- t in small sample. ANOVA- One way and Two way; Chi-square test of goodness of fit and Chi-square test of independence, comparison of means of two samples, three or more samples. Fundamentals of field experiments- randomization, replication and local control. CRD and RBD. Statistical packages	5	10
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative

Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Proteins, Structure and molecular properties, Thomas E Creighton
2. Fundamentals of Biostatistics: Irfan.A. khan, Atiya Khanum, Ukaaz publications
3. Principles of Biostatistics: Marcello Pagano, Kimberlee Gauvreau, Duxbury Press
4. Biochemistry: Donald Voet and Judith G Voet, Wiley Publications

Further Reading:

5. Biophysics-Hoope W etal
6. Biophysics-Volkenstain M.V
7. Molecular Biophysics- Volkenstain M.V
8. Introduction to thermodynamics of irreversible process-John Wiley
9. Statistical methods in Biology- Briley N.J.T
10. Introduction to Biophysics-Sokal R.R & Rohl F.J
11. Biostatistics: Pardeep.K.Jasra, Gurdeep Raj, Krishna prakashan Media.(P) Ltd
12. Bloomfield, V. (2009) Computer Simulation and Data Analysis in Molecular Biology and Biophysics. Springer

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MAHATMA GANDHI UNIVERSITY

BSM 21C 05: PHYSIOLOGY

SchoolName	**School of Biosciences**					
Programme	**MSc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**PHYSIOLOGY**					
Type of Course	Core					
Course Code	**BSM 21C 05**					
Names of Academic Staff & Qualifications	**Dr. R. Harikumaran Nair** MSc, PhD					
Course Summary & Justification	This course is designed to provide an overview of human physiology. Course topics will include the various systems of the body, functions of each system, and interrelationships to maintain the internal environment. The course also provides inputs to physiological stress and adaptive strategies to overcome stress					
Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours

	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics Knowledge in Biology**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Students should be capable of effectively communicating how the human body works	U	1
2	Students should be able to explain interrelationships among molecular, cellular, tissue, and organ functions in each system	E	2
3	Students should be able to describe the interdependency and interactions of the systems	A	2
4	Students should be able to explain contributions of organs and systems to the maintenance of homeostasis	A	1
5	Students should be able to identify causes and effects of homeostatic imbalances	E	2
6	Able to gain the approaches used to study various functional systems of the human body and physiologic adaptation	I	2
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	The system as a basic unit in physiology: different systems in physiological process, interaction of different systems in normal and stress conditions, homeostasis, Neuro-Musculo-Skeletal systems:	1	10

	brain and peripheral nervous systems, neurotransmitters, synapse, neuro-muscular junction, musculoskeletal systems		
2	Cardio-Pulmonary & Renal Physiology: Anatomy and general function of heart, blood and hemodynamic, blood pressure, heart rate, cardiac cycle, cardiac output, electrocardiography, echocardiography; anatomy of the respiratory system, principles of respiratory mechanisms, respiratory rate, lung volumes, oxygen uptake, lung function tests, gas transport; anatomy of the excretory system, nephron, glomerular filtration rate, urine formation, renal clearance test, renal regulation of electrolytes, dialysis	1,2,3	20
3.	Principles of endocrinology: Role of hormones for maintenance of the internal environment, hormone transport in blood, mechanism of hormone action, hormone metabolism and excretion, types of endocrine disorders, hypothalamus and pituitary, thyroid, adrenal glands, endocrine control of growth, sex hormones, pancreatic hormones, neurohormones	1,4,5	10
4	Gastrointestinal Physiology & Nutrition: Gastrointestinal structure, food digestion, and absorption, gastrointestinal hormones, central control of gastrointestinal functions, pathological situations of gastrointestinal functions. role of liver and bile in gastrointestinal functions.	3,6	10
5	Stress physiology: Stress-responses, the role of the hypothalamic-hypophyseal-adrenal axis, oxidative stress and mechanism, effect of stress-inducing and anti-stress agents, cardio-respiratory responses during high altitude acclimatization, stress-induced diseases, and remedy, Human tolerances to stresses in space including space flight: Physiological adaptation to space flight, physiology in deep-sea diving and other high-pressure operations	6	10
	Total Credits of the Course	3	60

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading

1. Vander's Human Physiology- The mechanism of body function. Widmaier, Raff & Strang
2. Textbook of Medical Physiology. Arthur.C. Guyton& John.E. Hall
3. Physiological basis of Medical Practice. John.B. West
4. Endocrinology- Mac E Hadley

Further Reading:

1. Review of Medical Physiology- Ganong, William F
2. Biochemistry and Physiology of the cell. An introductory text second edition- Edwards, N. A Hassall, K.A
3. Notebook of medical physiology: endocrinology, with aspects of maternal, fetal and neonatal physiology- Hawker, Ross Wilson
4. Human Physiology: an integrated approach- Silverthorn, Dee Unglaub
5. Principles of anatomy and physiology- Tortora, Gerald J Derrickson, Bryan
6. Textbook of Endocrine Physiology- Griffin, James E; Ed. Ojeda, Sergio R;Ed

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	MAHATMA GANDHI UNIVERSITY
	BSM 21C 06: LABORATORY COURSE 1BIOCHEMISTRY

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**LABORATORY COURSE 1: (GENERAL BIOCHEMISTRY)**					
Type of Course	Core					
Course Code	**BSM 21C 06**					
Names of Academic Staff & Qualifications	1. **Dr. Anie Y – M. Sc (Biochemistry), PhD**					
Course Summary & Justification	The course is designed to develop in students the essential skills to perform the basic biochemical assays, qualitative analysis of biomolecules and techniques for the separation of biomolecules. This will enhance the practical abilities of the students to carry out the analysis of biomolecules.					
Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg. Authentic learning Collaborative learning Independent learning	10	20	120	30	240
Pre-requisite	General idea on reagents and solvents					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To prepare reagents, buffers and other solutions in required concentrations and required pH.	Ap	1
2	To extract and estimate different bio-molecules (sugar, cholesterol, and proteins) in biological samples	Ap/S	2
3	To identify the different components in a mixture of carbohydrates	S	2
4	To detect the presence of albumin, casein and gelatin in biological samples	S	1
5	To perform separation by Paper and Thin layer chromatography	S	1
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hours
1	Preparation of solutions: Percentage solutions, Molar solutions, Normal solutions, Dilution of Stock solutions, Preparation of buffers using the Henderson Hasselbach equation	1	15
2	Spectrophotometric experiments: Verification of Beer Lambert's law, Determination of UV-Visible spectrum of compounds, Determination of Concentration of molecules from Molar Extinction Coefficient values Extraction of Polysaccharides (Starch/Glycogen), Proteins, and Lipids from appropriate sources and their estimations. Estimations: Estimation of reducing sugars by Dinitrosalicylic acid method, Estimation of proteins (Biuret and Lowry's methods), Estimation of Methionine by Nitroprusside method, Estimation of Cholesterol by Zak's method.	2	45
3.	Qualitative analysis of Carbohydrate mixtures (a combination of polysaccharide, disaccharide and monosaccharide) following systematic scheme for analysis. (Starch, dextrin, glycogen, glucose, fructose, xylose, galactose, sucrose, maltose, lactose) Qualitative analysis of proteins- Albumin, casein, gelatin	3,4	45
4	Chromatographic techniques: Separation of amino acids by Paper chromatography (Descending or Ascending), Separation of Plant pigments by Thin layer chromatography	5	15
	Total Credits of the Course	3	
Books for Reference			

Compulsory Reading: **1.** Introductory Practical biochemistry, S. K. Sawhney & Randhir Singh (eds) Narosa Publishing House, New Delhi, ISBN 81-7319-302-9, p 195 – 303 **2.** Standard Methods of Biochemical Analysis, S. K. Thimmaiah (ed), Kalyani Publishers, Ludhiana ISBN 81-7663-067-5, p 12 - 182.
Further Reading: **3.** Hawk's Physiological Chemistry, Bernard L. Oser (ed) TATA McGRAW Hill Publishing Company LTD, New Delhi, p 60 – 127, 1317- 1334 **4.** Experimental Biochemistry: A Student Companion, Beedu Sasidhar Rao & Vijay Deshpande (ed), I.K International Pvt. LTD, New Delhi ISBN 81-88237-41-8, p 13- 17, p 49 - 72 **5.** Practical Biochemistry, R.C. Gupta & S. Bhargava (eds) CBS Publishers and Distributors, New Delhi, ISBN 81-239-0124-0 p 9 – 27 **6.** Practical Clinical Chemistry, Harold Varley, CBS Publishers and Distributors, New Delhi,

TeachingandLearningApproach	**Laboratory Procedure (Mode of transaction)** Direct Instruction: lecture, Explicit Teaching, Demonstration, Hands on experimental sections, Skill acquisition by laboratory training
Assessment Types	**Mode of Assessment** **C. Continuous Internal Assessment (CIA)** Assessment of the performance of student in the lab- 10 marks Internal Test -20 marks Project report (student needs to perform experiments on a specific project and report should be prepared)– 10 marks **D. Semester End examination – 60 marks**

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	**MAHATMA GANDHI UNIVERSITY** **BSM 21C 07: LABORATORY COURSE-2-PHYSIOLOGY**

SchoolName	**School of Biosciences**					
Programme	**MSc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**LABORATORY COURSE-2-(PHYSIOLOGY)**					
Type of Course	Core					
Course Code	BSM 21C 07					
Names of Academic Staff & Qualifications	**Dr. R. Harikumaran Nair** MSc, PhD					
Course Summary & Justification	The purpose of this laboratory course is to provide the student with the opportunity to observe many physiological principles. The course is designed to understand the mechanisms related to cardiovascular and respiratory functions.					
Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours

	Authentic learning Collaborative learning Independent learning	5	5	120		130
Pre-requisite	**Basics Knowledge in Biology**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Apply appropriate safety standards in laboratory	A	2
2	Acquire laboratory skills in haematology, cardiovascular and respiratory physiology	S	1
3	Appropriately utilize laboratory equipment, such as microscopes, dissection tools, general labware, physiology data acquisition systems	S	1
4	Communicate results of scientific investigations, analyse data, and formulate conclusions	C	2
5	Students should be able to identify cell structure	U	2
6	Work collaboratively to perform experiments	I	2
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**Haematology** i) Determination of haemoglobin concentration ii) Enumeration of formed elements- red blood cells & white blood cells iii) Study of blood smear for the differential count and cell morphology iv) Erythrocyte sedimentation rate v) Determination of the bleeding time vi) Determination of clotting time	1,2,5,6	60
2	**Respiratory physiology- Pulmonary function testing** i) Demonstration on the recording of tidal volume	1,3,6	30

	ii) Demonstration on the recording of vital capacities iii) Demonstration on the recording of inspiratory & expiratory flow rates		
3.	**Cardiovascular physiology- Electrocardiography** i) Demonstration on ECG recording- human or animal model ii) Identification of ECG waves iii) Calculation of heart rate from ECG	1,3,4,6	30
	Total Credits of the Course	2	120

Teachingand LearningApproach	**Laboratory Procedure (Mode of transaction)** Direct Instruction: lecture, Explicit Teaching, Demonstration, Hands on experimental sections, Skill acquisition by laboratory training
Assessment Types	**Mode of Assessment** E. Continuous Internal Assessment (CIA) 1. Internal Laboratory Skill Tests of maximum 20 marks 2. Seminar Presentation – Laboratory material and methods Maximum marks 10 3. Write a detailed report on instrumentation – 10 marks F. **Semester End Practical examination – 60 marks**

REFERENCES

1. Medical Laboratory Technology-A Procedure Manual for Routine Diagnostic Tests- Kanai L Mukherjee
2. Pocket Guide to Spirometry- David P Johns and Rob Pierce
3. Spirometry in Practice- A practical guide to using spirometry in primary care- Dr. David Bellamy, British Thoracic Society COPD consortium.
4. ECGs made easy- Barbara J Aehlert

Further Reading:

1.ECG Assessment and Interpretation- Cascio, Toni
2.Introduction to medical laboratory technology- Baker, F J Silverton, R E
3.Practical haematology- Dacie, John V Lewis, S.M

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MAHATMA GANDHI UNIVERSITY

SBSNCC 1 Entry level orientation programme in applied life sciences

SchoolName	School of Biosciences					
Programme	M.Sc. Biochemistry, Biotechnology, Microbiology, Biophysics					
Course Name	Entry level orientation programme in applied life sciences					
Type of Course	Noncredit course					
Course Code	**SBSNCC 1**					
Course Summary & Justification	The proposed course is offered as a noncredit mandatory course at the entry level for all the PG students of school of Biosciences. The course content is inclusive of the scope and opportunities in various branches of applied life sciences along with suitable discussion on the preliminary aspects of lab training. It gives an orientation to the students coming from different disciplines of life science graduation and brings them to a common platform for further learning. This is a two week long bridging course					
Semester	First					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60		0		60

Pre-requisite	Fundamental Knowledge in Life Sciences

COURSEOUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1.	The students from various branches of life sciences are brought to a common platform	R/U	1
2.	The students will be getting a clear understanding of the different opportunities in their subject	R/U	1
3.	The course focusses on the requirement of awareness on good laboratory practices	U/ An	1
4.	The proposed entry stage training offers a good exposure to the field of research	U/An	1
5.	The students will be exposed to certain preliminary requirements for initiating startups, getting into QC,R&D	C/S	1
6.	The students on completing this entry stage course will be able to get along with the two-year course with a defined objective	A/S	1

**Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**Scope of the subject** Introducing the subject of Biochemistry, Biotechnology, Microbiology, Biophysics. Importance and recent trends, Opportunities. Method of teaching, learning and evaluation. Outcome based Education, Credit and semester system.	1	10
2	**Good laboratory practices** Laboratory instructions , Handling of Chemicals, Basics of weights and measureents, handling of equipment, Lab procedure, keeping of Lab record, Personal qualities and scientific conduct.	3	20

3.	**Basic Chemistry for Lab Work** Preparation of solutions, Methods for expressing concentration of solution, Colligative properties, Normality, Molarity, Molality, Mole fraction. pH, Buffering system, Examples Henderson Hasselbalch Equation.	5	10
4	**Research opportunities** Introduction to research, research aptitude, experimental design and research conduct, research problems, recent trends, Concept of research paper and review writing, plagiarism, Grammar editing softwares Regulatory bodies in life sciences, Patents and patent rules, Ethical Concepts-Research ethics, Bioethics. CSIR, UGC, GATE, DBT, DST, ICMR, ICAR, KSCSTE, fellowships, Projects, Opportunities.	1,2,4	10
5	**Job opportunities** Introduction to Entrepreneurial process and types of Business, opportunities, Startups, Basics of marketing, Quality control and management, R and D management, Innovation and knowledge management, Knowledge economy, Upskilling, Project preparation, team building,	1,5,6	10
	Total Credits		

Teaching And Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction, Active co-operative learning, Seminar, Group Assignments Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) B. Write a detailed report on a given topic based on research findings and literature search (Graded as very good, satisfactory and not satisfactory)

REFERENCES

Compulsory Reading:

1.Principles and techniques of Biochemistry and Molecular biology, Andreas Hofmann and Samuel Clokie, Cambridge University Press, 8 th edition,2018

2.Holmes D ., Moody P and Dine D.(2010).Research methods for the Biosciences,2 nd Editions, Oxford University Press,Oxford, UK.

3.Smith D (2003).Five Principles for research ethics, Monitor on Psychology 34. 56.

Further Reading: **4.Taylor P.L.(2007).Research sharing, ethics and public benefit. Nature Biotechnology, 25,398-401.** **5.Duke C.S. and Porter J.H (2013).The ethics of data sharing and reuse in Biology, Bioscience 63,483-489.**

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SECOND SEMESTER

MAHATMA GANDHI UNIVERSITY

BSM 21C 08IMMUNOLOGY

SchoolName	School of Biosciences
Programme	M.Sc. Microbiology/Biotechnology/Biochemistry/Biophysics
Course Name	IMMUNOLOGY
Type of Course	Core
Course Code	**BSM 21C 08**
Names of Academic Staff & Qualifications	Dr.Radhakrishnan E.K. M.Sc., Ph.D
Course Summary & Justification	This course on Immunology deals with various mechanisms and processes involved in the defense responses. This course is an important branch of life science. Human body has different lines of defense to fight against pathogens. The content in this course has been designed with an objective to provide detailed understanding on the process and mechanisms involved in the defense responses. Understanding on the

	functioning of immune system is highly essential for a student to explore its theoretical and practical aspects for the benefit of society.					
Semester	Second					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding on defense responses Knowledge in any branch of Life science					

COURSEOUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1.	Students will able to understand and explain basic principles of immunology	R/U	1
2.	Students will able to learn the recent advances in immunology	R/U	1
3.	Students will able to analyse the clinical importance of immunological reactions	U/ An	1
4.	Students will become able to identify the correlation between immunological abnormalities and health status of humans	U/An	1
5.	Students will get theoretical and technical know-how for the laboratory diagnosis of infectious diseases	C/S	1
6.	Students can apply the knowledge and skills for clinical and diagnostic applications	A/S	1
**Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*			

COURSE CONTENT

Module No	**Module Content**	**CO**	**Hrs**
1	Infection, Source and methods of transmission, Immunity- Types of immunity. Mechanisms of innate immunity, PAMPs, pattern recognition receptors, types, scavenger receptors and toll – like receptors, Phagocytes and Phagocytosis, Organs and cells with immune functions. Lymphocytes and lymphocyte maturation. PAMPs and PRRs in plants	1	10
2	Antigens, Epitopes and paratopes, B-cell and T-cell epitope, Antigenicity and Immunogenicity, Antibodies, Immunoglobulin – structure, classes and functions. Genetic basis of antibody diversity, Organization and Expression of Immunoglobulin Genes, V(D)J rearrangements; recombination signal sequences and their	1,2	20

	role, somatic hypermutation and affinity maturation Antigen-antibody reactions, Agglutination, Precipitation, Immunoflourescence, Complement fixation, Radioimmuno assay, ELISA, Western blotting		
3.	Immune response- Humoral and cell mediated, Receptors on T and B cells for antigens, MHC, TCR- mediated signalling, Signal transduction pathways associated with T-cell activation, Signal transduction by activated B- cell receptor, Antibody production, Primary and secondary immune response, Factors influencing antibody production, Clonal selection theory, Monoclonal antibodies – production and application, Antibody engineering. Complement system, Complement activation, Biological effects of complements, Antigen processing and presentation, Activation of T-cells, T cell function, Cytokines. Human microbiome and immunity	2,3,4	10
4	Immunology of organ and tissue transplantation, Allograft reaction and GVH reaction, Factors influencing allograft survival, Immunology of malignancy, Tumor antigens, Immune response in malignancy, Immunotherapy of cancer, Immunohematology, ABO and Rh blood group system, Immunology of blood transfusion, Hemolytic disease of new born	2,4,5	10
5	Immunological Tolerance, Autoimmunity, Mechanisms of autoimmunization, Autoimmune diseases. Inflammation, Hypersensitivity – immediate and delayed reactions, Clinical types of hypersensitivity, Immunodeficiency diseases, Immunoprophylaxis, Vaccines –types of vaccines, DNA vaccine, recent trends in vaccine development.	2,6	10
	Total Credits	**3**	

Teaching And Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction, Active co-operative learning, Seminar, Group Assignments Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative

Assessment Types	**Mode of Assessment** C. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar - Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks D. Semester End examination – 60 marks

REFERENCES

Compulsory Reading:

1. Immunology - Thomas J. Kindt, Barbara A. Osborne, Richard A. Goldsby, and Janis Kuby, W H Freeman and Co., 2013
2. Immunobiology - Charles A. Janeway Jr.,Paul Travers, Mark Walport and Mark J. Shlomchik, Garland Publishing., 2016

Further Reading:

3. Essential Immunology - Ivan M. Roitt and Peter J delves, Blackwell Publishing, 2016
4. Essential Clinical Immunology – Helen Chappel and Mansel Haeney, ELBS/Blackwell Scientific Publications, 2014
5. Introduction to Immunology – John W, Kimball Maxwell, Mac Millan International Edition, 1990
6. Text book of Microbiology – R. Ananthanarayanan and C K Jayaram Panicker. Orient Longman,2013

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MAHATMA GANDHI UNIVERSITY

BSM 21C 09 MOLECULAR BIOLOGY AND GENETIC ENGINEERING

School Name	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**Molecular Biology and Genetic Engineering**					
Type of Course	Core					
Course Code	**BSM 21C 09**					
Names of Academic Staff & Qualifications	**Dr Keerthi T R, Dr Jayachandran K, Dr. Linu Mathew**					
Course Summary & Justification	1. Molecular Biology and Genetic Engineering is one of the most dynamic and attractive courses in all branches of applied life sciences 2. The syllabus content in this paper is designed with an objective to train the students in both theoretical and practical aspects of the subject 3. This will also enable the students to get an idea about the latest developments taking place in this subject					
Semester	Second					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basics of cell and molecular biology, Basics of tools and techniques of genetic engineering					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	On completing this course the students will be able to Explain the processes of replication, transcription and translation and analyse the importance of these processes in health and disease	E	1,4,5
2	Explain the concepts of gene regulation in prokaryotes and RNA world	R/ E	1,4,5
3	Analyse the use of different tools and techniques of gene cloning in E coli and explain the applications of DNA technology	U	3,4,5

4	Ability to develop a protocol for cloning a gene from a selected organism	A	3,4,5
5	Ability to explain verbally and orally the concepts of molecular biology and genetic engineering	E	4,5
6	Ability to write a research proposal based on the concepts discussed in the course	An/ C	4,5
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**DNA Rr Replication–** Process of DNA replication, Semiconservative, discontinuous uni and bidirectional , Okazaki fragments, DNA polymerases in eukaryotes and prokaryotes , Klenov fragment, modes of replication, theta, rolling circle, d-loop replication, Primasome, SSB, Helicase, Ligase, methylation and control, repetitive DNA sequences, minisatellite, microsatellite, DNA protein interation DNA Linking number and topoisomerase, Inhibition of replication.	1,5,6	10
2	**Transcription**. Process of transcription, stages in transcription, RNA polymerases in prokaryotes and enkaryotes, sigmafactor in prokaryotes, Rho dependant and Rho independent termination. Enhancers, Transcription factors in Eukaryotes, Differences in transcription between prokaryotes and Eukaryotes, post transcriptional modifications-Polyadenylation, capping, r-RNA processing, Splicing-Spliceosome, lariat structure, Group 1, II and III Introns Rihozyme, Importance of ribozyme, properties, application, RNase P, RNAse III, RNAse H. monocistonic and polysistronic m-RNA, Joint transcript of r-RNA and t-RNA in prokaryotes and their processing, Transplicing, alternate splicing, inhibitors of Transcription. Molecular mechanism of gene regulation in prokaryotes-Transcriptional regulation in prokaryotes; Inducible & repressible system,+ & -ve regulation; Operon concept, structure of operon, Lac, Trp, Arc operon, Catabolic repression, Atteunation. Role of Hormones in gene regulation. RNA World, RNA based technology- Molecular mechanism of Ribozyme, Antisense RNA, SiRNA, MicroRNA, Ribozwitches & their applications; Telomerase structure and function, Nucleic acid as therapeutic agent	1,2,5,6	15
3.	**Translation:** Process of translation. Stages in translation, genetic code, properties, wobble hypothesis, eukaryotes and prokaryotes ribosomes, m-RNAs, t-RNAs, aminoacyl t-RNA synthatases, protein factors initiation complex, peptidyl transferase, releasing factors, differences between prokaryotic and eukaryotic systems, inhibition of translation. Post translation modification by cleavage, self assembly assisted self assembly chaperones, acylation,	2,5,6	10

	phosphorylation, acetylation and glycosylation, Histone acetylation and deacetylases, chromosome remodeling complex. Intein splicing. Protein targeting, cotranslational import, post translational import, SRP- structure and function, Blobel's concept, Lysosome targeting, M6P address Glycosylation core glycosylation terminal glycosylation, Dolichol phosphate.		
4	**Tools and techniques for genetic Engineering:** History of rDNA Technology ,Cohen And Boyer Patents, Isolation of DNA and RNA from different sources, enzymes used in genetic engineering with special reference to restriction enzymes, ligases, and other DNA modifying enzymes. End modification of restriction fragments, vaccinia topoisomerases mediated ligation of DNA, TA cloning, and homopolymer tailing Vectors for E coli with special reference to plasmid vectors (pSC101, pBR322,pUC,their development, features and selection procedures),direct selection plasmid vectors, low copy number plasmid vectors, runaway plasmid vectors, Bacteriophages (λ and M13) with special reference to Charon phages, λEMBL, λWES λB', λ ZAP- their development, features, selection procedures, *in vitro* packaging mechanisms for phage vectors, cosmids, features, advantages and cosmid cloning schemes, phagemids with special reference to pEMBL, pBluescript, pGEM3Z , pSP64, pcDNA, pLITMUS Construction of genomic libraries and cDNA libraries, procedures for recombinant selection and library screening, PCR enzymes, types of PCR, primer design, real time PCR, RTPCR, Nested PCR, Inverse PCR, Assymmetric PCR, applications of PCR Cloning, Chemical synthesis of DNA, DNA sequencing:- plus and minus sequencing, Sangers dideoxy sequencing, Maxam and Gilberts method. Advanced sequencing procedures: – pyrosequencing, Illumina, ABI / SOLiD and their applications	3,4,5,6	20
5	**Appications of Genetic Engineering:** Applications of transgenic Technology Improving quality, quantity and storage life of fruits and vegetables. Plants with novel features, Engineering metabolic pathways, Pharming. Animal cloning, Ethics of cloning. Applications of Molecular Biology in forensic sciences, medical science, archeology and paleontology	3,4,5,6	5
	Total Credits of the Course	3	

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** G. Continuous Internal Assessment (CIA)

	1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks H. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Principles of gene manipulation – Old and Primrose, Blackwell Scientific publishers, Edn.5th
2. Cell and Molecular Biology by Cooper

Further Reading:

7. Principles of gene manipulation – Old and Primrose, Blackwell Scientific publishers, Edn.5th
8. Principles of gene manipulation – Old, Primrose, and Twyman, Blackwell Scientific publishers, Edn. 6th
9. Principles of gene manipulation – Old, Primrose, and Twyman Blackwell Scientific publishers, Edn 7th
10. Molecular biotechnology, Principles and Applications of Recombinant DNA, Glick Pasternak and Patten, 4th edition ISBN 978-1-55581-498-4 Wiley International Publishers
11. From gene to genomes – Concepts and applications of DNA technology Jeromy W Dale and Malcom von Shantz , John Wiley and sons
12. Principles of plant biotechnology: An introduction to genetic engineering in plants – SH Mantell
13. Cell and Molecular Biology by Gerald Karp, Academic Press
14. Cell Biology by DeRobertis
15. Genes-Benjamin Lewin

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MAHATMA GANDHI UNIVERSITY

BSM 21C 10: METABOLISM AND BIOENERGETICS

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**METABOLISM AND BIOENERGETICS**					
Type of Course	Core					
Course Code	**BSM 21C 10**					
Names of Academic Staff & Qualifications	2. **Prof. M S Latha – M. Sc (Biochemistry), PhD** 3. **Dr. Anie Y – M. Sc (Biochemistry), PhD** 4. **Guest faculty – M. Sc (Biochemistry), PhD**					
Course Summary & Justification	The course is designed to get a deep knowledge of metabolic processes taking place in the biological systems and their regulation, which is needed to understand the more specialised areas of Biochemistry.					
Semester	Second					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg. Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding of chemical groups and bonding; basics of cell biology and physiology					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To be able to categorize, differentiate and predict the fates of different biomolecules via the metabolic pathways.	U/A	1
2	To draw conclusions on the energetics of the metabolic pathways and to find out the variations in ATP generation during physiological and pathological	A	2

	conditions		
3	To analyse different methods of regulation of the metabolic pathways.	A/An	2
4	Describe the different steps involved and the importance of metabolomics in toxicity analysis and health management	A	2
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hours
1	**Metabolic Pathways:** Detailed study on the catabolic pathways & anabolic Pathways -Carbohydrate, Protein, Amino acid and Nucleic acid metabolic pathways.	1	20
2	**Bioenergetics:** Functional significance of the mitochondrial respiratory chain and oxidative phosphorylation, Electron transport chain: structural components of the chain, complexes, free elements; Structure and functional properties of cytochromes, ferro-sulphurated proteins and CoQ; Generation of the electrochemical proton gradient: Chemiosmosis ATP synthesis- Proton flow through ATP synthase, Rotational catalysis. Inhibitors and uncouplers	1,2	15
3.	**Regulation of metabolism:** Hormonal and Allosteric regulation of pathways in carbohydrate, lipid, nucleotide, amino acid and protein metabolism; Coordinated regulation of opposing metabolic pathways; Regulation of mitochondrial electron transport and oxidative phosphorylation.	3	10
4	**Signal Transduction:** intracellular receptor and cell surface receptors signaling: Cyclic AMP-dependent protein kinase; Cyclic GMP-dependent protein kinase; Protein kinase C; Ca^{2+} - calmodulin-dependent protein kinases ; AMP-dependent protein kinase ; Receptor tyrosine kinases; Protein kinase B; Cytokine activation of the JAK'/STAT pathway; Cell cycle control; Receptor serine/threonine kinases; Other protein kinases ; Phosphoprotein phosphatases; Cancer Pathways: MAPK, P13K, TP53 network, NF*k*B pathways; Signalling by TGF β factor , STAT factor	3	10
5	**Metabolomics:** Introduction to origins of metabolomics; define terms: Metabolite, Metabolome, Metabonomics; Analytical techniques in study of Metabolomics (Principle & Methodolgy): Separation methods: Gas Chromatography, HPLC, Capillary Electrophoresis; Detection Methods: Mass spectroscopy, NMR. Applications of Metabolomics in toxicity assessment/ toxicology, diagnostics and health Screening	4	5
	Total Credits of the Course	3	

Books for Reference
Compulsory Reading: 1. Principles Of Biochemistry, 4/e (2006) by Robert Horton H , Laurence A Moran, Gray Scrimgeour K **Publisher:** Pearsarson **ISBN:** 0131977369, **ISBN-13:**9780131977365, 978-0131977365 2. Biochemistry 6th Edition (2007) by Jeremy M.berg John L.tymoczko Lubert Stryer **Publisher:** B.i.publicationsPvt.Ltd **ISBN:**071676766X **ISBN-13:** 9780716767664, 978-716767664 3. Lehninger Principles of Biochemistry, Fourth Edition by David L. Nelson Michael M. Cox Publisher: W. H. Freeman; Fourth Edition edition (April 23, 2004) ISBN-10: 0716743396 ISBN-13: 978-0716743392
Further Reading: • **E.S. West, W.R. Todd, H.S. Mason and J.T. van Bruggen,** A**Text Book of Biochemistry**, Oxford and IBH Publishing Co., New Delhi, 1974 • Biochemistry [with Cdrom] (2004) by Donald Voet, Judith G. Voet **Publisher:** John Wiley & Sons Inc **ISBN:** 047119350X **ISBN-13:** 9780471193500, 978-0471193500 • Principles Of Biochemistry (1995) by Geoffrey L Zubay, William W Parson, Dennis E Vance **Publisher:** Mcgraw-hill Book Company – Koga **ISBN:**0697142752 **ISBN-13:** 9780697142757, 978-0697142757 • Biochemistry (2008) by Rastogi **Publisher:** Mcgraw Hill **ISBN:**0070527954 **ISBN-13:** 9780070527959, 978-0070527959

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** **I. Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks **J. Semester End examination – 60 marks**

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MAHATMA GANDHI UNIVERSITY

BSM 21C11 BIOPHYSICAL TECHNIQUES AND BIOINSTRUMENTATION

SchoolName	**School of Biosciences**					
Programme	**MSc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**BIOPHYSICAL TECHNIQUES AND BIOINSTRUMENTATION**					
Type of Course	Core					
Course Code	**BSM 21C 11**					
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S**					
Course Summary & Justification	This course is designed to introduce different techniques used in life sciences This course gives knowledge of the principle of operation and design of scientific instruments It attempts to render a broad and modern account of scientific instruments					
Semester	Second					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of** Biophysics and Biostatistics					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To explain the methods used for gaining information about biological systems on an atomic or molecular level.	E	1
2	To describe different spectroscopic techniques	U/ An	2

3	To perform various biophysical fractionation and separation of biomolecules	R	2
4	To describe how to perform electrophoretic techniques	S	2
5	To describe the procedures and applications of hydrodynamic techniques	E	1
6	To perform different microscopic techniques	An/ C	1
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**Spectroscopic techniques**: Basic principles, nature of electromagnetic radiation, Interaction of light with matter, Absorption and emission of radiation; Atomic & Molecular Energy levels, Electronic, vibrational and Rotational spectroscopy of molecules, transition and selection rules; Atomic & Molecular spectra. Principle, Instrument Design, Methods & Applications of UV-Visible spectroscopy, Infrared spectroscopy, Raman Spectroscopy, Fluorescence spectroscopy, Nuclear magnetic Resonance Spectroscopy.	1,2	10
2	**Physicochemical Fractionation techniques**: Principle, Instrument Design, methods and Applications of all types of Adsorption and Partition Chromatography- Paper chromatography, Thin layer chromatography, High Performance Thin layer Chromatography, Gel filtration chromatography, Affinity chromatography, Ion-exchange chromatography, High Pressure Liquid Chromatography. Reversed phase chromatography, Hydrophobic interaction chromatography, Chiral chromatography, Counter current chromatography, Fast protein liquid chromatography, Two dimensional chromatography.	1,3	10
3.	**Electro analytical techniques and Hydrodynamic Techniques**: Principle, Electrophoretic mobility (EPM) estimation, factors affecting EPM, Instrument design & set-up, Methodology & Applications of Free and zone Electrophoresis – Paper electrophoresis, Gel electrophoresis, Poly Acrylamide gel electrophoresis, SDS PAGE, Capillary electrophoresis, Isoelectric focusing, Potentiometry, pH meter, Conductometry. Centrifugation & Ultracentrifugation-Basic principles, Forces involved, RCF Centrifugation, techniques- principles, types and applications. Viscometry- General features of fluid flow and nature	1,4,5	20

	of viscous drag for streamlined motion		
4	**Optical & Diffraction Techniques**. Principle, Instrument Design, Methods & Applications of Polarimetry, Refractometry, Circular Dichroism and optical rotatory dispersion: Plain, circular and elliptical polarization of light, Relation between CD and ORD, application of ORD in conformation and interactions of biomolecules. Flow cytometry	6	10
5	**Microscopic techniques**: Principle and working of Compound microscope, Phase contrast microscope, Interference microscope , Fluorescence microscope , Polarizing microscope , Scanning and Transmission Electron Microscopy, CCD camera, Introduction to Atomic force microscopy, Confocal microscopy.	6	10
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1.Principles and techniques of practical biochemistry: Keith Wilson and John walker, Cambridge

2. Modern Experimental Biochemistry. Rodney F Boyer. Nenjamin/ Cummings publishing company Inc. Redwood city, California

Further Reading:

1. Practical Biochemistry- Principles and techniques. Keith Wilson and John walker (Eds), University press, Cambridge UK.
2. Principles and Techniques of electron microscopy- Biological applications. M.A Hayat., Mac Millan Press, London UK.
3. Biophysical Chemistry: UpadhyayUpadhyay and Nath, Himalaya Publishing House
4. Chromatographic methods. A Braithwate and F J Smith. Chapman and hall, NewYork.
5. Gel Electrophoresis of Nucleic acids- A Practical approach. Rickwood D and BD Hames. IRL Press, New York. 53
6. Spectrophotometry and Spectrofluorimetry: A Practical Approach. Harris DA and CL Bashford (Ed.) IRL Press, Oxford.
7. Introduction to Spectroscopy. Donald L. Pavia Gary M Lipman, George S Kriz. Harcourt brace College Publishers, Orlands, Florida
8. Gradwohls Clinical Laboratory Techniques. Stanley s. Raphael. W.E. Company, London, UK
9. Fundamentals of molecular Spectroscopy: C N Banwell, Tata Mc Graw hill publishing Company Ltd.
10. Spectroscopic methods and analyses: Christopher Jones, Barbara Mulloy Adrian H.Thomas.
11. Methods in Modern Biophysics: Bengt Nolting, Springer.
12. Bio separations Science and Engineering: Roger G Harrison, Paul Todd, Scott .R. Rudge, Oxford University Press.

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MAHATMA GANDHI UNIVERSITY

BSM 21C 12 LABORATORY COURSE-3 MICROBIOLOGY AND IMMUNOLOGY

SchoolName	**School of Biosciences**					
Programme	**MSc.Microbiology/Biochemistry/Biophysics/Biotechnology**					
Course Name	**LABORATORY COURSE-3-MICROBIOLOGY AND IMMUNOLOGY**					
Type of Course	Core					
Course Code	**BSM 21C 12**					
Names of Academic Staff & Qualifications	**Dr. Jyothis Mathew** **MSc, PhD, Dr.Jisha M.S MSc, PhD and Dr.Radhakrishnan EK MSc, PhD**					
Course Summary & Justification	The course includes training on sterilization and disinfection techniques, morphological, cultural and biochemical study of microbes and antibiotic sensitivity tests. The content of the course also include serological techniques. The technical knowhow of basic microbiological and serological methods is essential for post graduate programmes in all branches of Biosciences.					
Semester	Second					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	5	5	120		130
Pre-requisites	Theoretical knowledge inMicrobiology and Immunology Basic laboratory skills					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Students will acquire skills on practice of sterile and safety precautions in a Microbiology laboratory.	A	1
2	Students will be able to prepare and sterilize media and to culture bacteria and fungi in laboratory	S	1
3	Students will be able to examine morphological, physiological and biochemical properties of bacteria	S/E	1
4	Students will be able to perform and interpret antibiotic sensitivity tests	S/E	1
5	Students will be able to test and analyse the efficacy of disinfectants	S/An	1
6	Students will be able to perform and interpret the various serological tests in a diagnostic laboratory	S/E	1
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	Microscopic examination of bacteria in living conditions Testing of motility Staining procedures	1,2,3	30
2	Sterilisation methodsCultivation of bacteria and fungi Study of cultural characteristics and biochemical reactions of bacteria Testing of disinfectants Antibiotic sensitivity tests	1,2,4,5	30
3.	Serological tests for the diagnosis of microbial infecdtions Agglutination and precipitation testsImmunodiffusion in gel ELISA	1,6	60
	Total Credits of the Course	2	120

TeachingandLearningApproach	**Laboratory Procedure (Mode of transaction)** Direct Instruction: lecture, Explicit Teaching, Demonstration, Hands on experimental sections, Skill acquisition by laboratory training
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Laboratory Skill Tests of maximum 20 marks 2. Seminar Presentation – Laboratory material and methods Maximum marks 10 3. Write a detailed report on instrumentation – 10 marks B. **Semester End Practical examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Medical Laboratory Manual for Tropical Countries Vol.2 Monica Cheesbrough ELBS, 2009
2. Mackie& McCartney Practical Medical Microbiology Churchil Livingstone, 1996

Further Reading:

1. Clinical Laboratory Methods Vol.2 Gradwohl The C.V.Mosby Company, 1981
2. London Practical Microbiology Dubey R.C.andMahaswari D.K. S.Chand& Company Ltd. New Delhi, 2002
3. Experiments in Microbiology, Plant pathology and Biotechnology, K.R.Aneja,, New Age International (P) Limited, New Delhi, 2003

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MAHATMA GANDHI UNIVERSITY

BSM 21C 13LABORATORY COURSE-4 MOLECULAR BIOLOGY AND GENETIC ENGINEERING

SchoolName	**School of Biosciences**					
Programme	**MSc.Microbiology/Biochemistry/Biophysics/Biotechnology**					
Course Name	**LABORATORY COURSE 4—(Molecular Biology and Genetic Engineering)**					
Type of Course	Core					
Course Code	**BSM 21C 13**					
Names of Academic Staff & Qualifications	**Dr Keerthi TR, Dr Jayachandran K, Dr Linu Mathew**					
Course Summary & Justification	The course is intended to provide experience to students in handling protein and DNA, its isolation, quantification and separation using electrophoresis. Also, the course focusses on the technique of PCR technology and proposes a training in PCR technique to equip the students for the present demand in the modern diagnostic methods.					
Semester	Second					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	5	5	120		130
Pre-requisites	Theoretical knowledge in **Molecular Biology and Genetic Engineering,**Basic laboratory skills					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	On completing the course, the students will be able to isolate nucleic acids and proteins from tissues/microorganisms	A	1
2	On completing the course, the students will be able to evaluate quantity and quality of nucleic acids	S	1
3	The students will be able to conduct PAGE and will be able to separate proteins using PAGE	S/E	2
4	The students will be able to amplify a DNA fragment selectively using the PCRtechnique	S/E	2
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I)			

and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	• PAGE- Protein separation Native PAGE-Reagent preparation, Apparatus handling, gel casting, electrophoresis and staining	1,3	45
2	• DNA isolation • Estimation of DNA • RNA isolation • Estimation of RNA • Separation of DNA and RNA by Agarose gel electrophoresis	1,2	60
3.	Selective PCR amplification of a desired fragment	1,2,4	15
	Total Credits of the Course	2	120

Teachingand LearningApproach	**Laboratory Procedure (Mode of transaction)** Direct Instruction: lecture, Explicit Teaching, Demonstration, Hands on experimental sections, Skill acquisition by laboratory training
Assessment Types	**Mode of Assessment** K. Continuous Internal Assessment (CIA) 1. Internal Laboratory Skill Tests of maximum 20 marks 2. Seminar Presentation – Laboratory material and methods Maximum marks 10 3. Write a detailed report on instrumentation – 10 marks L. **Semester End Practical examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Molecular cloning by Sambrook , Fritsch and Maniatis, Cold Spring harbour laboratories
2. Biochemical Methods Sadasivam and Manickam
3. Gel electrophoresis of proteins : A practical approach(second edition)B D H Ames and Rickwood D(eds) Oxford University press
4. Practical skills in Biomolecular Sciences, Weyers Jonathan, Reed Rob, Jones Allen, Holmes A D, Pearson publications

Further Reading:

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MAHATMA GANDHI UNIVERSITY

BSM 21E14: MICROBIAL TECHNOLOGY

School Name	**School of Biosciences**					
Programme	**MSc Biotechnology**					
Course Name	**Microbial Technology**					
Type of Course	**Elective**					
Course Code	**BSM 21E 14**					
Names of Academic Staff & Qualifications	**Dr. Keerthi TR**					
Course Summary & Justification	1. The course describe the application of microbes in various sectors 2. The course content explains the role of microbes and its utilization/application in various sectors especially in industrial & pharmaceutical area. 3. The course content also illustrates the various methods & process for production of bioactive compounds & products using microbes.					
Semester	Second					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisites	Basics of Microbiology					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**On completing this course, the student will be able to** Explain the methods for studying microbial genome and describe how metabolic & protein engineering help to enhance the production of microbial metabolites	**U/A**	1
2.	Describe the methods , process & production of various microbial based food and dairy products also students have able to explain microbes are food for animal and	**U/An**	1

	human		
3.	Students should explain the role of microbes as biofertilizer, biopesticide, fungicide, and herbicide and also able to describe the various plant microbe interactions	**U/A**	1
4.	Students have able to explain the methods and mechanism of microbes apply to protect various environmental sector.	**An/A**	1
5.	Illustrate the utilization of microbes in the production of industrial and pharmaceutical products	**S/C**	1
6.	Communicate effectively about a chosen topic in microbial technology both verbally and orally		1
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**Microbial Genomics:** Introduction to Microbial genomics, Structural Genomics, Functional genomics, Comparative Genomics, Meta Genomics - Genome analysis of extremophiles, Metabolic engineering and protein engineering for optimization of microbial products	**1,6**	**10**
2.	**Microbes in food & dairy industry:** Fermented foods-Introduction, Role & Advantages of fermented foods. Production of cheese, yoghurt, koji & Idli. Knowledge of other fermented dairy products. Single cell proteins-algae, bacteria, fungi, yeast & actinomycetes. Alcoholic beverages-Distilled and non distilled, Production of beer, wine & ethanol. Microbe as animal feed additives. Probiotics, Prebiotic & Synbiotics	**2,6**	**15**
3.	**Microbes in Agriculture**: Nitrogen fixation; Symbiotic & Non symbiotic Mechanism;Biofertilizers-Rhizobium, Azolla, Azospirillum, Algal Biofertilizers; Phosphate solubilizing microorganisms; Microbial biopesticide, biofungicide and herbicide; Micorrhiza; Plant –Microbe Interactions. Mushroom cultivation	**3,6**	**10**
4	**Microbes & Environment**: Biotechnology and pollution control; Use of immobilized microbial cell & enzyme in waste water treatment. Microbial biotransformation-Steroid, Microbial degradation of Herbicides, Insecticides & Pesticides; Bioremediation & Bioleaching	**4,6**	**10**
5.	**Industrial &Pharmaceutical Applications**: Methanogens & Biogas Production; Microbial Hydrogen production; Microbes in plastic industry - Bioplastics; Microbial biosensors- Micro oxygen electrode. Biochips; Biofilm; Bioactive compounds from microbes. Bioethanol & biodieseal production. Microorganism for Bioassay & as Bio weapon	**5,6**	**15**
	Total Credits of the Course	**3**	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** M. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks N. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Biotechnology Fundamentals and Applications, S.S. Purohit and S.S. Mathur; Agro Botanical Publishers India.
2. Microbial Biotechnology, Alexander N Glazer & Hiroshi Nikaido Cambridge University Press.
3. Microbial Biotechnology, Farshad Darvishi harzevili Hongzhang Chen.CRC Press.
4. Microbial Biotechnology Principle & Applications Lee Yuan Kein.World Scientific Press.

Further Reading:

1. Microbial Technology-Fermentation Technology Vol 1 & 11 Peppler Perinas Elsiver.
2. Biofertilizers in Agriculture, N.S.Subha Rao;Oxford & IBH Publishing Co.Pvt.Ltd New Delhi.
3. Essentials of Biotechnology, R.C.Sobti & Suparna.S.Pachauri. Ane Books Pvt.Ltd.
4. Fermentation Technology Vol I&II.
5. Soil Microbiology – N.S. Subha Rao, 1999
6. Agriculture Microbiology – Rangaswamy
7. Microbial control and pest Management – S. Jayaraj.
8. Food Microbiology – Frazier W.C and Westhoff D.C., Tata Mc Graw-Hill
9. Food Microbiology – Rose A.H. in Economic Microbiology, Academic Pr

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MAHATMA GANDHI UNIVERSITY

BSM 21E 15ECOLOGY AND ENVIRONMENT

School Name	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**ECOLOGY AND ENVIRONMENT**					
Type of Course	Elective					
Course Code	**BSM 21C 15**					
Names of Academic Staff & Qualifications	**Dr J G RAY**					
Course Summary & Justification	The course is designed to equip students in perceiving, understanding and analyzing environmental problems from an ecological perspective, and a critical analysis of the existing control measures from a holistic perspective.					
Semester	First					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg: Authentic learning Collaborative learning Independent learning	60	18	0	28	106
Pre-requisite	**Knowledge in Biology at Graduate level**					

No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**
1	Students will be able to understand and communicate the sustenance of natural biological systems on the earth effectively	R/U/A	1
2	They will acquire skills in explaining all kinds of interrelationships in natural biological systems	U/A	1

3	Students will be able to explain environmental degradation and pollution as outcomes of ignorant and irresponsible human actions	U/An/Ap	1
4	Students will be able to understand the significance of biodiversity and its conservation in the sustenance of natural ecosystems	An/Ap	1
5	Overall, students will be skilful in analyzing as well as designing and maintaining of environmental sustainability of all kinds of developmental activities	R/U/A/An /Ap	1

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

Module No	Module Content	CO	Hours
1	**Introduction to Ecology and different ecological objects**: Basic concept of the environment – components of the environment, the definition of ecology, ecological things. Autecological and Synecological concepts: **A.Population Ecology (Autecological concepts)**: **(a)** Characteristics of populations **(b)** Genecology - ecads, ecotypes, ecospecies, coenospecies; k-selection and r-selection populations **B. Synecological concepts(a)** Ecological processes of community formation, ecotone, edge effect. Classification of communities - criteria of classification, dynamic system of classification by Clement **(b)** Special plant communities - quantitative, qualitative and synthetic characteristics of plant communities**, (c)** Dynamic community characteristics - cyclic replacement changes and cyclic no-replacement changes	1,2,3	10 hrs
2	**Ecological succession -(a)** The concept – autogenic and allogenic succession, primary and secondary, autotrophic and heterotrophic **(b)** Retrogressive changes or the concept of degradation, concept of climax or stable communities, resilience of communities, ecological balance and survival thresholds **Biosphere and Ecosystem** - **(a)** Significance of habitat, biodiversity, ecological niche, trophic level, primary and secondary productivity, food chains, food webs, ecological pyramids, energy flow and nutrient cycles **(b**) Comparative study of the significant world ecosystems: Different aquatic and terrestrial ecosystems concerning their productivity, 0.5 57 biodiversity, energy flow, food chains and trophic levels	1,2,4	10 hrs

3.	**Natural Resources:** Soil, water and air Resources – soils and parent materials – ecology of soil fertility; Fresh water and marine resources – global distribution of water resources – surface and groundwater resources – water conservation – prevention of marine pollution – conservation of marine resources; Atmospheric resources – the structure of atmosphere – climate and weather – climatic factors – precipitation, wind temperature, aerosols	1,2	10 hrs
4	**Environmental pollution**: **(a)** Definition and classification **(b)** Water pollution: Water quality parameters and standards, different types of pollutants and their consequences. Types of water pollution, prevention and control - watershed management, different kinds of wastewater treatments; Phyto and bioremediation **(c)** Air pollution: Air quality standards and index, ambient air monitoring using high volume air sampler, types and sources of air pollutants, air pollution and human health hazards, control of air pollution **(d)** Noise pollution **(e)** Radioactive and thermal pollution: Causes and hazardous effects, effective management **(f)** Concept of solid wastes **(g)** Pollution Control - Bioremediation, Phytoremediation, bioaugmentation, biofilms, biofilters, bioscrubbers and trickling filters. Use of bioreactors in waste management	3	20 hrs
5	**Climate Change and other Global Environmental issues** - Factors responsible for climate change, Climate change mitigation – global conventions and protocols on climate change - El-Nino and La Nina phenomenon and its consequences; Environmental laws, environmental monitoring and bioindicators, environmental safety provisions in the Indian constitution, major ecological laws in free India; UNEP and its role in climate change control– IPCC, UNFCC, annual environment summits – 1973 Stockholm conference to 2015 Paris Conference – new developments of annual UNFCC meetings in the coming years - Future Earth Programme	5	10 hrs
	Total Credits of the Course	**3**	**60 hrs**

Books for Reference

Compulsory Reading:

1. MC Dash (1993) Fundamentals of Ecology, Tata McGraw Hills

2. Odum EP 3rd Edition (1991) Fundamentals of ecology, Saunders and Com

Optional Further Reading

1. Barbour MD et al. (1980) Terrestrial plant ecology. The Benjamin-Cummings Pub. Com 2.

2. Benton AH and Werner WE (1976) Field biology and Ecology, Tata McGraw Hill

3. Blanco-Canqui and Humberto LR (2008) Principles of Soil Conservation and Management, Springer

4. Molles MC (2012) Ecology – Concepts and applications, 6th Edition, Mc Graw Hill	
Course evaluation: **Assignments & Seminar (10 marks each); Two internal test papers (20 effects) end semester examination (60 marks)**	

MAHATMA GANDHI UNIVERSITY

BSM 21E16: NEUROBIOLOGY

SchoolName	**School of Biosciences**					
Programme	**MSc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**NEUROBIOLOGY**					
Type of Course	Elective					
Course Code	**BSM 21E 16**					
Names of Academic Staff & Qualifications	**Dr. R. Harikumaran Nair** MSc, PhD					
Course Summary & Justification	This course is designed to provide an overview of Neurobiology. Stress will be placed on methods and concepts rather than facts alone. The course will proceed from the basic biophysical properties of neurons and glia to the physiological basis of learning, memory, and sensory processing					
Semester	Second					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics Knowledge in Physiology**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Students should be capable of effectively communicating how neural system works	U	1
2	Students should be able to explain electricity and the biophysics of cell	E	2
3	Students should describe how do neurons talk to one-another	A	2
4	Students should be able to explain how neural circuits organize information	A	1
5	Students should be able to narrate how is information stored	E	2
6	Lastly, students should gain a general understanding how is information collected and processed.	I	2
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Introduction to neurobiology, the structure and distinguishing features of neurons, how is a neuron recognized? The architecture of nervous systems. Neuronal model systems. Chemical/electrical synapses. Recording/monitoring techniques.	1,6	10
2	Ionic basis of the resting potential. Maintenance of resting membrane potential, passive and active mechanisms, channels and pumps, ionic permeability	2,6	10
3.	Action potentials and ion channels, Mechanism of nerve action potential: Characteristics of action potential, initiation and propagation of action potential, voltage dependent sodium channels, mechanism of action potential propagation, factors affecting the speed of action potential propagation, molecular properties of voltage sensitive sodium channels, molecular	3,6	20

	properties of voltage dependent potassium channels, calcium dependent action potentials, voltage- clamp analysis of action potentials		
4	Synaptic transmission: Chemical and electrical synapse, neurotransmitter release, synaptic potential, excitatory synaptic transmission between neurons, excitatory neurotransmitters, inhibitory synaptic transmission, inhibitory neurotransmitters, neurotransmitter gated ion channels, presynaptic inhibition and facilitation, neuronal integration, synaptic transmission at neuromuscular junction	4,6	10
5	Synaptic plasticity, language and cognition: Short term changes in synaptic strength, long term changes in synaptic strength, modification of synaptic strength in reflex circuits, learning, language function and cortical areas involved in language, cognition, dementia and loss of cognitive abilities	5,6	10
	Total Credits of the Course	3	60

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading

1. Basic Neurochemistry- Molecular, cellular and medical aspects. George J Siegel, Bernard W Agra noff R, Wayne Albers, Stephen K Fisher & Michael D Uhler

2. Neurobiology: Molecules, cells and systems. Gary G Mattews

3. From Neuron to Brain- John G Nicholls, A Robert Martin, Bruce G Wallace & Paul A

Fuchs

Further Reading:

1. Neuroscience, edited by Purves, Augustine, Fitzpatrick, Hall, LaMantia, Mooney, Platt and White. Sinauer (2018) Sixth Edition.
2. Foundations of Neurobiology, Delcomyn, F. 1st edition W. H. Freeman and Company (1998)
3. Behavioral Neurobiology: An Integrative Approach, Zupanc, G. K. H. Oxford University Press. 2nd edition (2010)
4. Neurobiology: molecules cells and systems Gary G. Mathews 2nd edition. Blackwell Science Inc. (2001).
5. Neuroscience: exploring the brain. Bear, M., Connors, B.W. and Paradiso, M.A. 2nd edition Lippincott, Williams and Wilkins (2001)

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	MAHATMA GANDHI UNIVERSITY
	BSM 21E 17 ENVIRONMENT SCIENCE

SchoolName	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**ENVIRONMENT SCIENCE**					
Type of Course	Elective					
Course Code	**BSM 21E 17**					
Names of Academic Staff & Qualifications	**Dr.JISHAM.S.**					
Course Summary & Justification	This course on environmental Science deals with principles and scope of environment science. The objective of the course content is to create a sound awareness about the environment impact and its monitoring and Predict the consequences of human actions on the web of life, global economy and quality of human life The course content is designed with a view to augment CSIR/UGC syllabus					
Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of cell biology and genetics**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Gain in-depth knowledge on natural processes that sustain life and govern economy.	U/A	1
2	Able to describe the principles of ecology	U/ C	2
3	Develop critical thinking for shaping strategies (scientific, social, economic and legal) for environmental protection and conservation of biodiversity, social equity and sustainable development.	R/An	2
4	Acquire values and attitudes towards understanding complex environmental-economic social challenges	U/R	1
5	Understand the current environmental problems and preventing the future ones.	U/R	1
6	Create an insight to the strategies and methodologies of environmental impact assessment	An/ C	2
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Definition, principles and scope of environmental science, Earth, Man and environment, ecosystem, pathways in ecosystem. Physic-Chemical and Biological factors in the environment Geographical classification and Zones. Structure and functions of ecosystem, Abiotic and biotic components, energy flows, food chains, Food, web, Ecological pyramids, types and diversity Terrestrial (Forest, grass land) and Aquatic (Fresh water, marine, eustarine) ecosystems. mineral cycling. Habitat and niche. Major terrestrial biomes. Impact of microorganisms on global ecology, microorganisms in extreme environment	1,2,3	10
2	Definition, Principles and scope of ecology, Human ecology and Human settlement, evolution, origin of life and speciation Population ecology characteristics and regulation. Community ecology structure and attributes. Levels of species diversity and its management, Edges and ecotones. Ecological succession. Concept of climax. Common Flora and fauna in India. Endangered and Threatened Species	2,3	10

3.	Biodiversity status, monitoring and documentation Biodiversity management approaches. Conservation of biological diversity, methods and strategies for conservation. Natural resources, conservation and sustainable development. Hotspots of biodiversity, National parks and Sanctuaries	3,4	20
4	Environmental pollution- Air: Natural and anthropogenic source of pollution, Primary and Secondary pollutants , Methods of monitoring and control of air pollution, effects of pollutant on human beings, plants animals, material and on climate, Acid rain, Air Quality standards Water: types, Sources and consequences of water pollution, Physio-chemical and Bacteriological sampling and analysis of water quality, Soil: Physio-chemical and Bacteriological sampling as analysis of soil quality, Soil pollution-control, Industrial waste effluents, and heavy metals Their interaction with soil components, Noise: Sources of noise pollution, Noise control and battement measures. Impact of noise on human health, Radioactive and thermal Pollution. Bioremediation- Strategies for bioremediation, Biosensors, biological indicators of pollution and monitoring. Detoxification of hazardous chemicals, mycotoxins. Biological weapons	5	20
5	Introduction to environmental impact analysis, Impact Assessment Methodologies Generalized approach to impact analysis, Guidelines for Environmental Audit Introduction to environmental Planning, Environmental priorities in India and Sustainable development, Environment protection-issues and problems, International and national efforts for environment Protection. Global environmental problems-Ozone depletion, global warming, climatic change, desertification, green movement, ecofeminism. Current environmental issues in India	6	10
	Total Credits of the Course		

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** O. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks P. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Jonathan B (2016) Principles of Evolution, Garland Science, Taylor and Francis.
2. Odum E. P and Barret G W. Fundamentals of ecology. W. B Saunders company, Philadelphia
2. Chapman and Reiss, Ecology principles and applications. Cambridge University

Further Reading:

1. Jobes A. M., Environmental biology, Routledge, London.
2. Odum E. P. Basic ecology. Saunders College.
3. A textbook of environmental sciences, Arvind kumar.
4. Alleby M.Basics of environmental science. Routledge, Newyork
5. Cunningham, W. P and Siago, B. W, Environmental science.
6. Kewin T. P and Owen C. A., Introduction to global environmental issues. Routledge, London.
7. Chiras,D.D, Environmental science Cell and Molecular Biology by De Robertis E.D.P, 8th Edition

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BSM 21E 18 MOLECULAR MICROBIOLOGY

SchoolName	School of Biosciences					
Programme	M.Sc. Microbiology/Biotechnology/Biochemistry/Biophysics					
Course Name	MOLECULAR MICROBIOLOGY					
Type of Course	Elective					
Course Code	**BSM 21E18**					
Names of Academic Staff & Qualifications	Dr.Radhakrishnan E.K. M.Sc., Ph.D					
Course Summary & Justification	This course on Molecular Microbiology deals with the applications of various molecular biological techniques in Microbiology. This course is an important branch of Microbiology. Rapid identification of microorganisms is very important for the clinical, diagnostic and research purposes and the methods used for the same have developed significantly with the advances in Molecular biology. The content in this course has been designed with an objective to provide detailed understanding on the techniques, principle and applications of molecular biology for the microbial identification, production of recombinant proteins and also for studying the unculturable microorganisms through metagenomics. This will enable the students to identify the research, learning and job opportunities based on the latest developments in this subject.					
Semester	Second					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding on microorganisms and molecular biology Knowledge in any branch of Life science					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1.	Students will able to understand and explain molecular biological applications in microbiology	R/U	1
2.	Students will able to learn rapid methods used for the microbial identification	R/U	1
3.	Students will able to understand the functioning of human microbiome and its beneficial role	U/ An/E	1
4.	Students will become able to understand molecular basis of microbial virulence	U/An/A	1
5.	Students will able to apply the knowledge for advanced microbiological applications	C/S	1
6.	Students will able to identify the research and technical opportunities in molecular microbiology	A/S	10
**Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	Molecular biology of Microbial evolution, rRNA sequence and cellular evolution, Signature sequence and phylogenetic probe. Identification and characterization of microorganisms, Molecular methods for microbial identification, Molecular typing methods: Bacterial strain typing, Pulsed Field Gel Electrophoresis, PCR-based microbial typing, Genotyping by Variable Number Tandem Repeats, Multilocus Sequence Typing, Automated Ribotyping	1,2	20
2	Unculturable bacteria and Metagenomics, Methods used in metagenomics, New generation sequencing technologies for metagenome study, Human microbiome, Importance of human microbiome in relation to human health and disease.	3	20
3.	Molecular basis of microbial virulence. Bacterial adherence: basic principles, effects of adhesion on bacteria and host cells. Bacterial invasion of host cells; mechanism. Bacterial toxins: classification based on molecular features, Molecular detection and characterisation of bacterial pathogens, detection of bioterrorism.	4	10
4	Microbial production of recombinant proteins: expression, purification and applications, Microbes in plant transformation, Agrobacterium tumefaciens T-DNA transfer process, Application of microorganisms for combinatorial and engineered biosynthesis, Engineering *E.coli* for the production of curcumin	5,6	10
	Total Credits of the Course	3	

Teaching And Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction, Active co-operative learning, Seminar, Group Assignments Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar - Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. Semester End examination – 60 marks

REFERENCES

Compulsory Reading:

1. Molecular Microbiology – Diagnostic Principles and Practice, David H. Persing, Fred C. Tenover, James Versalovic, Yi-Wei Tang, Elizabeth R. Unger, David A. Relman, Thomas J. , ASM Press., 2016
2. Brock Biology of Microorganisms- Michael T. Madigan and John M.Martinko, Prentice Hall, 2015

Further Reading:

3. Microbial Physiology – Albert G. Moat, John W. Foster and Michael P. Spector , 2002
4. Metagenomics for Microbiology, Jacques Izard Maria Rivera , 1st edition, Academic Press Published Date: 12th November 2014
5. Production of Recombinant Proteins: Novel Microbial and Eukaryotic Expression Systems, Gerd Gellissen, May 2005Longman,2013

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MAHATMA GANDHI UNIVERSITY

BSM 21E 19DEVELOPMENTAL BIOLOGY

School Name	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**DEVELOPMENTAL BIOLOGY**					
Type of Course	Elective					
Course Code	**BSM21E 19**					
Names of Academic Staff & Qualifications	**Dr J G RAY**					
Course Summary & Justification	The course is designed to equip students in perceiving, understanding, and analyzing reproductive and embryological developmental processes in plants to apply the principles towards increasing plant productivity through breeding.					
Semester	First					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	E.g., Authentic learning Collaborative learning Independent learning	60	18	0	28	106
Pre-requisite	**Knowledge in Botany at the Graduate level**					

No.	Expected Course Outcome	Learning Domains	PSO No.
1	Students will be able to understand and communicate the reproductive and developmental events in plants effectively	R/U/A	1
2	They will acquire the skills to explain all kinds of reproductive parts and seed developmental processes, including seed storage in plants	U/A	1

3	They will be able to explain how developmental processes initiates and proceeds in plants	U/An/Ap	1
4	Students will be able to explain the specific developmental process and its ultimate impact on the productivity or successful completion of lifecycle in plants	An/Ap	1

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

Module No	Module Content	CO	Hours
1	**Introduction:** Basic concepts of developmental Biology; An overview of plant and animal development, Potency, Commitment, Specification, Induction, Competence, Determination and Differentiation morphogenetic gradients; cell fate and cell lineages; stem cells; genomic equivalence and the cytoplasmic determinants; imprinting; mutants and transgenics in the analysis of development	1,2,3	20 hrs
2	**Development in flowering plants**: **(a)** Angiosperm life cycle **(b)** Anther: Structure and development, microsporogenesis, male gametophyte development. Palynology: Pollen morphology, exine sculpturing, pollen kit, NPC formula. Applications of palynology- palynology concerning taxonomy. Viability of pollen grains Pollination, pollen germination, growth and nutrition of pollen tube**. (c)** Ovule: Structure, ontogeny and types. Megasporogenesis. Embryosac – development, classes, ultrastructure, and nutrition of embryosac. Female gametophyte development.	1,2,3	10 hrs
3.	**Fertilization in Plants:** Double fertilization; embryo development - different types. Endosperm development, types of endosperm, haustorial behaviour of endosperm. Xenia and metaxenia. Polyembryony – types and causes. Seed formation, dormancy and germination. Apomixis, Parthenogenesis.	1,2,3	10 hrs
4	**Morphogenesis and organogenesis in plants:** Shoot and root development; Leaf development and Phyllotaxy. Transition to flowering, floral meristems and floral development; Homeotic genes in plants; Senescence, programmed cell death and hypersensitive response in plants	4	20 hrs
	Total Credits of the Course	**3**	**60 hrs**

Books for Reference	
Compulsory Reading: 1. Maheswari P. 1950. An introduction to the embryology of Angiosperms. McGraw Hill 2. Wolpert L, C Tickle and AM Arias (2015) Principles of development	
Optional Further Reading 1. Krishnamurthy KV (2015) Growth and Development in Plants 2. Raghavan V (2000) Developmental Biology of Flowering Plants 3. Gilbert SF (2000) Developmental Biology 4. Developmental Biology, 8th Ed, Gilbert 5. Developmental Biology Paperback – 2008 by Werner A. Muller	
Course evaluation: **Assignments, 1 Seminar, and one assignment (10 marks each) Two internal test papers (20 marks) end semester examination (60 marks)**	

THIRD SEMESTER

	MAHATMA GANDHI UNIVERSITY
	BSM 21C 20ENZYMOLOGY

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**BIOCHEMISTRY**					
Type of Course	Core					
Course Code	**BSM 21C 20**					
Names of Academic Staff & Qualifications	**Prof. M S Latha – M. Sc (Biochemistry), PhD**					
Course Summary & Justification	The course is designed to get a deep knowledge of the mechanisms by which cellular reactions are accelerated. The course builds a base for the students to understand and predict the metabolism of all living things and provide basics of drug development process related to enzyme targets and enzyme therapy					
Semester	Third					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Eg. Authentic learning Collaborative learning Independent learning	80	20	0	40	140
Pre-requisite	Basic idea about protein structure and function					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To give details of different characteristics of enzymes, enzyme classification and nomenclature, different factors that affect their activity and their mechanisms of action.	U	5
2	To contrast different modes of enzyme inhibition and regulation.	An/E	5
3	To describe the structure and functions of vitamins and hormones	U	8
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I)			

and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hours
1	**Introduction to enzymes:** Holoenzyme, apoenzyme, and prosthetic group; Interaction between enzyme and substrate- lock and key model, induced fit model, Features of active site, activation energy. Enzyme specificity and types; Enzyme Commission system of classification and nomenclature of enzymes (Class and subclass with one example) **Mechanisms of enzyme action-** Rate Enhancement Through Transition State Stabilization, Acid-base catalysis, covalent catalysis, metal ion catalysis (eg: Serine Proteases: cystenyl protease, aspartyl protease, metalloprotease, Lysozyme). **Coenzymes and their functions** - NAD, $NADP^+$, FAD, FMN, lipoic acid, TPP, pyridoxal phosphate, biotin and cyanocobalamin **Isolation and characterization of enzymes:** Isolation of enzymes and the criteria of purity; Characterization of enzymes- active site mapping, Measurement and expression of enzyme activity, enzyme assays. Definition of IU, katals, enzyme turnover number and specific activity.	1	20
2	**Enzyme kinetics:** Factors affecting the velocity of enzyme catalyzed reaction- enzyme concentration, temperature, pH, substrate concentration, inhibitors and activators (explanation with graphical representation). Derivation of Michaelis -Menten equation and Km value determination and its significance, Definition of V_{max} value of enzyme and its significance, Lineweaver- Burk plot, Eadie-Hofstee and Hanes plots.Bi-substrate reactions: Classification, Reaction mechanisms.	1	20
3.	**Enzyme inhibition:** Reversible and irreversible – examples. Reversible- competitive, noncompetitive, uncompetitive inhibition and mixed inhibition, Irreversible inhibition- mechanism based inactivators, affinity labels, group specific inhibitors; Graphic Determination of Inhibitor Type; Dose—Response Curves of Enzyme Inhibition; Mutually Exclusive Binding of Two Inhibitors; Structure—Activity Relationships and Inhibitor Design; Tight Binding Inhibitors: Identifying Tight Binding Inhibition, examples; Time-Dependent Inhibition: examples; Distinguishing between modes of inhibitor interaction with enzyme	2	20
4	**Regulation of Enzyme activity: Different c**ovalent modifications,; Zymogen form of enzyme and zymogen activation; Multienzyme complexes and their role in regulation of metabolic pathways; Allosteric regulation: example Aspartate trascarbamoylase, Sigmoidal kinetics of allosteric enzymes, Models of Allosteric Behavior, Effects of Cooperativity on Velocity Curves. Isoenzymes- Lactate dehydrogenase and creatine phosphokinase.	2	10

5	**Application of enzymes:** Applications of enzymes in industry (eg: in food industry, paper and leather industry, detergent industry and waste management).Diagnostic and therapeutic enzymes; Applications of enzymes in life science research, Ribozymes, Abzymes, Immobilised enzymes, Biosensors, synthetic enzymes, Enzyme engineering	2,3	10
	Total Credits of the Course	4	80

Books for Reference

Compulsory Reading:

1. Fundamentals of Enzymology: The Cell and Molecular Biology of Catalytic Proteins by Nicholas C. Price, Lewis Stevens, and Lewis Stevens (2000) Publisher: Oxford University Press, USA ISBN: 019850229X ISBN-13: 9780198502296, 978-0198502296
2. Enzyme Kinetics: A Modern Approach Book: Enzyme Kinetics: A Modern Approach by Alejandro G. Marangoni (2003) Publisher: Wiley-Interscience ISBN: 0471159859 ISBN-13: 9780471159858, 978-0471159858
3. Principles Of Biochemistry, 4/e (2006) by Robert Horton H , Laurence A Moran, Gray Scrimgeour K **Publisher:** Pearsarson **ISBN:** 0131977369, **ISBN-13:**9780131977365, 978-0131977365
4. Biochemistry 6th Edition (2007) by Jeremy M.berg John L.tymoczko Lubert Stryer **Publisher:** B.i.publicationsPvt.Ltd **ISBN:**071676766X **ISBN-13:** 9780716767664, 978-716767664
5. Lehninger Principles of Biochemistry, Fourth Edition by David L. Nelson David L. Nelson (Author)

Further Reading:

- Enzyme Kinetics and Mechanisms by Taylor Publisher: Spring ISBN: 8184890478 ISBN-13: 9788184890471, 978-8184890471
- Enzyme Mechanism by P.K. Shivraj Kumar (2007) Publisher: RBSA Publishers ISBN: 8176114235 ISBN-13: 9788176114233, 978-8176114233
- Enzymes and Enzyme Technology by Kumar (2009) Anshan Pub ISBN: 1905740875, ISBN-13: 9781905740871, 978-1905740871
- Enzymes in Industry: Production And Applications by Aehle W (2007) Publisher: John Wiley & Sons Inc ISBN: 3527316892 ISBN-13: 9783527316892, 978-3527316892
- Enzymes: Biochemistry, Biotechnology, Clinical Chemistry (second Edition) by Trevor Palmer, Philip Bonner (2007) Publisher: Horwood Publishing Limited ISBN: 1904275273 ISBN-13: 9781904275275, 978-1904275275

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** **Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks **Semester End examination – 60 marks**

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MAHATMA GANDHI UNIVERSITY

BSM 21C 36: MOLECULAR BIOPHYSICS

SchoolName	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**MOLECULAR BIOPHYSICS**					
Type of Course	Core					
Course Code	**BSM 21C 36**					
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S**					
Course Summary & Justification	This course is designed to provide an overview of chemical structure of various macromolecules involved in propagation of life. This course comprehends the influence of macromolecular three dimensional structure on their function Appreciate the relevance of bioinformatics to the function of biological macromolecules					
Semester	Third					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of** Molecular biophysical techniques and bioinformatics					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To describe and discuss the relationship between the structure and function of biological macromolecules	E	8
2	To determine the role of biological receptors	U/ An	8
3	To narrate the cell membrane structure and function	R	9
4	To explain how to use bioinformatics tools and software's	S	3
5	To know different molecular biology databases and formats in which data is stored	E	3
6	To describe the concepts of computer aided drug discovery	An/ C	3
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**Structure and conformations of proteins**: Structure stabilizing forces, Super secondary structural domains, Domain and motifs, Classification and role of reverse turn, Hydrogen bonding in globular proteins. Main chain and side chain conformation of globular proteins. Globular and fibrous proteins. Conformation, organization and interactions of structural proteins such as collagen, alpha-keratin, silk fibroin, actin and myosin. Examination of 3D structure of chymotrypsin and Rubisco. Folded conformation of globular proteins- lysozyme and cytochromes. **Protein evolution**: Concept of protein evolution, Protein speciation, phylogeny, phylogenetic tree- cladogram, phylogenetic study of cytochrome c. Homologies and analogies- convergent, divergent and parallel evolution **Structure and conformation of polysaccharide** - Amylase, Cellulose, Chitin.	1	20
2	**Biological receptors**: Drug-receptor interaction. Protein ligand interaction: Ligand- binding sites of Immunoglobulins, substrate-binding sites of Seriene proteases, Haem-binding sites, Nucleotide-binding sites, Binding sites for phosphoryl groups, Interaction of	2	10

	proteins with other macromolecules- lipoprotein, Glycoprotein and Nucleoprotein.		
3.	**Membrane biophysics**: Various membrane models, Carbohydrate, Lipids & Proteins components of cell membrane, Electrical properties of lipids and proteins, Principles of membrane organization & stability, Molecular motion in membrane & membrane fluidity, Structure of membrane proteins- Integral membrane proteins, Peripheral membrane proteins, Polypeptide toxins, Membrane protein complex, Transport across membrane, Selective permeability of biomembrane, Selectivity & ion specificity of biomembrane, Ion channel structure and gating function, Ion channel types and characterization, Role of carriers in ion transport.	3	20
4	**Bioinformatics:** Nature and scope of bioinformatics, Biological databases and various file formats, Sequence retrieval and submission, Overview of available Bioinformatics resource on the web- NCBI, EBI, EXPASY etc, Sequence alignment – BLAST and FASTA, Pairwise sequence alignment:- global and local alignment, multiple sequence alignment, Clustal W, Clustal X, Pattern and profile databases – PROSITE and BLOCKS, Structure databases – understanding structures from Protein Data Bank, Metabolic databases – post translational modification databases, Tools for viewing and interpreting macromolecular structures- Swiss pdb viewer, Webmol, Rasmol, PyMol, Chimera.	4,5	20
5	**Application of Bioinformatics in Drug discovery**: Role of bioinformatics in drug discovery, Different methods of target identification and validation, Ways of lead identification and optimization, *In-silico* prediction of ADMET properties of drug molecules, Molecular docking and prediction of drug quality. Bioinformatics companies.	5,6	10
	Total Credits of the Course	4	

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10

	3. Write a detailed report on a given topic based on research findings and literature search – 10 marks **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading: 1. Introduction to Proteomics Tools for the New Biology: Daniel C. Liebler, Humana press 2. Proteins, Structure and molecular properties, Thomas E Creighton
Further Reading: 1. Molecular Biophysics- Volkenstain M.V 2. Biopolymers, AP- Watson,A.G & Blackwell,J 3. Principles of protein structure Schulz, G.E. & Schimmer, R.H.. 4. Structure and Molecular Principles, W.H. Freeman & Co.Creighton, T.E. 5. Saenger,W., Principles of Nucleic Acid Structure, Springer. 6. Protein-Protein Interactions: Erica Golemis, CSHL Press. 7. Protein Architecture A practical Approach: A.M.Lesk, 8. Molecular and Cellular biophysics, Jack A. Tuszynski, Chapman & Hall/ CRC

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	MAHATMA GANDHI UNIVERSITY
	BSM 21C 37: ELECTROPHYSIOLOGY

SchoolName	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**ELECTROPHYSIOLOGY**					
Type of Course	Core					
Course Code	**BSM 21C 37**					
Names of Academic Staff & Qualifications	**Dr. R. Harikumaran Nair**					
Course Summary & Justification	The course will deal with the methodological aspects ofelectrophysiology The course will cover the main methods used for humanelectrophysiology The course will empower the students to carry out and interpret the various neurophysiological techniques					
Semester	Third					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of** Biophysics and Biostatistics					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.

1	Estimate the feasibility of recording and stimulating any electrophysiological signal from first principles of biophysics	E	1
2	Describe the working principles of all currently available medical devices for therapeutic modulation of neuralsignals	U/ An	1
3	Identify technological and biological limitations in the treatment of clinical disorders of the heart, motor control and special senses	R	1
4	Record and analyze common electrophysiological signals, including ECG, EMG andEEG	An/ C	2
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	Basic principles of electricity, overview of electrophysiological instrumentation, types of electrophysiological recordings, current voltage relationship and the membrane potential	1,2	10
2	Electrophysiology of heart, electrocardiogram (ECG), source of ECG voltage- dipole theory, vector analysis of ECG, Brain Potentials, electroencephalogram (EEG), source and mechanism of formation of rhythmic pattern of EEG, characteristics of EEG waves	1,2, 4	10
3.	Event related potential (evoked potential)-types, characteristics, and significance electromyogram (EMG) – motor unit potential, physiological significance and analysis of EMG	1,2, 3,4	20
4	Structure of retina, electrical response of the photoreceptors to light, molecular mechanism of phototransduction process, auditory system in brain, mechanoreceptors of vibration sense, cochlear mechanics	1	10
5	Chemical sense, structure of olfactory epithelium, chemo transduction in olfactory receptor cells, processing of olfactory information in brain, structureofthetastebuds,chemotransductionintastereceptorcells,processing of taste information in brain	1	10
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. From Neuron to Brain. -John G. Nicholls, A. Robert Martin, Bruce G. Wallace & Paul A. Fuchs.
2. Ion channels. Molecules in Action-David J. Aidley & Peter R.Stanfield
3. The Physiology of Excitable Cells- David J.Aidley

Neurobiology: Molecules, cells and systems. Gary GMattews

Further Reading:

1. Principles of Neuroscience.Kandel, Schwartz and Jessell, ed., McGraw-Hill, 4th ed.,2000.
2. Glover,BM,Brugada,P.eds.ClinicalHandbookofCardiacElectrophysiology.NewYork, NY: Springer;2016.
3. Sra,JS,Akhtar,M.eds.PracticalElectrophysiology.Minneapolis,MN:Cardiotext Publishing; 2014.

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MAHATMA GANDHI UNIVERSITY

BSM 21C 38LABORATORY COURSE 5

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry**					
Course Name	**LABORATORY COURSE 5 BIOPHYSICS**					
Type of Course	Core					
Course Code	**BSM 21C 38**					
Names of Academic Staff & Qualifications	**Dr.Anie Y – M. Sc (Biochemistry), PhD** **Guest faculty– M. Sc (Biochemistry), PhD**					
Course Summary & Justification	The course is designed to develop in students the essential skills to perform enzyme assays and related techniques. This will enhance the practical skillsto perform enzyme-related methods and computational drug discovery process.					
Semester	Third					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg. Authentic learning Collaborative learning Independent learning	10	20	120	30	180
Pre-requisite	Generalidea on reagents and solvents					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.

1	To design and perform enzyme assays	C/S	5
2	To extract and purify enzymes from different sources and to examine their kinetic behavior	A/An	5
3	To prepare and charaterise immobilized enzymes	A	5
4	To assess the activity of enzymes by computational methods	E	5
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hours
1	Enzyme assays: Practical concepts	1	15
2	Extraction of enzymes and assay: Acid phosphatase from Fresh Potato (*Solanum tuberosum*) β- amylase from Sweet potato (*Ipomoea batates*) Urease from Jack bean (*Canavalia ensiformis*) Phytase from Seeds	2	30
3.	Enzyme Kinetics: Effect of Substrate Concentration on velocity of Enzyme catalyzed reaction: Determination of K_M and V_{max} using Line weaver- Burk plot Effect of Temperature on velocity of Enzyme catalyzed reaction: Determination of Q_{10} Effect of pH on velocity of Enzyme catalyzed reaction: Effect of activators on velocity of Enzyme catalyzed reaction: Determination of type of inhibition using Line-weaver Burk plot	2	45
4	Immobilized enzyme: Immobilisation of α- amylase enzyme Assay of activity of immobilized enzyme Effects of Temperature and pH on Immobilized enzymes	3	15
5	Docking of Enzymes with ligand molecules using docking softwares Determine the drug likeliness of ligand molecules Determining Binding energies of ligand with receptors	4	15

	Determining Ki values		
	Total Credits of the Course	2	

Books for Reference

Compulsory Reading:

Introductory Practical biochemistry, S. K. Sawhney &Randhir Singh (eds) Narosa Publishing House, New Delhi, ISBN 81-7319-302-9, p 195 – 303

Standard Methods of Biochemical Analysis, S. K. Thimmaiah (ed), Kalyani Publishers, Ludhiana ISBN 81-7663-067-5, p 12 - 182.

Further Reading:

Hawk's Physiological Chemistry, Bernard L. Oser (ed) TATA McGRAW Hill Publishing Company LTD, New Delhi, p 60 – 127, 1317- 1334

Experimental Biochemistry: A Student Companion, BeeduSasidhar Rao & Vijay Deshpande (ed), I.K International Pvt. LTD, New Delhi ISBN 81-88237-41-8, p 13- 17, p 49 - 72

Practical Biochemistry, R.C. Gupta & S. Bhargava (eds) CBS Publishers and Distributors, New Delhi, ISBN 81-239-0124-0 p 9 – 27

. Practical Clinical Chemistry, Harold Varley, CBS Publishers and Distributors, New Delhi,

TeachingandLearningApproach	**Laboratory Procedure (Mode of transaction)** Direct Instruction: lecture, Explicit Teaching, Demonstration, Hands on experimental sections, Skill acquisition by laboratory training, Journal Club
Assessment Types	**Mode of Assessment** **Continuous Internal Assessment (CIA)** Assessment of the performance of student in the lab- 10 marks Internal Test -20 marks Project report (student needs to perform experiments on a specific project and report should be prepared)– 10 marks **Semester End examination – 60 marks**

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MAHATMA GANDHI UNIVERSITY

BSM 21C 39 **LABCOURSE-6**

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biophysics**					
Course Name	**LABCOURSE-6**					
Type of Course	Core					
Course Code	BSM 21C 39					
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S**					
Course Summary & Justification	To familiarize the student to various Biophysical and Molecular techniques. To develop laboratory experience on spectrometry, chromatography, electrophoresis, To develops practical skills about Isolation of DNA and PCR amplification					
Semester	Third					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	10	10	120	20	160

Pre-requisite	**Basics of** molecular biophysical methods

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To explain and characterize UV absorption spectrum	E	4
2	To describe the separation and of compounds	U/ An	2
3	To perform purification of compounds by column chromatography and HPLC	R	2
4	To perform the structural elucidation of purified compounds	S	2
5	To perform the DNA isolation and PCR amplification	E	1
6	To perform various electrophoretic techniques	An/ C	2
7	To perform various bioinformatic analysis		3
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Absorption spectra analysis** To study the characteristics of UV absorption spectra of Aromatic Amino Acids To study the characteristics of UV absorption spectra of Proteins To study the characteristics of absorption spectra of Nucleic Acids and Nucleotides	1	30
2	**Chromatography- I** Paper chromatography of Plant Pigments Paper chromatography of aminoacids Paper chromatography of sugars. Thin layer chromatography of aminoacids. HPTLC separation of plant pigments HPTLC of Amino acids & sugars	2,3	30

	To isolate the proteins- Casein from milk and Hb from RBC To analyse the Erythrocytes membrane lipids by TLC Fractionation of Sugars from fruit juice using TLC/HPTLC		
3.	**Chromatography- II** Column Chromatography separation of plant derived compounds. UV Visible spectrum of the separated compounds To perform the separation of column fractions using HPLC GCMS analysis of purified compounds LCMS analysis of purified compounds FTIR analysis of purified compounds	2,4	25
4	**Electrophoresis** Paper Electrophoresis of Amino acids Cellulose acetate strip Electrophoresis of Amino acids Polyacrylamide Gel Electrophoresis (PAGE). SDS- Polyacrylamide Gel Electrophoresis (PAGE) DNA isolation from bacteria and plants. Agarose gel electrophoresis of DNA PCR amplication of 16SrDNA	2,5,6	15
5	**Bioinformatics** Literature databases: PubMed, PMC and PLOS. Nucleic acid sequence databases: NCBI, EMBL and DDBJ. Protein sequence databases: Uniprot and TrEMBL. Protein structure databases: PDB and SCOP. Metabolic pathway databases: KEGG and Reactome. Protein interaction databases: STRING and BioGRID. Homologous sequence search by BLAST and FASTA. Multiple sequence alignment and tree construction	7	20
	Total Credits of the Course	2	120

Teachingand LearningApproach	**Laboratory Procedure (Mode of transaction)** Direct Instruction: lecture, Explicit Teaching, Demonstration, Hands on experimental sections, Skill acquisition by laboratory training
Assessment Types	**Mode of Assessment** **. Continuous Internal Assessment (CIA)** Assessment of the performance of student in the lab- 10 marks Internal Test -20 marks Project report (student needs to perform experiments on a specific project and report should be prepared)– 10 marks **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Biochemical methods by S Sadasivan and A Manickam. New Age international publishers
2. Biotechnology: A laboratory course by Becker J.M.
3. Bioinformatics Practical Manual by Mohammed Iftekhar and Mohammed Rukunuddin Ghalib

Further Reading:

1. An introduction to practical Biochemistry by D. T. Plummer, Mc Graw Hill.
2. Laboratory manual in Biochemistry by J. Jayaraman, Wiley Eastern Limited.
3. Introductory Practical Biochemistry by S. K. Sawhney and Randhir Singh, Narosa
4. Experimental Biochemistry: A student companion by Beedu Sashidhar Rao and Vijay Deshpande, I.K. International Pvt. Ltd., New Delhi.

5. Lab manual in Biochemistry by J. Jayaraman, Wiley Eastern Limited 6. An introduction to practical Biochemistry by D. T. Plummer, Mc Graw Hill. 7. Introductory Practical Biochemistry by S. K. Sawhney and Randhir Singh, Narosa

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BSM 21C 58: BIOPHYSICAL CHEMISTRY

SchoolName	**School of Biosciences**
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**
Course Name	**BIOPHYSICAL CHEMISTRY**
Type of Course	Core
Course Code	**BSM 21C 58**
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S**

Course Summary & Justification	To understand the student about the link between Physics, Chemistry and Biology To perceive the knowledge about ligand binding, models of ligand binding, light scattering, calorimetry, x-ray crystallography The course also provides information about mass spectrometry and nanotechnology					
Semester	Fourth					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	80	20	0	40	140
Pre-requisite	**Basics of** biophysical chemistry methods					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To explain covalent and coordinate bond formation	E	7
2	To describe the process of ligand binding interaction and various models	U/ An	8
3	To narrate the bio-molecular interactions and their role in modulation of biological processes	R	8
4	To describe Isothermal titration and Differential scanning calorimetry in the study of proteins and carbohydrates	S	2
5	To explain the mechanism of X-ray Crystallography	E	4
6	To determine the instrumentation and mechanism of different type of mass spectrometers and to describe applications of nanotechnology.	An/ C	4
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**Atoms and chemical bonds**: Electron theory of valance, Chemical bonding and interaction, synthesis and cleavage of covalent bonds, Reactive species- electrophiles, nucleophiles and radicals, Structure, bonding and special properties of water. Coordinate bond,	1	10

	coordinate bond formation in transition metals Bonding of iron in haemoglobin, cobalt in Vitamin B12, magnesium m in chlorophyll,		
2	**Ligand binding interaction**: Ligand interaction at equilibrium, identical and independent sites model, Scatchard plot, multiple classes of independent sites, interaction between binding sites, Allosterism, MWC model, sequential model, oxygenhaemoglobin binding, binding of two different ligands, cooperative binding, anti-cooperative binding and excluded site binding, energetics & dynamics of binding, binding of immunoglobulin and DNA binding protein. Static and dynamic light scattering, Surface Plasmon resonance, isothermal titration calorimetry and differential scanning calorimetry	2,3	20
3.	**X-Ray Crystallography**-Crystals, Molecular crystal symmetry, Miller indices, reciprocal Lattice, Ewalds Construction, X ray diffraction by crystals, Bragg's Law & Bragg's diffraction equation, laue powder and rotation methods & Laue's equations, diffraction methods-Laue's method, Weissenberg diffraction camera and powder method. Protein X-ray crystallography- production of suitable crystals, Acquisition of the diffraction pattern, Determination of phases, Heavy atom replacement method, Calculation of the electron density and refinement. Phase problem in crystallography. Neutron diffraction, Electron diffraction, Synchrotron diffraction, Fibre diffraction and its application	5	20
4	**Mass spectrometry:** Principles of operation and instrumentation, ion formation and types; molecular ions, meta stable ions, fragmentation processes, fragmentation patterns and fragment characteristics in relation to parent structure and functional groups, mass spectrum; its characteristics, presentation and interpretation. Sector Mass spectrometer, Quadrupole Mass spectrometer, Ion trap Mass spectrometer, Time-of flight Mass spectrometer, Fourier transform Mass spectrometer. Combination with chromatographic methods, Biological application	6	20
5	**Introduction to Nanotechnology:** Orientation and introduction towards Nanotechnology, Classification and nomenclature of Nanomaterials, synthesis and Characterizationof Nanomaterials, Structural and functional principles of Bionanotechnology, protein and DNA based Nanostructures, Nanoparticles in agricultural and food diagnostics, Applications of Nanotechnology	6	10
	Total Credits of the Course	4	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Biophysical chemistry, Principles and Techniques- Himalaya Publ.House
2. X-ray structure determination, George.H. Stout, Lyle H Jensen, John Wiley & Sons.
3. Methods in Modern Biophysics, Bengt nolting, II edition

Further Reading:

1. Biophysical Chemistry, P.R.Bergethon, E.R.Simons
2. Principles of Instrumental Analysis, Skoog, Holler, Nieman

3. Biophysics, VasanthaPattabhi, N.Gautham
4. Biophysical Chemistry, Techniques for the study of biological structure and function, CantorSchimmel part II
5. Bioseparations Science and Engineering: Roger G Harrison, Paul Todd, Scott R. Rudge, Oxford University Press.
6. Methods in Modern Biophysics, Bengt Nolting.
7. Practical Protein crystalloigraphy, Duncan.E, Mc Rec
8. Biomaterials: A Nano Approach,S Ramakrishna, M Ramalingam, T.S. Sampath Kumar, Winston O. Soboyejo,Published by CRC Press
9. Bionanotechnology: Lessons from Nature, D S. Goodsell, by John Wiley & Sons, Inc
10. Klabunde, K.J. (Ed.), "Nanoscale Materials in Chemistry", John Wiley & Sons Inc. 2001

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SchoolName	**School of Biosciences**					
Programme	**M.Sc.Biophysics**					
Course Name	**Laboratory Course 7 Biophysics**					
Type of Course	Core					
Course Code	**BSM 21C 59**					
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S**					
Course Summary & Justification	To familiarize the student to various Biophysical, Physiological, Radiation and Bioinformatics techniques To develop laboratory skills on Separation techniques, Physiological experiments, effects of radiation and Bioinformatics tools					
Semester	Fourth					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	10	20	180	10	230
Pre-requisite	**Basics of** physiological and biophysical methods					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To describe the UV analysis of nucleic acids and proteins	E	PSO2

2	To perform IR spectroscopy and Ultracentrifuge	U/ An	PSO2
3	To perform molecular docking of target versus inhibitors	R	PSO3
4	To isolate RNA and plasmids	S	PSO1
5	To determine spirometry analysis	E	PSO2
6	Molecular Dynamic Simulation	An/ C	PSO3
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Analytical Biophysics** To study the Erythrocytes Membrane Permeability and Transport effects of Hypotonic & Hypertonic shock To determine the partial characteristics of Membrane Protein by SDS-PAGE. To determine the effects of UV on E. coli and elucidation of cell survival curve. Determination of effects of UV on cell membrane	1	30
2	**Spectroscopic analysis** • Determination of effects of UV on cell membrane. • To study the renal stone using Infra-Red (IR) Spectroscopy. • To determine the oil content of oil seeds using Nondestructive IR Spectrophotometry. • Denaturation & Renaturation of DNA. • To isolate the chloroplast and characterize the chloroplast membrane protein To determine the molecular weight of biomolecules using ultracentrifuge	1,2	30
3.	**Molecular Docking** • Identification of Nucleic Acid Binding Proteins Using Nondenaturing Sodium DecylSulfate Polyacrylamide Gel Electrophoresis (SDecS-Page) • Multiple sequence alignment and Conserved Amino acid residues.	3,6	60

	• Analysis and study of sequence using different types of BLAST tool • Molecular docking by using AutoDock4 software		
4	**Molecular Biophysics** • Conformation of Nucleic acid by Spectral study. • To isolate RNA • To isolate and characterize Plasmid DNA. • To hydrolyze the t-RNA and separation of Nucleotides by TLC and paper chromatography • Restriction digestion and agarose gel electrophoresis of DNA • Demonstration of various advanced microscopic techniques	4	30
5	**Physiological Biophysics** • ECG, (demonstration). • Spirometry, Body temperature, pulse sensors, • Breath holding time, Measure the pulse rates, Heart beat rate, BP measurement	5	30
	Total Credits of the Course	3	

TeachingandLearningApproach	**Laboratory Procedure (Mode of transaction)** Direct Instruction: lecture, Explicit Teaching, Demonstration, Hands on experimental sections, Skill acquisition by laboratory training
Assessment Types	**Mode of Assessment** **A. Continuous Internal Assessment (CIA)** Assessment of the performance of student in the lab- 10 marks Internal Test -20 marks Project report (student needs to perform experiments on a specific project and report should be prepared)– 10 marks **B. Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Biochemical methods by S Sadasivan and A Manickam. New Age international

publishers

2. Biotechnology: A laboratory course by Becker J.M.

3. Bioinformatics Practical Manual by Mohammed Iftekhar and Mohammed Rukunuddin Ghalib

Further Reading:

1. An introduction to practical Biochemistry by D. T. Plummer, Mc Graw Hill.

2. Laboratory manual in Biochemistry by J. Jayaraman, Wiley Eastern Limited.

3. Introductory Practical Biochemistry by S. K. Sawhney and Randhir Singh, Narosa

4. Experimental Biochemistry: A student companion by BeeduSashidhar Rao and Vijay Deshpande, I.K. International Pvt. Ltd., New Delhi.

5. Lab manual in Biochemistry by J. Jayaraman, Wiley Eastern Limited

6. An introduction to practical Biochemistry by D. T. Plummer, Mc Graw Hill.

7. Introductory Practical Biochemistry by S. K. Sawhney and Randhir Singh, Narosa

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MAHATMA GANDHI UNIVERSITY

BSM 21E61: QUALITY CONTROL IN HERBAL DRUGS

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**QUALITY CONTROL IN HERBAL DRUGS**					
Type of Course	Elective`					
Course Code	**BSM 21E 61**					
Names of Academic Staff & Qualifications	**Guest faculty – M. Sc (Biochemistry), PhD**					
Course Summary & Justification	The course is designed to get a clear idea on quality control approaches in natural herbs and products and modern analytical techniques for the analysis of the herbal drugs.					
Semester	Fourth					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg. Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding of plant-based drugs					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To estimate the quality assurance of herbal materials.	C	1
2	To isolate, purify and characterize the photochemical from medicinal plants.	A	2
3	To interpret the structure of natural products	U/E	1

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	CO	Hours
1	WHO Guidelines for Quality Control of herbal raw materials. Determination of pesticide residue, arsenic and heavy metals, afflatoxins and microbial contaminants	1	10
2	Definition, principle of the various extraction techniques like maceration, percolation, hot continuous extraction, pilot scale extraction, microwave assisted extraction and supercritical fluid extraction. GMP for the production of quality botanicals.	2	20
3.	General methods for isolation and purification of active principles from medicinal plants. Application of chromatographic techniques in isolation & characterisation of phytochemical constituents viz., paper chromatography, thin layer chromatography, column chromatography, gas chromatography (GC), high performance liquid chromatography (HPLC) and high performance thin layer chromatography(HPTLC).	2,3	10
4	Role of chemical and biological markers in standardization of herbal products	1,3	10
5	General methods for structural elucidation of natural products, Application of spectroscopy for characterization of phytoconstituents	2,3	10
	Total Credits of the Course	3	

Books for Reference

Compulsory Reading:

1. Herbal Drug Technology, S. S. Agrawal, M. Paridhavi, Publisher Universities Press, 2007, ISBN 8173715793, 9788173715792

Further Reading:

2. Pharmaceutical Analysis Hiquchi, Bechmman,Hassan.

3. Methods of Drug Analysis Gearien,Graboski.

4. Text Book of BioPharmaceutic Analysis Robert Smith and JamesStewart.

5. Pharmaceutical Analysis Modern methods Part A and B Munson James.W.

6. Quantitative Analysis of DrugsGarrot.

7. Quantitative Analysis of Drugs in Pharmaceutical Formulations P. D.Sethi.

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** **CC. Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks **DD. Semester End examination – 60 marks**

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MAHATMA GANDHI UNIVERSITY

BSM 21E 62: ENVIRONMENT BIOTECHNOLOGY

School Name	**School of Biosciences**					
Programme	**Msc Biotechnology/ Biochemistry/ Biophysics/ Microbiology**					
Course Name	**BSM 21E 62: ENVIRONMENT BIOTECHNOLOGY**					
Type of Course	Elective					
Course Code	**BSM 21E 62**					
Names of Academic Staff & Qualifications	**Dr.Jayachandran.K**					
Course Summary & Justification	Environmental Biotechnology is offered to train the students both in the theoretical and practical aspects of identifying environmental problem where a solution is possible through Biotechnological methods Enabling students in formulating ideal solution to environment problems based on green chemistry concept is the need of this time . Students have to earn a sense of Environmental concern and to get experience in the applications of Biotechnological methods for environmental protection.					
Semester	Fourth					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisites	None					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**On completing this course, the student will be able to** Understand the effect of a specific environmental problem identified	U	1
2	**Analyse** Apply the most suitable biological method for	An	2

	the effective treatment of the pollutant		
3	**Compare** Explore into the possibility of applying the developed method in the field.	U	1
4	**Acquiring awareness about the emerging challenges in Environmental threats**	S	1
5	**Communicate effectively about a choosen topic of current environmental issue**	An/ C	10
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	Introducti I**ndustrial pollution causes, problems:** Air, Soil and Water pollutants, Types of pollutants characterization, Persistence and Biomagnification of Xenobiotics, recalcitrant molecules, nitroaromatic polychlorinated, biphenyls and dioxans, synthetic polymers, alkylbenzyl sulphonates, Hydrocarbons, Pesticides, Phenolics, Anilines, Inorganic pollutants, Heavy metals. Detection and Quantification of pollutants. Environmental laws	1,5,4	10
2	**Biodegradation, Process and application:** Microbial infallibility, types of biodegradation, factors affecting biodegradation, enzymes involved in biodegradation, catabolic plasmids, Molecular Approaches, Biogeochemical cycles, Bioleaching. Biodegradation of Hydrocarbons, cellulose, lignin, Phenoland pesticides. Application of TOC, FT/IR, GC-MS analysis in biodegradation studies	2,5	10
3.	**Industrial wastewater:** Types of industrial effluents, characterization of the wastewater. Chemical Oxygen Demand, Biological Oxygen Demand, Total organic carbon, Nitrogen contents, Suspended solids. Total heterotrophic bacterial population. Bacteriological analysis of drinking water, Presumptive, completed, and confirmed test. Treatment strategies primary, Secondary and tertiary treatment Physical, Chemical and Biological treatment. Floc based and film based strategies, aerobic and anaerobic methods	1,2,3,5	20
4	**Biological treatment of industrial wastewater:** Activated sludge process, different stages, Types. Oxic/Anoxic, Extended aeration methods, Nitrification and denitrification. Trickling filter process, Different stages Types, Biofilm applications, Rotating Biological contactor,UASB, Submerged aerobic filters, Fluidized Bed Reactor, Packed bed reactor, Oxidation lagoons. Bioreactors for wastewater treatment. Advanced treatment strategies Teritiary treatment methods, Disinfection, Chlorination, Chlorination	4,5	10

	dosage chlorination derived byproducts		
5	**Solid waste management:** Solid waste, Types, Problems, Characterization and sorting of wastes. Municipal and industrial waste management, Land fills composting, stages in composting, Types of composting vermicomposting. Methanogenesis, stages in anaerobic digestion, methanogens Anaerobic reactors Biogas generation, Household treatment strategies, Present problem and Possible remedies	4,5	10
	Total Credits of the Course	3	

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** EE. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks FF. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Microbial Ecoology, Atlas and Bartha, Pearson Publication
2. Comprehensive Biotechnology—2 nd Edition,Murray Moo Young ISBN-9780444533524,Pergman
3. Industrial Microbiology, Samuel Cate Prescot and Cecil Gordan Dunn,Third edition Mac Graw-Hill
4. Waste water microbiology, Gabriel Bitton,Third edition, Wiley, ISBN-

9780471717966 5.
Further Reading: 1. Environmental Biotechnology -Theory and application , Gareth m Evans and Judith C Furlong , Wiley 2003 2. Envoronmental Chemistry-Anilkumae DE,

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MAHATMA GANDHI UNIVERSITY

SBS BSM 21E 63: IPR AND PATENTING

School Name	**School of Biosciences**					
Programme	**Msc Biotechnology/ Biochemistry/ Biophysics/ Microbiology**					
Course Name	**IPR AND PATENTING**					
Type of Course	Elective					
Course Code	**BSM 21E 63**					
Names of Academic Staff & Qualifications	**Dr. Linu Mathew**					
Course Summary & Justification	To introduce students the concept of intellectual property and IPR					
Semester	Fourth					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisites	None					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**On completing this course, the student will be able to** **Define different international agreement on IPR**	u	1
2	**Analyse the patentability of an invention and laws on plant variety protection**	An	1
3	**Compare the patentability of biological entities**	U	1

4	**File a patent**	S	1
5	**Communicate effectively about a patent related topic both verbally and in writing**	An/ C	10

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Introducti Introduction. Definitions General Agreement on Trade and Tariff (GATT) and World Trade Organizations Establishment and functions of GATT, WTO, and WIPO. WTO Guidelines and Summits. Physical and Intellectual Property	1,5	10
2	TRIPS Different types of intellectual property rights (IPR) - Patents, Trade mark, Trade secret, copyright and Geographical indications Requirement of patentability, Biotechnological examples of patents, trademark, trade secret and copy right	1,5	10
3.	Patenting research tools and the law: Patents as a Strategy for Protection of Intellectual Property, Benefits and Costs of Patents, Requirements for Patent Protection, patentable subjects and protection in biotechnology, international convention for the protection of new varieties – Strasbourg convention, UPOV convention. Experimental Use Exemption	2,5	10
4	Patent filing and Infringement Patent application- forms and guidelines, fee structure, time frames; Types of patent applications: provisional and complete specifications; PCT and convention patent applications; International patenting-requirement, procedures, and costs; financial assistance for patenting-introduction to existing schemes; Indian Patent Act, 1970 and recent amendments Publication of patents in India Status of patenting in Europe and US. Patenting by research students, lecturers, and scientists University/organizational rules in India and abroad, credit sharing by workers, financial incentives, Patent infringement- meaning, scope, litigation, case studies and examples	4,5	20
5	The patentability of microorganisms, legal protection for plants and other higher organisms, new plant varieties by rights, tissue culture protocols, transfer of technology. Patentability of vectors. Licensing - Flavr Savr™ tomato as a model case, Biopiracy and case studies on patents (Basmati rice, Turmeric, and Neem)	3,5	10
	Total Credits of the Course	3	

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** GG. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks HH. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

6. Patents (2003), N.Subbaram, Pharma Book Syndicate, Hyderabad.
7. WIPO Hand book on Intellectual Property
8. IPR , Biosafety, and Bioethics Deepa Goel and Shomoni Parashar

Further Reading:

1. Revised guidelines for research in Transgenic plants (August 1998), Department of Biotechnology, Ministry of Science & Technology, Government of India, New Delhi.
2. Intellectual Property, W.R. Cornish, Sweet and Maxwell publishers, London

Web resources

3. https:// worldwide. espacenet.com
4. https:// patentscope. wipo. int
5. https:// ipindiaservices.gov.in

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MAHATMA GANDHI UNIVERSITY

BSM 21E 64: OMICS IN BIOTECHNOLGY

School Name	**School of Biosciences**					
Programme	**MSc Biotechnology**					
Course Name	**Omics in Biotechnology**					
Type of Course	**Elective**					
Course Code	**BSM 21E 64**					
Names of Academic Staff & Qualifications	**Dr. Keerthi TR**					
Course Summary & Justification	1. The course describes new approach, the concept of "OMICS" in various levels.It is a multi-disciplinary emerging field that encompasses genomics, epigenomics, transcriptomics, proteomics, and metabolomics. 2. The course content explain the high-quality techniques, methods & analysis from genome level will help in the complete understanding of a biological process.These approaches are targeted towards understanding complex systems more thoroughly at the molecular level.					
Semester	Fourth					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours

	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisites	Basics of Molecular Biology					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**On completing this course, the student will be able to Explain genome and types of genomics, tool and methods in genomic study, as well as Genome structure of selected organisms.**	**U/E**	1
2.	**Explain the Proteomics, Transcriptomics &Metabolomics & Describe the tool and methods employed to study. Students have able to explain the various application of Proteomics, Transcriptomics & Metabolomics study**	**An/A**	**1**
3.	**Students have able to illustrate the techniques employed for metagenomic analysis and application of metagenomic study**	**S/I**	**2**
4.	**Describe the classification and types of databases & applications of data bases**	**U/R**	**3**
5.	**Communicate effectively about a chosen topic in Omics in Biotechnology both practically and theoratically.**	**C/S**	**1**
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**Genome & Genomics**: Definition of Genome & Genomics.Types of genomics,, Functional Genomics.Structural genomics&Comparative genomics, Tools in Genomics,Structural genomics:-Classical ways of genome analysis, large fragment genomic libraries; Physical & Genetic mapping of genomes; Genome sequencing, sequence assembly, annotation& bioinformatics.Functional genomics:-DNA chips and their use in transcriptome	**1,5**	**20**

	analysis; Mutants and RNAi in functional genomicsNext generation sequencing methods; Structure of genomes: bacteria, yeast, nematode, Arabidopsis, rice, zebra fish, mouse and man.Applications of genomics		
2.	**Proteomics,Transcriptomics & Metabolomics:** Basic concepts , Introduction to transcriptomics,proteomics and metabolomicsTools of proteomics- SDS PAGE, 2D PAGE , Liquid chromatography , Mass Spectrometry (ESI and MALDI) ,Protein identification by peptide mass fingerprinting ,Applications of proteomics-. Protein identity based on composition, Motifs and patterns, Analysis and characterization of proteins and metabolites:. Proteomics approaches to the analysis of protein-protein interactions, and metabolic profiling through emerging metabolomic techniques like 2D gel electrophoresis and Mass spectrometric and computational techniques.Applications of proteomics in agriculture, human health and industry	**2,5**	**10**
3.	**Metagenomics:** Definition of metagenomics**,**Techniques in metagenomics-Isolating DNA from an environmental sampleClone DNA,Insert into plasmid,Develop sample library,Screen or sequence,Analysis of metagenomic data.Application of metagenomics	**3,5**	**10**
4	**Biological data bases:** Classification databases. Biological databases-primary sequence databases- Composite sequence databases- Secondary databases-composite protein pattern databases,Pattern and profile databases Genome Information Resources: DNA sequence databases-specialized genomic resources, GRAIL, GENSCANProteome databases Protein sequence databases - SWISS-PROT and TrEMBL — PROSITE and BLOCKS - 2D PAGE databases – Structure databases - PDB- Metabolic databases – post translational modification databases	**4,5**	**20**
	Total Credits of the Course	**3**	

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** II. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and

	literature search – 10 marks JJ. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Introduction to proteomics, Daniel. C. Libeler, Humana Press 2002
2. Thompson, J.D., Schaeffer-Reiss, C., and Ueffing, M. 2008. *Functional Proteomics. Methods andProtocols*. Humana Press, New York.
3. Metabolomics- Methods and Protocols by Wolfram Weckwerth, Humana Press.
4. Aurthur M Lesk Introduction to Bioinformatics .Oxford University press.

Further Reading:

1. Bostjan Koba.,Mitchell Guss & Thomas Habs Structural Proteomics.Humana Press.
2. Twyman, R.M. 2004. *Principles of Proteomics*. Taylor & Francis
3. Mass Spectrometry for Biotechnology by Gary Siuzdak, Academic Press.
4. Proteomics for Biological Discovery by Timothy Veenstra and John Yates, Wiley.
5. Lipidomics- Technologies and Applications by Kim Ekroos, Wiley-VCH.
6. Web/Journal Resources.
7. Transcriptomics: Expression Pattern Analysis, Virendra Gomase, Somnath Tagore; VDM Publishing, 2009 – Science
8. Brown TA. 2007. Genome III. Garland Science Publ.
9. Campbell AM & Heyer L. 2004. Discovery Genomics, Proteomics and Bioinformatics. Pearson Education.
10. Jollès P & Jörnvall H. 2000. Proteomics in Functional Genomics: Protein Structure Analysis.
11. Kamp RM. 2004. Methods in Proteome and Protein Analysis. Springer.
12. Primrose SB & Twyman RM. 2007. Principles of Genome Analysis and Genomics
13. Blackwell. Sensen CW. 2005. Handbook of Genome Research. Vols. I, II. Wiley CVH.

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MAHATMA GANDHI UNIVERSITY

BSM 21E 65: MOLECULAR PHYLOGENY

School Name	**School of Biosciences**
Programme	**Msc Biotechnology/ Biochemistry/ Biophysics/ Microbiology**
Course Name	**MOLECULAR PHYLOGENY**
Type of Course	Elective
Course Code	**BSM 21E 65**
Names of Academic Staff & Qualifications	**Dr. Linu Mathew**
Course Summary & Justification	1. This elective course deals with the tools and techniques of Molecular phylogeny. The course has a theoretical and a practical

	dimension 2. The learner will develop an understanding about models of nucleic acid substitution, tree building algorithms, data mining tools and submission tools for nucleic acid data and applications of Molecular phylogeny					
Semester	Fourth					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisites	Basics of genome organisation and organic evolution, concepts of biological classification					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**On completing this course, the students will be able to** **Compare and narrate the models of nucleic acid substitution, tree building algorithms, data mining tools, and submission tools for nucleic acid data**	An	3
2	**Deposit nucleic acid sequences in databases and able to perform data mining**	S	3
3	**Perform sequence alignment and editing**	S	3
4	**Analyse sequence alignments by suitable software and perform phylogenetic analysis**	S	3
5	**Carry out a phylogenetic analysis from raw sequence data up to final conclusions**	S	3
6	**Communicate effectively about a phylogenetic problem both verbally and in writing.**	An/ C	10
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs

1	Basic concepts of molecular evolution: Genetic information, population dynamics, evolution and speciation, data used for molecular phylogenetics, phylogenetic tree, methods for inferring phylogenetic trees, networking, RNA world	1,6	15
2	Sequence databases and data base searches: Sequence databases, composite databases, database mirroring, and search tools, data base searching by sequence similarity – BLAST and FASTA, multiple sequence alignments CLUSTAL, MUSCLE, T-COFFEE	2,3,6	10
3.	Phylogenetic inference: Genetic distances and nuclear substitution models, phylogenetic inference based on distance methods- UPGMA, Neighbour Joining, Minimum Evolution, Least square	4,5,6	10
4	Phylogenetic inference: Maximum Likelihood and Bayesian phylogenetic analysis, phylogenetic analysis based on parsimony, phylogenetic analysis using protein sequences, testing tree reliability – Bootstrapping and jackknifing	4,5,6	10
5	Testing models and trees: Models of evolution and phylogeny reconstruction, model fit, likelihood ratio tests, Practising MEGA, Paup*, RaxML, Mr Bayes, J Model Test, Sequence submission tools- SEQUIN and BankIt	4,5,6	15
	Total Credits of the Course	3	

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** KK. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks LL. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Molecular evolution And Phylogenetics, Masatoshi Nei and Sudhir Kumar, Oxford University Press, ISBN 0195135857
2. Baldauf, SL (2003) "Phylogeny for the faint of heart: a tutorial." Trends in Genetics; 19(6):345-351.

Further Reading:

3. The phylogenetic Hand book, 2nd Edition, Philippe Lemey, Marco Salemi, Anne – Mieke Vandamme, Cambridge University Press, ISBN-13 978-0-511-71963-9
4. Hall, BG. (2004) Phylogenetic Trees Made Easy: A How-To Manual, 2nded. Sinauer Associates, Inc.: Sunderland, M A. I*SBN: 978-0-87893-606-9*
5. Hartwell, LH, L Hood, ML Goldberg, AE Reynolds, LM Silver, RC Veres (2008) Genetics: From Genes to Genomes, 3rd Ed. McGraw-Hill: New York ISBN-13: 978-0073525266ISBN-10: 007352526X

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MAHATMA GANDHI UNIVERSITY

BSM 21E 66: HUMAN VIROLOGY

SchoolName	**School of Biosciences**
Programme	**M.Sc Microbiology/Biochemistry/Biotechnology/Biophysics**
Course Name	**HUMAN VIROLOGY**
Type of Course	Core

Course Code	**BSM 21E 66**					
Names of Academic Staff & Qualifications	**Dr. Jyothis Mathew M.Sc, Ph.D**					
Course Summary & Justification	This course on Human Virology deals with an important area of Medical Microbiology The objective of the course content is to create a sound awareness in human viruses and viral diseases. their The course will augment the student's knowledge in pathogenesis of viral diseases and their laboratory diagnosis and prophylaxis.					
Semester	Fourth					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding on Human Anatomy, Physiology and Biochemistry Knowledge in Basic Virology, Molecular Biology and Immunology					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1.	On completing this course student will be able to analyse comparatively the structure and properties of important human viruses	**U/An**	**1**
2.	Students will be able to understand and evaluate the mechanism of pathogenesis of viral diseases	**U/E**	**1**
3.	Students will become aware of the methods applicable in viral diagnostics	**U/A**	**1**
4.	Students will be able to analyse the various mechanisms of viral oncogenesis	An	1
5,	Students will be able to understand and compare the mechanisms of action of various antiviral agents	U/An	1
6	Students will be able to understand and evaluate the methods of prophylaxis of viral diseases in humans	U/E	1
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I)			

and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Study of properties of human DNA viruses viz. Pox, Herpes, Adeno, Papova, and Parvo viruses. Pathogenesis and laboratory diagnosis of diseases caused by these viruses	1,2,3	
2	Study of properties of human RNA viruses viz. Picorna, Orthomyxo, Paramyxo, Rhabdo, and Rubella viruses	1,2,3	
3.	Arboviruses and Hepatitis viruses - Properties. Pathogenesis and laboratory diagnosis of diseases caused by these viruses. Viral haemorrhagic fevers, SARS CoV-2, HIV,Properties, pathogenesis and laboratory diagnosis of Slow virus infections, Prion diseases	1,2,3	
4	Viruses and cancer, Viral oncogenesis, Viruses implicated in the cancers of humans, Prophylaxis of viral diseases, Types of viral vaccines, antiviral agents and their mechanisms of action, Interferons	4,5,6	
	Total Credits of the Course	3	

TeachingandLearning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** MM. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks NN. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Jawetz,Melnick& Adelberg's Medical Microbiology27 th Edition Carrol, Butel, Morse, Mietzner Mc Graw Hill

2. Ananthanarayan & Panicker's Text book of Microbiology.9th Edition Arti Kapil (Ed) University Press (India) Pvt.Ltd.

Further Reading:

Further Reading:

1. Human Virology Fourth Edition Leslie Collier, John Oxford & Paul Kellam University Press.

2. Fundamental Virology 5th Edition David M.Knipe& Lippincott Williams & Wilkins

3. Viruses Biology,Applications & Control

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MAHATMA GANDHI UNIVERSITY

BSM 21E 67ADVANCED TECHNIQUES IN DIAGNOSTIC MICROBIOLOGY

SchoolName	School of Biosciences
Programme	M.Sc. Microbiology/Biotechnology/Biochemistry/Biophysics

Course Name	ADVANCED TECHNIQUES IN DIAGNOSTIC MICROBIOLOGY					
Type of Course	Elective					
Course Code	**BSM 21E 67**					
Names of Academic Staff & Qualifications	Dr.Radhakrishnan E.K., M.Sc., Ph.D					
Course Summary & Justification	Different methods are used to detect the diseases caused by microorganisms. The syllabus content in this course has been designed with an objective to provide the basic principle and applications of various methods used in diagnostic microbiology. This will enable the students to learn the basic and advanced methods in diagnostic microbiology which will enable them to identify the research and job opportunities based on the latest developments in this subject					
Semester	Fourth					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding on diseases caused by microorganisms, different methods used to detect the diseases					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1.	Students will able to understand the process and methods in medical microbiology lab	R/U	1
2.	Students will able to understand various clinical samples used for diagnostic applications	R/U	1
3.	Students will able to explain the principles of methods used in medical microbiology	U/ An/E	1
4.	Students will get exposed to both the conventional and rapid methods used for the microbial identification	U/An/A	1
5.	Students will able to identify research and job opportunities in diagnostic microbiology	C/S	10
6.	Students will able to analyze scope of technological advancement for rapid microbial identification	S/I	10
**Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*			

COURSE CONTENT

Module No	**Module Content**	**CO**	**Hrs**
1	Introduction to diagnostic microbiology, laboratory safety, hospital epidemiology. Lab methods in Medical Microbiology, basic virology, basic mycology, Clinical material - collection and transport. Etiological agents recovered from different clinical materials	1,2,3	20
2	Biochemical profile based microbial identification systems, Urea breath test, Rapid antigen tests, Enzyme-Linked Immunoassay, Western blot, Advanced antibody detection, Bacterial antimicrobial susceptibility tests	1,4	20
3.	Polymerase chain reaction, Principle, applications and types of PCR in medical diagnostic field, Microbial Identification Based on PCR amplification of 16S rDNA, Sequence analysis, Application of Real	4,5	10

	Time PCR in Diagnostic Microbiology, Microbial Strain Typing Using Repetitive Sequences Advances in the Diagnosis of *Mycobacterium tuberculosis* and methicillin resistant *Staphylococcus aureus.*		
4	Probe-Based Microbial Detection and Identification, Southern Blot Hybridization, Microarray- Based Microbial Identification and Characterization, Recent advances in medical microbiology	5,6	10
	Total Credits of the Course	3	

REFERENCES

Teaching And Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction, Active co-operative learning, Seminar, Group Assignments Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** OO. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar - Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks PP. Semester End examination – 60 marks

Compulsory Reading:

1. Bailey and Scott's Diagnostic Microbiology Publisher: Elsevier Health, 28 Jun 2013
2. Advanced Techniques in Diagnostic Microbiology Editors: Wu, Shangwei, Stratton, Charles, 2012

Further Reading:

3.Textbook of Diagnostic Microbiology Hardcover, by Mahon (Author), Publisher:

Elsevier Health - US; 5 edition (18 February 2014) 4.Koneman's Color Atlas and Textbook of Diagnostic Microbiology 7th Edition by Gary W. Procop MD MS , Elmer W. Koneman, Publisher: LWW; 7 edition (June 15, 2016)

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MAHATMA GANDHI UNIVERSITY

BSM 21E 68: RADIATION BIOPHYSICS

SchoolName	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**RADIATION BIOPHYSICS**					
Type of Course	Elective					
Course Code	**BSM 21E 68**					
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S**					
Course Summary & Justification	To introduce the student to an important division of Biophysics- Radiation Biophysics To familiarize the topics of Radiation and Radioactivity, its interactions, biological effects, dosimetry, hazards, protection and application in medicine, industry and agriculture The course is designed to provide an overview of different imaging techniques used in medical field					
Semester	Fourth					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120

Pre-requisite	**Basics of** Radiation biophysics

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To describe various kinds of radiation and radiation units	E	6
2	To explain the various biological effects of radiation	U/ An	6
3	To narrate how to detect and measure radiation	R	6
4	To explain how to protect from radiation exposure	S	6
5	To describe the use of radioisotopes in medicine, industry and agriculture	E	6
6	To discuss about the biomedical imaging techniques	An/ C	2
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	**Radioactivity:** Laws of radioactivity, α, β, γ rays. Properties of electromagnetic radiation. Radiation units; Exposure and Dose, Dose equivalent unit, KERMA, Absorbed dose and Derived Units- Equivalent Dose and Effective dose, Dose rate. Interaction of radiation with matter- Bremsstrahlung, Photoelectric effect, Compton effect, Ion pair production. Interaction, absorption and scattering of electron. Heavy charged particles and Neutrons. Attenuation coefficient and absorption coefficient. HVL, Mean free path, Absorption edges, LET, Relative biological effectiveness (RBE)	1	10
2	**Biological effects of radiation:** Radiolysis of water, Production of free radicals & their interactions, Competition kinetics, Diffusion kinetics & Physicochemical effects, Role of scavengers, G-value, Genetic Effect of radiolysis, Chromosomal breakage and Aberrations Direct and Indirect action, Oxygen and temperature effect, OER. Target theory, Single hit & Multi hit theory, Multi target theory, Calculation of target, Mass, Volume & Molecular weight, Effect of radiation on Nucleic acids, Proteins, Enzymes &	1,2	10

	Carbohydrates, Somatic and genetic effects of radiation, Stochastic and deterministic effects, early and late effects, Radiation sickness, Radiation syndrome, Haemopoietic syndrome, G.I syndrome, CNS syndrome, Acute radiation damage, Early and late effects of radiation, Effect of chronic exposure to radiation. Acute radiation damage, LD-50, Dose effect relationship. Cell recovery and modification of Radiation damage		
3.	**Radiation dosimetry**: Principles of radiation detection and measurement- Dosimetry- General requirements of Dosimeters, Radiation sources, Telegamma Unit (Cobalt unit), Gamma chamber, Nuclear reactors, Thermal & fast neutron sources. Dosimeters- Basic principles, Design & Working of physical dosimeters- Ionization chamber, Proportional counters, GM- Counter, Concepts of Gas amplification, Resolving time & Dead time, Scintillation Detectors, Thermolumeniscent Dosimeter, Semiconductor, Surface barrier & Lithium detectors, Area survey meter & Pocket dosimeter, Film badge, General principle of chemical dosimetry, Salient Features of Chemical dosimeter, Dose evaluation formula for chemical dosimetry, Principles of radiolytic reaction, Experimental methods- Influencing factors of Fricke dosimeter methyl orange, FBX dosimeter, Free radical dosimeter, Ceric sulphate dosimeter, PMMA, PVC, chlorobenzene dosimeter, High & low dose indicators	3	20
4	**Radiation Hazards and Protection**: Natural and man-made radiation exposures, maximum possible dose, Radiation hazards- external and internal radiation hazards. Radiation protection measurement in industrial establishment, Radioisotope labs, diagnostic and therapeutic installation and during the transportation of radioactive substances, Disposal of radioactive wastes.	4	10
5	**Applications of radiation**- Radioisotopes in Biology, Agriculture, Plant breeding, Plant Physiology, Medicine. Internally administered isotopes. Radioiodine in thyroid function analysis. Renal, liver and lung function analysis. Radio Immuno Assay, Radiotracer techniques. Auto radiography. Specialized radio isotopic applications in industries **Biomedical imaging techniques**- Principle of analogue and digital imaging, Ultra sound imaging, Nuclear resonance imaging, X-ray imaging and CT scan, Principle of tomographic techniques, Computerised tomography, positron emission tomography,	5,6	10

	application and interpretation of image		
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** QQ. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks RR. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Glenn.F.knoll., Radiation detection and Measurement; III Edition,John Wiley & Sons, Inc.
2. Edward L. Alphen., Radiation Biophysics©, Prentice Hall

Further Reading:

8. Frank.H. Attix., Introduction to Radiological Physics & Radiation dosimetry
9. Wagner, Szabo, Buchanan., Principles of Nuclear medicine.
10. Orton, C.G., Radiation Dosimetry: Physical and Biological aspects.
11. Girish Lahari- Nuclear Physics,Mohit Books International.
12. S.P.Yarmonenko;Radiobiology, Mir Publishers.

13. JozsefKonya.Noemi M. Nagy; Nuclear and Radiochemistry,Elsevier insight

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MAHATMA GANDHI UNIVERSITY

BSM 21E 69: GOOD LABORATORY PRACTICES

SchoolName	**School of Biosciences**					
Programme	**MSc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**GOOD LABORATORY PRACTICES**					
Type of Course	Elective					
Course Code	**BSM 21E 69**					
Names of Academic Staff & Qualifications	**Dr. R. Harikumaran Nair** MSc, PhD					
Course Summary & Justification	To equip the students with appropriate knowledge, skills to undertake general and quality management of laboratory practices and procedures. To adequately address quality issues and improve the overall delivery of clinical and public health laboratory services in their facilities/organizations. To sensitize the students with medical and public health ethics issues and to ensure its application in teaching and practice.					
Semester	Fourth					
Total StudentLearning	Learning Approach	Lecture	Tutorial	Practical	Others	Total

Time (SLT)						LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics Knowledge in Biosciences**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Understand basic good laboratory practice	U	1
2	Appreciate how to conduct research safely and efficiently	Ap	1
3	Understand the requirements for safe working practices and risk assessment	U	1
4	Apply experimental design and the need for controls	A	1
5	Consider ways in which student can maximise research effort	C	10
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	Introduction to good laboratory practices (GLP) and its application, history of GLP, fundamental points of GLP	1	10
2	Resources-personnel, Facilities - buildings and equipment, Characterization- test item, test system, rules for performing studies-the study plan or protocol, standard operating procedures (SOPs) raw data and data collection- records and recording, study report, archives and archiving, quality assurance, audit and	2	20

	inspections, implementation of GLP		
3.	Applications of the GLP principles to field studies, applications of the GLP principles to short term studies, applications of the GLP principles to in vitro studies	3	10
4	Ethics in research- locating ethics in research, justice in research, science and society, ethical issues in biotechnology, ethical guidelines related to human experimentation, guidelines regarding animal use in research, institutional biosafety monitoring mechanisms.	4,5	20
	Total Credits of the Course	3	60

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** SS. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks TT. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading

1. Handbook on Good Laboratory Practice- World Health Organization
2. Ethical Guidelines for Biomedical Research on Human Participants- Indian Council of Medical Research
3. Guidelines on the regulation of scientific experiments on animals- Ministry of Environment and Forests, India
4. Textbook on Ethics in Research- European Commission, Publications Office of the European Union

Further Reading:

1. **Good Laboratory Practice Regulations, 4th edition edited By Sandy Weinberg- CRC**

Press, 2007

2. **The Indispensable Guide to Good Laboratory Practice (GLP): Second Edition 2nd Edition- Mark Gregory Slomiany- Springer, 2009**

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MAHATMA GANDHI UNIVERSITY

BSM 21E 70: HEALTH AND NUTRITION

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**HEALTH AND NUTRITION**					
Type of Course	Elective					
Course Code	**BSM 21E 70**					
Names of Academic Staff & Qualifications	**Dr. Anie Y – M. Sc (Biochemistry), PhD**					
Course Summary & Justification	The course is designed to provide basic information on nutrition and its importance in providing health.					
Semester	Four					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg. Authentic learning Collaborative learning Independent learning	60	30	0	40	130
Pre-requisite	Basic understanding of food and food ingredients					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To describe the basic principles of nutritional biochemistry and different methods of nutritional analysis.	R/U	1
2	To identify and compare the different ingredients and nutritional value of food components	A	1
3	To identify different diseases associated with nutritional deficiency and overnutrition	U	1
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	CO	Hours
1	**Introduction to nutrition -** Food as source of nutrients, functions of food, definition of nutrition, nutrients & energy, adequate, optimum & good nutrition, malnutrition. Basics of energy metabolism, nutrition & dietetics - Unit of measuring energy, calorific value of food, BMR & factors affecting it, SDA of food, calculation of energy requirement, balanced diet, nutrition in health & disease. Nutritional disorders- Epidemiology, clinical features, prevention and dietary treatment for Protein Energy malnutrition, nutritional anaemias.	1,3	20
2	**Food sources:** Carbohydrates : Functions, classification, food sources, storage in body. Fats & oils : composition, saturated and unsaturated fatty acids, classification, food sources, function of fats. Proteins - composition, sources, essential & non-essential amino acids, functions, Protein deficiency	2	10
3.	**Water, Vitamins and minerals-** Water - as a nutrient, function, sources, requirement, water balance & effect. Minerals - macro & micronutrients. - functions, sources. Bioavailability and deficiency of Calcium, Iron, Iodine, Sodium & Potassium (very briefly). Vitamins (water & fat soluble) - definition, classification & functions. Effect of cooking & heat processing on the nutritive value of foods. Processed supplementary foods.	2	10

4	**Nutritional problems affecting the community-**Etiology, prevalence, clinical features and preventive strategies of- Undernutrition - Protein energy malnutrition: Nutritional Anaemias, Vitamin A Deficiency, Iodine Deficiency Disorders. Overnutrition – obesity, coronary heart disease, diabetes. Fluorosis	3	20
	Total Credits of the Course	3	

Books for Reference

Compulsory Reading:

1. Mudambi, SR and Rajagopal, MV. Fundame ntals of Foods, Nutrition and Diet Therapy; Fifth Ed; 2012; New Age International Publishers
2. Mudambi, SR, Rao SM and Rajagopal, MV . Food Science; Second Ed; 2006; New Age Publ.

Further Reading:

1. Srilakshmi B. Nutrition Science; 2012; New Age International (P) Ltd.
2. Swaminathan M. Handbook of Foods and Nu trition; Fifth Ed; 1986; BAPPCO.
3. Bamji MS, Krishnaswamy K and Brahmam GNV (Eds) (2009). Textbook of Human Nutrition, 3rd edition. Oxford and IBH Publishing Co. Pvt. Ltd. New Delhi.

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** **UU. Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks **VV. Semester End examination – 60 marks**

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MAHATMA GANDHI UNIVERSITY

BSM 21E 71: NEUTROPHIL BIOLOGY

SchoolName	**School of Biosciences**
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**
Course Name	**NEUTROPHIL BIOLOGY**
Type of Course	Elective
Course Code	**BSM 21E 71**
Names of Academic Staff & Qualifications	**Dr. Anie Y – M. Sc (Biochemistry), PhD**
Course Summary & Justification	The course is designed to get a detailed idea about the functioning of neutrophils in providing immune response and the mechanisms behind it. This would be helpful for the students, in case they take up research in immunology, cell biology or cellular biochemistry.
Semester	Four

Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg. Authentic learning Collaborative learning Independent learning	60	20	10	40	130
Pre-requisite	Basic understanding of immunology and blood cells					

COURSE OUTCOMES (CO)

CO No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**
1	To describe the role of neutrophils in imparting and fine-tuning immune response	R/U	1
2	To identify and compare different functions of neutrophils	U/A	1
3	To identify different techniques to perform neutrophil functional analysis	S	1
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	**Module Content**	**CO**	**Hours**
1	**Introduction to immune system-** innate and adaptive immune system, cells involved in immune system, humoral immunity, cytokines, antibodies, complement system. cell- mediated and humoral immune response	1	10
2	**Neutrophil Physiology**-Neutrophil structure, Granule types- azurophilic, specific, gelatinase, secretory vesicles, Antimicrobial peptides. Neutrophil Subpopulations. Neutrophil activation, apoptosis and clearance. Neutrophils in the resolution of inflammation. Neutrophil in immune cross-talk	2	10
3.	**Neutrophil defense mechanisms-** Chemotaxis, Phagocytosis, degranulation, ROS generation,NADPH oxidase, Neutrophil extracellular trap formation, NETosis vs. apoptosis and necrosis, Cytokine secretion. Diseases associated with altered neutrophil defence- Autoimmunity, cancers, thrombosis.	2	20

4	**Techniques to study neutrophils:** Neutrophil isolation and maintenance, Cell counting, Phagocytic assays, chemotactic assays, NBT assay, MTT assay, other assays of ROS production, Granule isolation, Neutrophil protein analysis, microscopic analysis of neutrophils and granules – Light and fluorescent microscopy, SEM and TEM	3	20
	Total Credits of the Course	3	

Books for Reference

Compulsory Reading:

1. Neutrophil Methods and Protocols, Quinn, Mark T., DeLeo, Frank R., Bokoch, Gary M. (Eds.). ISBN 978-1-59745-467-4.
2. Biochemistry and physiology of the neutrophil, Steven W Edwards, Cambridge university press Online ISBN-9780511608421
3. The Neutrophil, Murphy, Patrick , Springer, ISBN- ISBN 978-1-4684-7418-3

Further Reading:

1. Neutrophil function: Mechanisms to diseases. Borko Amulic, Christel Cazalet,Garret L. Hayes, Kathleen D. Metzlerand Arturo Zychlinsky; Annu. Rev. Immunol. 2012. 30:459–89.
2. Neutrophil biology: an update. Yoshiro Kobayashi, EXCLI J. 2015; 14: 220–227. doi: 10.17179/excli2015-102.
3. Advances in neutrophil biology: clinical implications. Cowburn AS, Condliffe AM, Farahi N, Summers C, Chilvers ER. Chest. 2008 Sep;134(3):606-12. doi: 10.1378/chest.08-0422.
4. The Neutrophils: New Outlook for Old Cells. 3rd Edition.Edited by: Dmitry Gabrilovich (H Lee Moffitt Cancer Center, USA & University of South Florida, USA). ISBN: 978-1-84816-836-7

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, demonstrations, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** **A. Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks **B. Semester End examination – 60 marks**

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MAHATMA GANDHI UNIVERSITY

BSM 21E 72:PLANT-MICROBE INTERACTIONS

SchoolName	**School of Biosciences**
Programme	M.Sc. Microbiology
Course Name	**PLANT-MICROBE INTERACTIONS**
Type of Course	Elective
Course Code	**BSM 21E 72**
Names of Academic Staff &	**Dr.JISHA.M.S**

Qualifications						
Course Summary & Justification	This course develops concepts in plant- microbe interaction The major objective of this paper is to give an insight into the consequences, on population and ecosystem level, of compatible and incompatible interactions, to understand infection process and control measures and to familiarize with the microbial production of plant metabolites.					
Semester	First					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of agricultural microbiology**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Comprehensively discuss interactions between plants and microbes as well as the defense reactions of the host plant	U/R/ An	1
2	Gain insight into genetics of host-pathogen interactions and resistance mechanism in plants.	C/ I/An	1
3	Comprehend various methods to analyse plant diseases and biological methods of disease control	S/An/A	1
4	Analyse why plants and microbes react in certain ways in pathogenic and symbiotic interactions	U/R/An	1
5	Understands the role of microbes in developing plant immunity	U/R	1
6	Have an in-depth knowledge on biopesticides and their role in pest control	An/ C	1
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I)			

and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	CO	Hrs
1	Different interfaces of interactions -soil-plant-microbe interactions leading to symbiotic (rhizobial and mycorrhizal), associative, endophytic and pathogenic interactions	1,2	10
2	General concepts of plant immunity. PAMP-triggered immunity (PTI) and Effector triggered immunity (ETI). Outer membrane vesicles (OMVs) and their involvement in plant immunity. The type III secretion system. Hypersensitive response. Genetic basis of plant defences. Quorum-sensing in bacteria and their role in plant defence mechanisms. Phytohormones and antibiotics as plant therapeutics.	2,3,4	20
3.	Plant pathogens and molecular basis of pathogenesis .Genetics of host-pathogen interactions, resistance genes, resistance mechanisms in plants. basal and induced defence mechanisms. Systemic Acquired Resistance (SAR) and Induced Systemic Resistance (ISR), Recognition mechanism and signal transduction during plant - pathogen interaction. Virulence determinants of plant pathogenic bacteria-Enzymes, Toxins, pili, siderophores, secretion systems	4,5	20
4	Microbial pest control: Bacillus thuringiensis-mode of action, Biocontrol agents– uses and practical constraints Biofungicide and bioherbicides. Plant growth promoting rhizobacteria. Use of plant–microbe symbiosis for remediation of pollutants and carbon (C) sequestration	6	10
	Total Credits of the Course	3	

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction: Active co-operative learning, Seminar, Group Assignments Authentic learning: Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** WW. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to

	prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks XX. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Subba Rao, N.S. 2005. Soil Microorganisms and Plant Growth, Oxford and IBH Publishing Co.
2. B. Lugtenberg (ed). 2015.Principles of plant microbe interactions, Springer

Further Reading:

1. Microbial control and pest Management – S.Jayaraj.
2. Paul, E.A. 2007. Soil Microbiology, Ecology and Biochemistry, Academic Press.
3. M.Gillings and Holmes .2004.Plant microbiology-Bios Scientific publishers.
4. Kosuge T & Nester EW. 1989. Plant-Microbe Interactions: Molecular and Genetic Perspectives .Vols I-IV. McGraw Hill.
5. Verma DPS & Kohn TH. 1984. Genes Involved in Microbe-Plant Interactions. Springer Verlag.
6. Gary Stacey, Noel T. Keen, 1995. Plant-Microbe Interactions. Vols I-VI Springer Science & Business Media.
7. Jeng-Sheng Huang **2001.**Plant Pathogenesis and ResistanceBiochemistry and Physiologyof Plant-Microbe Interactions .Springer Verlag

Approval Date	
Version	
Approval by	
Implementation Date	

MAHATMA GANDHI UNIVERSITY

BSM 21E 73: SUSTAINABLE AGRICULTURE

School Name	**School of Biosciences**

Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**SUSTAINABLE AGRICULTURE**					
Type of Course	Elective					
Course Code	**BSM 21E 73**					
Names of Academic Staff & Qualifications	**Dr J G RAY**					
Course Summary & Justification	The course is to introduce the concept of sustainable agriculture, especially its principles of ecological sustainability. The course will equip students to understand the concept of organic farming. It will enable an understanding of plant nutrient management as well as pest management in sustainable agriculture. Organic farming is becoming an internationally significant agricultural practice, and the knowledge has global significance. Interdisciplinary biology students with a good understanding of organic farming will enable our students to find suitable job opportunities in such farming industries.					
Semester	First					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	E.g., Authentic learning Collaborative learning Independent learning	60	18	0	28	106
Pre-requisite	**Knowledge in Botany at the Graduate level**					

No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**	
1	Students will develop a critical knowledge of the basic principles of sustainable agriculture	R/U/A	1	
2	They will be able to analyze environmental issues related to chemicalized agriculture	U/A	1	
3	They will acquire the basic skills of sustainable organic agriculture	U/An/Ap	1	
4	They will develop the skills to evaluate different kinds of farming	An/Ap	1	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)				

Module No	Module content	CO	
1	Introduction to Sustainable agriculture: Concept of ecological sustainability and sustainable agriculture-Natural, Ecological and organic farming – definition, concepts, and practices – management, principles, methods, merits and demerits.	1,2	10
2	Challenges to Sustainable agriculture – Productivity vs sustainability; Soil organic matterdecomposition, C: N ratios, mineralization and immobilization processes, hummus, the role of organic matter in soil quality – natural way to prevent soil degradation and erosion, types and control measures. Soil related water pollution- sources, different pollutants in soils and their managements Plant nutrient management in sustainable agriculture: Bio-availability of nutrients in soils, deficiency symptoms on plants, nutrient interactions and chelated micronutrients.Bio-fertilizers – benefits - classifications, production - maintenance and application	1,2	20
3	Organic Manures – bulky and concentrated – FYM – Biocomposting, Compost – rural, urban, vermicompost and coirpith; Panchagavya preparation and other organic nutrients application - Enrichment of organic manures; Sewage and sludge; Green manures – potentials and limitations; Quality parameters of organic manures and specifications – Biofertilizers -	3,4	10
4	Biopesticides and biological control agents: Types of biocontrol agentsbiological agents and pheromones, control of weeds, diseases and insect pests and field sanitation - competition, predation, antibiosis and fungistatic Efficacy of traditional biopesticides - Botanical insecticides- beneficial insects like the honeybee, lac insect, silkworm and pollinators Biological control - concepts and potentialities for managing soil-borne pathogens. Types of biological interactions, competition, 1.078 mycoparasitism; Mycorrhizal associations, Biodynamic products, Biodynamic composting, Liquid manure, Influence of Bio-dynamic products on crop production. Visit Organic Farms	3,4	20
	Total Credits of the course	3	

Books for References

Compulsory Reading:

1. Dahama AK (2007). Organic Farming for Sustainable Agriculture. 2nd Edn. Published by AGROBIOS (India) Jodhpur
2. National Standards Programme for Organic Production and Organic Products (2000) Department of Commerce, Ministry of Commerce and Industry, Govt. of India

Further Reading:

3. Gehlot D (2005). Organic Farming: Standards, Accreditation, Certification and Inspection, AGROBIOS (India) Jodhpur
4. Gupta PK (2007). Soil, Plant, Water and Fertilizer Analysis Published by AGROBIOS (India), Jodhpur

5. Sadasivam S and Manickam A (1992). Biochemical Methods for Agricultural Sciences Wiley Eastern Limited and Tamil Nadu Agricultural University, Coimbatore	

Rubrics selected for OBE implementation

1.Overall performance in each course of the semester on a continuous basis

2. Response to **critical theoretical questions** in each course

3. Procedural approach adopted towards **lab oriented critical questions** in each practical course

4. Response to **socially relevant issues and recent trends** in each course

5.Aptitude to research and **specific research problem** in each course

PART 1Task Description

1. Written Examination
2. Assignment
3. Seminar
4. Practical Exam
5. Viva voce

PART II Scale- Continuous mode

Excellent, Satisfactory, Needs improvement (Remedial practices recommended)

PART III Dimensions

Written Examination-Content, Communicating

Assignment -Content, level of Comprehension

Seminar-Content, Performance

Practical exam- Conduct of practical, Observation and recording

Viva voce -Response to questions, Attitude

PART IV Description of the dimensions

Content-Brief and meaningful

Comprehension- Precise and effective

Communicating-Direct and orderly

Procedure adopted- Scientific Suitability and easiness

Conduct of practical-Accuracy and reproducibility

Observation and recording- Sharp and systematic

Response to questions- Analytical approach and level of accuracy

Attitude- Positive and self-inspiring

PREVIOUS QUESTION PAPERS

School of Biosciences

Mahatma Gandhi University

SBS MIC1703: CELL BIOLOGY, GENETICS &EVOLUTION

Model Questions: (5 marks each for all the three)

TIME 3 HRS MAX MARKS 60

A. Easy

1. What are the common characteristics of phenotypic expressions in Quantitative Inheritance with the example of skin colour in human beings. Explain the basic differences between multiple allelic and Multigenic inheritance.
2. Comment on the role of p21 in the DNA replication checkpoint. Explain with appropriate diagram.
3. What are the factors affecting recombination frequencies?
4. Compare apoptosis and necrosis?

B. Medium

5. Explain why monohybrid and dihybrid ratios are different? Solve the following problem: A tall Red flowered plant when crossed with a dwarf white flowered plant there four different offspring such as Tall Red, Tall white, Dwarf Red and Dwarf white. But when the same Tall Red flowered plant was crossed with another Tall Red flowered plant, all the 2229 offspring produced were Tall Red. Using a checker-board analysis find out the genotype of all the parents and offspring in both cases.
6. Specify the role of Cdc 14 phosphatase in the regulation of cell cycle. Explain how it prompts the exit of mitosis. What is the significance of sequestration of Cdc 14 phosphatase?

7. The father supplies a mutated UBEA3 and the mother supplies a mutated SNRPN to a child. What will be the phenotype of the child? Explain the reason for the said phenotype?
8. What is the application of transduction by blue white screening?

C. Difficult

9. (**a**) Explain the evidences provided by Charles Darwin to prove his theory of origin of new species through natural selection? (**b**) How do the Lamarckian and Darwinian theories become identical as per genetic principles followed by them? (**c**) Explain how modern findings in population genetics defeats Lamarckian concepts while it is supportive of Darwinian theory of origin of species by natural selection?
10. Intensive research in Cell Biology has provided significant contributions in the treatment of various types of cancer. Justify the statement
11. Hardy Weinberg equilibrium is shown only by cross pollinated plants and not by self-pollinated plants? Justify the statement based on HW theorem?
12. Describe any two recent research using yeast as a genetic system.

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School of Biosciences
Mahatma Gandhi University
I Semester MSc Biochemistry/Biotechnology/Microbiology/Biophysics Examination
SBS MIC 1701 Biochemistry

***Draw structures wherever needed.**

Time 3 hours **Max marks 60**

Answer all of the following briefly. Each question carries *five* marks

1. Draw the predominant conformation of glucose and explain why it is predominant in nature.
2. Explain the characteristics of amphipathic lipid aggregates that form in water?
3. Describe the differences in the mechanism of action of progesterone and prolactin.
4. Does sucrose exhibit mutarotation? Give the reason for your answer.
5. Draw the structures of tryptophan and arginine at pH 7.Explain the importance of charge-based separation techniques during the isolation of proteins.
6. Suppose that the Sanger dideoxy method shows that the template strand sequence is 5'-TGCAATGGC-3'. Sketch the gel pattern that would lead to this conclusion. What is the purpose of the dideoxynucleoside triphosphate in this reaction?
7. Give a list of three vitamins that act as coenzymes and explain their importance.
8. Distinguish between Watson-crick base pairing and Hoogsteen base pairing.
9. Discuss the chemistry and physiological functions of prostaglandins and thromboxanes?
10. Explain the following terms (with examples) related to protein structure
    a. Supersecondary structures
    b. Motifs
    c. Domains
    d. oligomer
11. Which is more common - alpha helix or beta pleats? Substantiate your answer.
12. Mention the importance of lipid molecules containing sphingomyelin.

(12 x 5= 60 marks)

School of Biosciences
Mahatma Gandhi University
I Semester MSc Biochemistry/Biotechnology/Microbiology/Biophysics
(Supplementary) Examination
SBS MIC 1701 Biochemistry

***Draw structures wherever needed.**

Time 3 hours **Max marks 50**

Answer all of the following briefly. Each question carries *five* marks

1. Explain the role of Ramachandran plot in protein structure prediction.
2. Glycogen, starch and cellulose are made of glucose molecules. Yet they are so different in their structure and function. Explain the reason.
3. Design a set of experiments for the isolation and purification of a membrane protein.
4. Write a note on sugar derivatives.
5. Write notes on any two physiologically relevant molecules formed from cholesterol.
6. Distinguish between secondary structures and supersecondary structures**.**
7. What happens to glucose and mannose when they are treated with phenyl hydrazine? Draw the chemical reaction.
8. Explain the relevance of base pairing. Why is RNA mostly single stranded?
9. What happens if glucose is stored as such in the cells instead of as glycogen?
10. Explain the structural characteristics of t-RNA.

(10 x 5= 50 marks)

School of Biosciences
Mahatma Gandhi University
I Semester MSc Biochemistry/Biotechnology/Microbiology/Biophysics Examination

SBS MIC 1701 Biochemistry

Answer all questions. Each question carries 5 marks.

1. Glucose and fructose form same osazone. Why?
2. Distinguish between secondary structures and supersecondary structures**.**
3. Detail the importance of different proteins in DNA compaction.
4. Design a set of experiments for the isolation and purification of a membrane protein.
5. What is the role of Ramachandran plot in the structure determination of proteins?
6. If starch, glycogen and cellulose are made up of glucose molecules, what is the basis for the difference in their structure and function?
7. The densities of lipoproteins increase as their particle diameters decrease. Why?
8. Eicosanoids can act as hormones. Explain.
9. How does vitamin and hormones differ in their action?
10. Describe in your own words the structural features of
    a. a ceramide and how it differs from a cerebroside.
    b. a phosphatidylethanolamine and how it differs from a phosphatidylcholine.
    c. a ganglioside and how it differs from a cerebroside

11. A polypeptide when treated with 2-mercaptoethanol yielded two polypeptides with the following amino acid sequences:
    a. Leu-Phe-Cys-Met-Tyr-Cys-Leu-Trp-Cys-Asn
    b. Val-Cys-Trp-Val-Ile-Phe-Ala-Cys-Lys

    These two polypeptides on chymotrypsin treatment yielded small peptides with the following amino acid compositions:
    a. (Leu, Phe)
    b. (Asn, Cys2, Met, Tyr)
    c. (Cys, Ala, Lys)
    d. (Cys2, Leu, Trp2, Val)
    e. (Ile, Phe, Val)
12. Find out the positions of the disulfide bonds in the parent polypeptide.

The following DNA fragment was sequenced by the Sanger method. The fragments were produced by this reaction was separated by gel electrophoresis. Sketch the gel pattern.

School of Biosciences

Mahatma Gandhi University

I Semester MSc Biochemistry/Biotechnology/Microbiology/Biophysics Examination, July 2021

SBS MI C 1704: BIOPHYSICS & BIOSTATISTICS

TIME: 3 HRS **MARKS: (12 X 5 = 60)**

1. Explain multimolecular and macromolecular colloids? Give one example of each. How are associated colloids different from these two types of colloids?

2. Describe the following processes
a) Redox potential
b) Surface tension
c) Osmosis
d) Adsorption
e) Carnot cycle

3. A simple sample of the height of 6,400 Englishmen has a mean of 67.85 inches and a standard deviation of 2.56 inches while a simple sample of heights of 1,600 Austrians has a mean of 68.55 inches and standard deviation of 2.52 inches. Do the data indicate that the Austrians are on the average taller than the Englishmen?

4. Mention the physical interactions of proteins with nucleic acids. Comment on the Shape, flexibility and packing of proteins and nucleic acids in complexes

5. Discuss about different types of electromagnetic radiation. Compare the properties of ionizing and non- ionizing radiations.

6. In a simple random sample of 600 men taken from a big city 400 are found to be smokers. In another simple random sample of 900 men taken from another city 450 are smokers. Do the data indicate that there is significant difference in the habit of smoking in the two cities?

7. How can we propose a protein secondary structure prediction method based on Ramachandran map? Diagrammatically represent the allowed and disallowed regions of Ramachandran plot.

8. Describe the biophysical method to purify bovine serum albumin from a mixture of salts and diagrammatically represents the instrumentation setup for that.

9. Explain the applications of electromagnetic radiation in the field of Agriculture, Medicine and Industry?

10. Two types of drugs were used on 5 and 7 patients for reducing their weight.
Drug A was imported and drug B indigenous. The decrease in the weight after using the drugs for six months was as follows:

Drug A: 10 12 13 11 14
Drug B: 8 9 12 14 15 10 9
Is there a significant difference in the efficacy of the two drugs?

11. From the following data estimate y when x =92

X Girth	Y Wood volume
90	0.5
95	0.6
100	0.7
85	0.5
88	0.53
70	0.40
72	0.41
74	0.42
70	0.39
69	0.38

12. From the following data, examine whether there is significant difference between three groups

Self (T1)	Govt (T2)	Aided (T3)
65	**60**	**74**
75	**62**	**78**
76	**70**	**80**
78	**74**	**82**
80	**76**	**88**

School of Biosciences
Mahatma Gandhi University
I Semester MSc Biochemistry/Biotechnology/Microbiology/Biophysics Examination
SBS MIC 1703 Cell Biology, Genetics and Evolution

Time 3 hours **Max marks 60**

Answer all of the following briefly. Each question carries *five* marks

1. Find out the Caucasian admixture of Afro Americans in two different cities of the United States of America by analysing the data given in the table. The data is about the presence of Duffy blood group allele in different populations.

Locality	Afro Americans population	African population	Caucasian population
Oakland	0.094	0.0	0.429
Charleston	0.016	0.0	0.429

2. A hypothetical mutant phenotype, wrinkled chloroplast shows a maternal pattern of inheritance in the dandelion plant. What would be the outcome of
   a) mating male wrinkled to female smooth (wild)
   b) mating male smooth to female wrinkled
3. What are the factors affecting recombination frequencies?

4 .(**a**) Explain the phenotypic expressions of Comb pattern in fowls using a checker board? (**b**) How does the mode of inheritance of 'Comb pattern in fowls' differ from that of 'Fruit colour in summer squashes'?

5.(**a**) Compare and contrast a typical Mendelian dihybrid appearance of four phenotypic classes of offspring with the four phenotypic classes of offspring of a cross between heterozygous 'A-blood group' and heterozygous 'B-blood group' persons? (**b**) If a 'B-blood group' woman who is married to an 'O-blood group' man have three children, and one among them is an 'O-blood group' boy, explain the genotypes of the parents and that of all the possible children to them.

6.(**a**) Compare and contrast Lamarkism and Darwinism (**b**) Explain the advantages of Darwinism over that of Lamarkism in the explanation of evolution of species, especially in relation to the modern theory of genetics? (**c**) How do natural selection work as per the concept of population genetics?

7.Illustrate the Biotechnological application of bacterial genetic transformation.

8. Describe suitably features and valuable research conducted in *Neurospora* to establish a model organism

9.Specify the differences between cancerous cells and normal cells.

10. Comment on the significance of DNA damage check points in the regulation of cell cycle.

11.What are polarity and segmentation genes? Discuss their role in the genetic control of embryonic development in drosophila

12. Discuss the regulatory role of cell survival pathway in apoptosis.

(12 x 5= 60 marks)