MAHATMA GANDHI UNIVERSITY
School of Biosciences

Priyadarshini Hill PO Kottayam – 686560

Learning Outcomes based Curriculum Framework (LOCF)for Post Graduate Programme

Under the CSS scheme for University

(EFFECTIVE FROM 2020 ADMISSIONS)

Preface

Mahatma Gandhi University

Mahatma Gandhi University is an Indian collegiate public University based in Kerala, established in 1983, approved by UGC, and accredited with NAAC "A" Grade, 3.24 CGPA. With its academic excellence, the University has bagged Chancellor's Award twice for the best University (2015-16 and 2017-18) within the state of Kerala. It has also secured 30th position in NIRF ranking (April 2019) and 11th position in India Today-MDRA ranking, 2018. CSIR has ranked the University 13th for its intellectual productivity and NISTADS has rated it as 19th in terms of h-index.

At present, Mahatma Gandhi University offers research programs in forty disciplines through its own Schools and approved Research Centers. It has close collaboration for academic, research and extension programs with a number of national agencies and institutions including the UGC, DST-FIST, DRS, ISRO, COSIT, DIT, DST (Nano Mission), CSIR, DAAD, STEC, ICMR, BARC and MOEF. The University is also involved in active collaboration with research institutions of international reputation such as the Max Planck Institute of Technology, Germany; Brown University, USA; University of Nantes, France; California Institute of Technology, USA; University of Toronto, Canada; Catholic University, Belgium; Heidelberg University, Germany; the Institute of Political Studies, Rennes, France; Trent University, Canada; IPF Dresden, Germany; University of Paris and University of Strasbourg.

Mahatma Gandhi University has made immense strides in the fields of inter disciplinary teaching and research. The faculty comprises of outstanding scholars, many of whom havemade original contributions in their respective fields of specialization. The faculty and research scholars of several departments have gained widespread recognition for the commendable

quality of their research publications. The web enabled University library has large collection of books, journals, e-journals and online theses. The digital library provides open access to its enviable collection of digitized Ph.D dissertations. All these work in tandem with the academic business transacted by the University, making the whole experience a holistic one. The University has a well-established instrumentation facility with many sophisticated equipments functioning at the various departments and also at the platform provided by the common Inter University Instrumentation Centre (IUIC). The University also has an Inter University Centre for Organic Farming and Sustainable Agriculture (IUCOFSA) which is a renowned centre for promoting interdisciplinary research in organic farming and conservation of traditional knowledge base, through effective networking and innovative extension activities among the public. The centre has developed many organic methods of agricultural activities for the cultivation of different varieties of rice and vegetables.

The University has well established and internationally reputed facility and academic expertise in various areas like Nanoscience, Environmental science, Bioscience, Chemical science, Physics, Arts and Humanities. The Centre for Nanoscience and Nanotechnology focus on the enhancement of research and higher studies in the cutting-edge areas of Nanoscience and Nanotechnology. The Centre is motivated to thrust its research and development focusing on developing novel materials and devices prospering the outrage of Nanoscience. With a vision to consolidate the existing and to pay focus attention to the frontier areas of Environmental Science, the University has established the School of Environmental Sciences as a Centre of learning for advanced studies in different branches of environmental science. The major mandate of the School is to develop appropriate technologies and skilled human resource for sustainable utilization, management and conservation of natural resources. The school has established a Centralized Remote Sensing and GIS facility, the first ofits kind in a University in the

state, with the support of Indian Space Research Organization (ISRO). It has also established a regional center, the Highrange Environmental Research center (HERC) at Nedumkandam, Idukki district. The School has a live laboratory named as “Jeevaka” which consists of areas with rich biodiversity within the Mahatma Gandhi University Campus.

Vision and Mission of MGU

Vision of Mahatma Gandhi University

"Mahatma Gandhi University envisions to excel in the field of higher education and cater to the scholastic and developmental needs of the individual, through continuous creation of critical knowledge base for the society's sustained and inclusive growth."

Mission of Mahatma Gandhi University

- **To conduct and support undergraduate, postgraduate and research-level programmes of quality in different disciplines**
- **To foster teaching, research and extension activities for the creation of new knowledge for the development of society**
- **To help in the creation and development of manpower that would provide intellectual leadership to the community**
- **To provide skilled manpower to the professional, industrial and service sectors in the country so as to meet global demands**
- **To help promote the cultural heritage of the nation and preserve the environmental sustainability and quality of life**
- **To cater to the holistic development of the region through academic leadership**

Preamble

OUTCOME BASED EDUCATION (OBE)FROM THE ACADEMIC YEAR 2020-21**MAHATMA GANDHI UNIVERSITY SCHOOL OF BIOSCIENCES**

1. Introduction

A high priority task in the context of education in India is improvement of quality of higher education for equipping young people with skills relevant for global and national standards and enhancing the opportunities for social mobility. Mahatma Gandhi University has initiated an Outcome Based Education (OBE) for enhancing employability of graduates through curriculum reforms based on a learning outcomes-based curriculum framework, upgrading academic resources and learning environment.

Learning outcomes specify what graduates completing a particular programme of study are expected to know, understand and be able to do at the end of their programme of study. The fundamental premise underlying the learning outcomes-based approach to curriculum development is that higher education qualifications are awarded on the basis of demonstrated achievement of outcomes, expressed in terms of knowledge, understanding, skills, attitudes and values. Outcomes provide the basis for an effective interaction among the various stakeholders. It is the results-oriented thinking and is the opposite of input-based education where the emphasis is on the educational process.

Benefits of OBE

The OBE Framework is a paradigm shift from traditional education system into OBE system where there is greater focus on programme and course outcomes. It guarantees that curriculum, teaching and learning strategies and assessment tools are continuously enhanced through a continuous improvement process. All decisions including those related to

curriculum,delivery of instruction and assessment are based on the best way to achieve the predetermined outcomes. Traditionally, educators have measured learning in terms of standardised tests. In contrast, outcome-based education defines learning as what studentscan demonstrate that they know.

Benefits of OBE:

*More directed & coherent curriculum.
*Graduates will be more "relevant" to industry & other stakeholders (more well-roundedGraduates)
*Continuous Quality Improvement is in place.
*OBE shifts from measuring input and process to include measuring the output (outcome)

Outcome Based Education (OBE) process

OBE is a comprehensive approach to organise and operate a curriculum that is focused onand defined by the successful demonstrations of learning sought from each learner. The term clearly means focusing and organising everything in an education system around "what is essential for all learners to be able to do successfully at the end of their learning experiences".

OBE is an approach to education in which decisions about the curriculum and instruction are driven by the exit learning outcomes that the students should display at the end of aprogramme or a course. By the end of educational experience, each student should have achieved the outcomes.

Learning Outcomes based Curriculum Framework (LOCF) for Post Graduate Programmes-

IQAC MG University

One of the main objectives of OBE is to ensure continuous improvement of programmes in terms of maintaining the relevance in

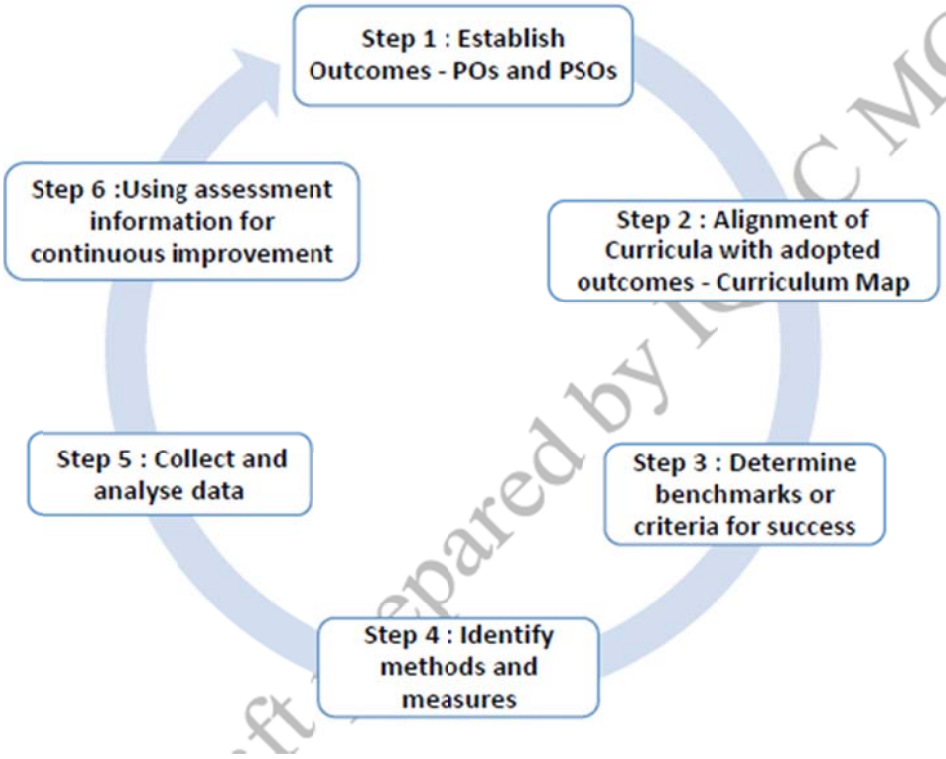
curriculum as well as responding to the requirements of the stakeholders. In other words, it ensures that Post graduate programme next year is better than Post graduate programme this year, offered by a department.

An OBE system has been proposed and to be implemented at various Departments of Mahatma Gandhi University, as a quality-assurance approach to improve teaching and learning outcomes and processes. This OBE plan incorporates the "outcomes assessment" process to be followed in the departments. OBE should be a key driver of the curriculum management in allthe departments of the university.

The OBE is a 6 step process as shown in the figure

Figure: OBE Process

The process is presented as a cycle or a loop. The cycle represents the continuous nature of assessing learning outcomes.



As envisaged by the IQAC of Mahatma Gandhi university, an OBE based curricular framework has been proposed for the School of Biosciences from the academic year 2020-2021 which is presented hereafter.

School of Biosciences

The Life Science research of the University is carried out under the School of Biosciences, which is another prestigious department of the University and it provides academic expertise to students in advanced areas of Biochemistry, Microbiology, Biotechnology and Biophysics. The established research areas at School of Biosciences specifically include the Bioprocess technology, toxicology, ethnopharmacology, inflammation, ecology, ecotechnology, agricultural microbiology, immunobiology, medicinal plant research, probiotic development, microbial and natural product research,

molecular microbiology etc. The department harbours a state -of-the-art instrumentation facility, animal maintenance facility and animal cell culture facility as well. The institute has been a successful aspirant in producing a large number of PhDs, and has completed several funded projects with significant number of publications.

Our Vision

* An Institution of excellence developing professional competence, ambition and determination in students to face new challenges and find new opportunities in the field of Biological Sciences and facilitating the wellbeing and prosperity of mankind especially our Mother Land by utilising the opportunities in advanced Biological research.

Key points

- Institution of excellence
- Professional competence, ambition and determination
- New challenges and new oppurtunities
- Well being and prosperity of nation and humanity
- Utilise opportunities in research

Our Mission

- To provide advanced knowledge and technological knowhow to the students in the field ofBiological sciences.
- To utilise the expertise of faculty in diverse areas of biology for benefitting the students inachieving their career goals.
- To conduct cutting-edge research in areas of life Sciences and to extend the knowledge gainedfrom lab to land and benchtop to bedside.

Key points

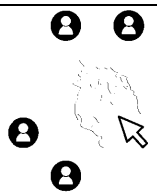
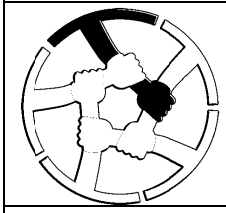
1. provide advanced knowledge and technological knowhow
2. To utilise the expertise of faculty
3. benefitting the students in achieving their career goals.

4. conduct cutting-edge research

5. extend the knowledge gained from lab to land and benchtop to bedside.

	Mahatma Gandhi University **Graduate attributes**

	**Critical thinking and analytical reasoning**	Capability to analyze, evaluate and interpret evidence, arguments, claims, beliefs on the basis of empirical evidence; reflect relevant implications to the reality; formulate logical arguments; critically evaluate practices, policies and theories to develop knowledge and understanding; able to envisage the reflective thought to the implication on the society.
	**Scientific reasoning and Problem solving**	Ability to analyze, discuss, interpret and draw conclusions from quantitative/qualitative data and experimental evidences; and critically evaluate ideas, evidence and experiences from an unprejudiced and reasoned perspective; capacity to extrapolate from what one has learned and apply their competencies to solve problems and contextualize into researchand apply one's learning to real life situations.
	**Multidisciplinary/ Interdisciplinary/ Transdisciplinary approach**	Acquire interdisciplinary /multidisciplinary/ transdisciplinary knowledge base as a consequence of the learning they engage with their programme of study; develop a collaborative-multidisciplinary/interdisciplinary/transdisciplinary-approach for formulate constructive arguments and rational analysis for achieving common goals and objectives.

	**Intra and Interpersonal skills**	Ability to work effectively and respectfully with diverse teams; facilitate collaborative and coordinated effort on the part of a group, and act together as a group or a team in the interests of a common cause and work efficiently as a member of a team; lead the team to guide people to the right destination, in a smooth and efficient way.
	**Digital literacy**	Capability to use ICT in a variety of learning situations, demonstrate ability to access, choose, collect and evaluate, and use a variety of relevant information sources; structure and evaluate those data for decision making.
	**Global Citizenship**	Building a sense of belonging to a common humanity and to become responsible and active global citizens. Appreciation and adaptation of different sociocultural setting and embrace and promote equity.
	**Social competency**	Possess knowledge of the values and beliefs of multiple cultures, appreciate and adapt to a global perspective; and capability to effectively engage in a multicultural society and interact respectfully, manage and lead with diverse groups.
	Equity, Inclusiveness and Sustainability	Appreciate and embrace equity, inclusiveness and sustainability and diversity; acquire ethical and moral reasoning and values of unity, secularism and national integration to enable to act as dignified citizens; able to understand and appreciate diversity
	**Lifelonglearning**	Continuous acquisition of knowledge and skills. Learn, unlearn and re-learn based on changing ecosystem. “Learning how to learn”, that are necessary for participating in learning activities throughout life, through self-paced and self-directed learning aimed at personal development, meeting economic, social and cultural objectives, and adapting to changing trades and demands of work place through knowledge/skill development/reskilling.

	**Mahatma Gandhi University** **Programme Outcome**

Programme Outcomes (PO)

PO 1: Critical Thinking and Analytical Reasoning

Capability to analyse, evaluate and interpret evidence, arguments, claims, beliefs on the basis of empirical evidence; reflect relevant implications to the reality; formulate logical arguments; critically evaluate practices, policies and theories to develop knowledge and understanding; able to envisage the reflective thought to the implication on the society.

PO 2 : Scientific Reasoning and Problem Solving

Ability to analyse, discuss, interpret and draw conclusions from quantitative/qualitative data and experimental evidences; and critically evaluate ideas, evidence and experiences from an unprejudiced and reasoned perspective; capacity to extrapolate from what one has learned and apply their competencies to solve problems and contextualise into researchand apply one's learning to real life situations.

PO 3: Multidisciplinary/Interdisciplinary/Transdisciplinary Approach

Acquire interdisciplinary /multidisciplinary/transdisciplinary knowledge base as a consequence of the learning they engage with their programme of study; develop a collaborative-multidisciplinary/interdisciplinary/transdisciplinary-approach for formulate constructive arguments and rational analysis for achieving common goals and objectives.

PO 4: Communication Skills

Ability to reflect and express thoughts and ideas effectively in verbal and nonverbal way; Communicate with others using appropriate channel; confidently share one's views and express herself/himself; demonstrate the ability to listen carefully, read and write analytically, and present complex information in a clear and concise manner and articulate in a specific context of communication.

PO 5: Leadership Skills

Ability to work effectively and lead respectfully with diverse teams; setting direction, formulating an goal, building a team who can help achieve the goal, motivating and inspiring team members to engage with that goal, and using management skills to guide people to the right destination, in a smooth and efficient way.

PO 6: Social Consciousness and Responsibility

Ability to contemplate of the impact of research findings on conventional practices, and a clear understanding of responsibility towards societal needs and reaching the targets for attaining inclusive and sustainable development.

PO 7: Equity, Inclusiveness and Sustainability

Appreciate equity, inclusiveness and sustainability and diversity; acquire ethical and moral reasoning and values of unity, secularism and national integration to enable to act as dignified citizens; able to understand and appreciate diversity, managing diversity and use of an inclusive approach to the extent possible.

PO 8: Moral and Ethical Reasoning

Ability to embrace moral/ethical values in conducting one's life, formulate a position/argument about an ethical issue from multiple perspectives, and use ethical practices in all work. Capable of demonstrating the ability to identify ethical issues related to one's work and living as a dignified person in the society.

PO 9: Networking and Collaboration

Acquire skills to be able to collaborate and network with scholars in an educational institutions, professional organizations, research organizations and individuals in India and abroad.

PO 10: Lifelong Learning

Ability to acquire knowledge and skills, including "learning how to learn", that are necessary for participating in learning activities throughout life, through self-paced and self-directed learning aimed atpersonal development, meeting economic, social and cultural objectives, and adapting to changing trades and demands of work place through knowledge/skill development/reskilling.

Programme outcome of MSc courses in School of Biosciences (PS)

To develop competent personnel in applied branches of life sciences with goodacademic standards, skill, technical knowhow, research aptitude, scientific ethics and societal consciousness.

Programme specific outcomes of M.Sc. Biophysics

PSO1. To get in depth scientific knowledge about the prokaryotic and eukaryotic cellular, physiological and biochemical processes, interaction of microorganisms among themselves, with physical and chemical agents and beneficial and harmful effects and applications of microorganisms and to explore its immense scope both at the basic and application aspects.

PSO2. To meet the global demand for skilled scientific resources in microbiology, the program has been designed to understand the basics and advances of microbiology to accelerate the microbiological contributions for diverse applications by integrating the knowledge through interdisciplinary approach.

PSO3. To translate the technical knowhow in various branches of microbiology like the medical, industrial, food, agricultural and environmental microbiology for the well being of humans, other living forms and the environment and to motivate the innovation in microbiological applications for the emerging and existing challenges

PSO4. To generate trained human resources with the skills to use the microbiological methods, instrumentation tools, and technologies by following the standard operation procedures and scientific ethics for the research, clinical, agricultural, industrial and other commercial applications through the design and execution of microbiological experiments for both the local and global societies.

PSO5. To provide a vibrant and internationally competitive educational environment that fosters the development of scientific vocabulary, reasoning skills, entrepreneurial skills, communication abilities and societal consciousness

SCHEME OF MSc BIOPHYSICS PROGRAMME

FIRST SEMESTER SCHEME

Course Code	Course Title	Credits
SBS M P C 01	Biochemistry	3
SBS M P C 02	Microbiology	3
SBS M P C 03	Cell Biology, Genetics &Evolution	3
SBS M P C 04	Biophysics &Biostatistics	3
SBS M P C 05	Physiology	3
SBS M P C 06	Laboratory Course – 1	3
SBS M P C 07	Laboratory Course – 2	3

SECOND SEMESTER SCHEME

SBS M P C 08	Immunology	3
SBS M P C 09	Molecular Biology and Genetic Engineering	3
SBS M P C 10	Metabolism and Bioenergetics	3
SBS M P C 11	Biophysical Techniques and Bioinstrumentation	3
SBS M P C 12	Laboratory Course – 3	4
	Elective Course to be selected from the optionsgiven below	3
	Total Credits of the 2nd Semester Programme	19
	Elective Courses Offered by Different Teachers in the 2nd Semester	

SBS M P E 13	Microbial Technology	3
SBS M P E 14	Ecology and Environment	3
SBS M P E 15	Neurobiology	3
SBS M P E 16	Environment Science	3
SBS M P E 17	Molecular Microbiology	3
SBS M P E 18	Developmental Biology	3

SCHEME OF THIRD SEMESTER BIOPHYSICS

Course No	**Subject of the Course**	Credit
SBS M P C 19	Enzymology	3
SBS M P C 31	Molecular Biophysics	3
SBS M P C 32	Electrophysiology	3
SBS M P C 33	Laboratory Course 4-Biophysics	4
Course taken by the student from other departments	Open course	4
One Elective course to be selected from the options given below		3
Total Credits of the 3rd Semester Programme in M Sc Biophysics		20

SCHEME OF THIRD SEMESTER OPEN ELECTIVE COURSES

Students need to select one open elective course offered by other departments

	Course No.	**Subject of the Course**	**Credits**

	SBS M P O 34	Biotechnology and Society	4
	SBS M P O 35	Microbiology in Everyday Life	4
	SBS M P O 36	Environment Lead Auditor Course	4
	SBS M P O 37	System Biology	4
	SBS M P O 38	Sustainable Agriculture	4
	SBS M P O 39	Ecology of Soil Fertility	4
	SBS M P O 40	Infectious Disease Management	4
	SBS M P O 41	Probiotics and Nutraceuticals	4

SCHEME OF THIRD SEMESTER ELECTIVE COURSES			
31	SBS M P E 42	Quality Control in Herbal Drugs	3
32	SBS M P E 43	IPR and Patenting	3
33	SBS M P E 44	Advanced Techniques in Diagnostic Microbiology	3
34	SBS M P E 45	Radiation Biophysics	3
35	SBS M P E 46	Algal Biofuel Technology	3

SCHEME OF FOURTH SEMESTER BIOPHYSICS		
Course No	**Subject of the Course**	Credit
SBS M P C 56	Biophysical chemistry	4
SBS M P C 57	Laboratory Course 5- Biophysics	3
SBS M P C 58	Major Research Project	7
Elective 1	To be selected from among the elective courses offered	3
Elective 2	To be selected from among the elective courses offered	3
Total Credits of the 4th Semester Programme in M Sc Biophysics		20

SCHEME OF FOURTH SEMESTER ELECTIVE COURSES Students need to select any two of the following elective courses				
43	SBS M P E 59	Environment Biotechnology		3
44	SBS M P E 60	Omics in Biotechnology		3
45	SBS M P E 61	Molecular Phylogeny		3
46	SBS M P E 62	Plant Microbe Interactions		3
47	SBS M P E 63	Human Virology		3
48	SBS M P E 64	Physiological Biophysics		3
49	SBS M P E 65	Good Laboratory Practices		3
50	SBS M P E 66	Medical Biophysics		3
51	SBS M P E 67	Biofertilizers and Biopesticides		3
52	SBS M P E 68	Health and Nutrition		3
53	SBS M P E 69	Neutrophil Biology		3
54	SBS M P E 70	Medicinal Plants		3

	MAHATMA GANDHI UNIVERSITY
	SBS M P C 01: BIOCHEMISTRY

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**BIOCHEMISTRY**					
Type of Course	Core					
Course Code	**SBS M P C 01**					
Names of Academic Staff & Qualifications	1. **Prof. M S Latha – M. Sc (Biochemistry), PhD** 2. **Dr. Anie Y – M. Sc (Biochemistry), PhD** 3. **Guest faculty – M. Sc (Biochemistry), PhD**					
Course Summary & Justification	The course is designed to get a clear idea on the basic biomolecules and their importance in the various biochemical processes in life so that the course builds a base for the students to comprehend and articulate the advanced concepts in life sciences.					
Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Eg. Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding of chemical groups and bonding; basics of cell biology and physiology					

O No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**
1	To identify the different types of biomolecules such as	U	

	lipids, carbohydrates, proteins and nucleic acids		
2	To differentiate the structural and functional characters of different biomolecules	A	
3	To narrate the coordinated functions of different biomolecules in a complex living system	A/Ap	
4	To compare the structure and functions of biomolecules in plants, animals and microbes	A	
5	To describe the structure and functions of vitamins and hormones	U	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hours
1	**Carbohydrates:** Classification of Carbohydrates with examples- monosaccharides, disaccharides and oligosaccharides; their structure and functions; Polysaccharides - occurrence, structure, isolation, properties and functions of homoglycans- starch, glycogen, cellulose, dextrin, inulin, chitins, xylans, arabinans, galactans. Occurrence, structure, properties, and functions of heteroglycans – bacterial cell wall polysaccharides, glycoaminoglycans, agar, alginic acid, pectins, amino sugars and deoxv sugars, blood group substances and sialic acids. Glycolipids and Glycoproteins and their biological applications. Lectin- structure and functions.	0.5	10
2	**Lipids:** Classification of lipids with examples; their structure and functions Complex lipids- phospholipids -classification, structure and functions. Ceramides and sphingomyelins. Eicosanoids, structure and functions of prostaglandins, thromboxanes, leukotrienes Types and functions of plasma lipoproteins. Amphipathic lipids -membranes, micelles, emulsions and liposomes. Steroids -cholesterol structure and biological role -bile acids, bile salts. Sterols in Plant system: Phytohormones: Brassinosterroids (functions); Sterols in microbial system: mycosterols.	0.5	10
3.	**Proteins:** Amino acids- Structure amd properties**,** Classification of proteins on the basis of solubility and shape, structure, and biological functions. Isolation, fractionation and purification of proteins. Denaturation and renaturation of proteins. Primary structure - determination of amino acid sequence of proteins. Ramachandran plot, Secondary, tertiary and quartenary structures of proteins. Detailed study	0.5	15

	on structure and function with an example: Fibrous Protein (Collagen) Globular protein (Hemoglobin)., Enzymes- Different classes and functions.		
4	**Nucleic Acids: Components of nucleic acids,** Watson -Crick model of DNA structure. A, B and Z DNA Cruciform structure in DNA, miscellaneous alternative conformation of DNA. Higher order organization of DNA. Methods for nucleic acid sequence determination, isolation and purification of DNA, molecular hybridization, Cot value curve, Reassociation kinetics, RNA Structure: Types of RNA; structure of mRNA, tRNA and rRNA ,Si RNA, micro RNA with emphasis on importance of structure to its function	0.5	10
5	**Vitamins and Hormones:** Vitamins -water soluble -thiamine, riboflavin, niacin, pyridoxine, folic acid, ascorbic acid-source, structure, biochemical functions, deficiency diseases, daily requirements; fat soluble -vitamin A, vitamin D2, vitamin E and vitamin K -sources, structure, biochemical functions, deficiency diseases, daily requirements. Hormones: different types, structures, their biological role and disorders. Mechanism of action of peptide and steroid hormones.	1.0	15
	Total Credits of the Course	3	

Books for Reference

Compulsory Reading:

1. Principles Of Biochemistry, 4/e (2006) by Robert Horton H , Laurence A Moran, Gray Scrimgeour K **Publisher:** Pearsarson **ISBN:** 0131977369, **ISBN-13:**9780131977365, 978-0131977365
2. Biochemistry 6th Edition (2007) by Jeremy M.berg John L.tymoczko Lubert Stryer **Publisher:** B.i.publicationsPvt.Ltd **ISBN:**071676766X **ISBN-13:** 9780716767664, 978-716767664
3. Lehninger Principles of Biochemistry, Fourth Edition by David L. Nelson Michael M. Cox
13. Publisher: W. H. Freeman; Fourth Edition edition (April 23, 2004) ISBN-10: 0716743396 ISBN-13: 978-0716743392

Further Reading:

- Biochemistry: A Students survival Guide by Hiram. F. Gilbert (2002) Publishers: McGraw-Hill ISBN 0-07-135657-6
- Introduction to Biophysics by Pranab Kumar Banerjee (2008) Publishers: S. Chand & Company ltd ISBN: 81-219-3016-2
- **E.S. West**, **W.R. Todd**, **H.S. Mason and J.T. van Bruggen**, A **Text Book of Biochemistry**, Oxford and IBH Publishing Co., New Delhi, 1974
- Biochemistry [with Cdrom] (2004) by Donald Voet, Judith G. Voet **Publisher:** John Wiley & Sons Inc **ISBN:** 047119350X **ISBN-13:** 9780471193500, 978-0471193500
- Principles Of Biochemistry (1995) by Geoffrey L Zubay, William W Parson, Dennis E Vance **Publisher:** Mcgraw-hill Book Company – Koga **ISBN:**0697142752

ISBN-13: 9780697142757, 978-0697142757

- Molecular Biology of the Cell by Bruce Alberts, Alexander Johnson, Julian Lewis, Martin Raff, Keith Roberts, Peter Walter Publisher: Garland Science; 5 edition ISBN-10: 0815341059 ISBN-13: 978-0815341055
- Genes IX by Benjamin Lewin (2008) Publisher: J&b ISBN:0763752223 ISBN-13: 9780763752224, 978-0763752224
- Molecular Biology Of The Gene 5/e (s) by James D Watson, Tania A Baker, Stephen P Bell (2008) Publisher: Dorling Kindersley (India) Pvt Ltd ISBN: 8177581813 ISBN-13: 9788177581812, 978-8177581812
- Cell and Molecular Biology, 3e (2003) by Karp Publisher: Jw ISBN: 0471268909 ISBN-13: 9780471268901, 978-0471268901

Molecular Cell Biology (2002) by H.S. Bhamrah Publisher: Anmol Publications ISBN: 8126111429 ISBN-13: 9788126111428, 978-8126111428

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** **A. Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks **B. Semester End examination – 60 marks**

Approval Date	
Version	
Approval by	
Implementation Date	

MAHATMA GANDHI UNIVERSITY

SBS M P C 02: MICROBIOLOGY

SchoolName	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**MICROBIOLOGY**					
Type of Course	Core					
Course Code	**SBS M P C 02**					
Names of Academic Staff & Qualifications	**Dr.JISHA.M.S**					
Course Summary & Justification	is course on Microbiology introduces the milestones of Microbiology key components and their functions. e objective of the course content is to impart Knowledge on Landmark discoveries in Microbiology and different domains classification of living organisms. develop a very good understanding of the characteristics of different types of microorganisms, methods to organize/classify these into and basic tools to study these in the laboratory.					
Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours

	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of General microbiology**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	mmarize the contributions made by prominent scientists in microbiology and bacterial taxonomy	E	
2	derstanding of basic microbial structure and similarities and differences among various groups of microorganisms	U/ An	

3	Exemplify basic tools to study these in the laboratory	S	
4	Explain various factors affecting the microbial growth and nutritional requirements and will be acquainted with methods of measuring microbial growth	U/R	
5	alyze various methods for identification and sterilization of isolated microorganisms.	An	
6	Create an insight to the interactions and characteristics of microorganisms	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**History and scope of microbiology:** The historical foundations and development of microbiology. An overview of microbial world. The bacteria and the archae. Classification of bacteria and Archaea according to the Bergey's Manual of Systematic Bacteriology. Tools for Systematics: Numerical taxonomy, Phylogenetic analysis, Polyphasic approach; Modern methods of studying microbial diversity; Microbial culture collections.	0.5	10
2	**Microbial Diversity:** Prokaryotic and eukaryotic microbial diversity. General characteristics of various groups of prokaryotes: bacteria including, Rickettsiae, Chlamydiae and Actinomycetes, Cyanobacteria and Mycoplasmas. Morphology and structure of bacteria. Viruses unique properties, morphology, structure and cultivation; Viroids and Prions.Viral replication.Viral diversity–bacterial, plant and animal viruses; Fungi - properties and classification. Microorganism in extreme environments..	1.0	10
3.	**Microbial physiology:** Factors influencing microbial growth. Environmental and nutritional factors. Nutritional types of bacteria. Microbial growth curve. Mathematical expression of growth- continuous and batch cultures. Diauxic and synchronous growth. Measurement of bacterial growth.Cultivation of bacteria-culture media and methods. Aerobic and Anaerobic culture methods. Culture preservation techniques. Microbial locomotion – flagellar motility, gliding motility and amoeboid motion. Chemotaxis, Phototaxis and other taxes. Microbial photosynthesis.	1.0	20
4	**Identification of bacteria and Sterilization methods:** Identification of bacteria. Staining reactions. Cultural, physiological and biochemical properties. Molecular methods for identification. Sterilisation – Principles and methods, physical and chemical methods. Disinfectants – modes of action. Testing of disinfectants.	0.5	10

	Antibiotics – mechanism of action. Drug resistance in bacteria. Antibiotic sensitivity tests		
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction: Active co-operative learning, Seminar, Group Assignments Authentic learning: Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** C. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks D. **Semester End examination – 60 marks**

References

Compulsory Reading:

1. Prescott, L. M., Harley, J. P. and Klein, D. A.2014. *Microbiology*. 9th Edition. Edition, McGraw Hill Higher Education.
2. . Pelczar, M. J. Jr., Chan, E. C. S. and Krieg, N. R. 1993. *Microbiology*, 5th Edition, Tata MacGraw Hill Press.

Further Reading:

1. Jeffrey C.Pommerville.2016.Alcamos fundamentals of microbiology. Tenth Edition. Jones and Bartlett Learning.
2. Tortora G. J., Funke B. R. and Case C. L. 2015. *Microbiology: An Introduction*. 12th Edition. Pearson Education Inc.
3. Madigan, M. T. and Martinko, J. M. 2015. *Brock's Biology of Microorganisms*. 14th Edition. Pearson Education Inc.
4. .Willey, J. M., Sherwood, L. M. and Woolverton, C. J. 2013. *Prescott's Microbiology*. 8th Edition, McGraw-Hill Higher Education.
5. Stanier, R. Y., Adelberg, E. A. and Ingraham, J. L. 1987. *General Microbiology*, 5th Edition. Macmillan Press Ltd.
6. Russell, A. D., Hugo, W. B., and Ayliffe, G. A. J. 2013. *Principles and practice of disinfection, preservation and sterilization*, 5th Edition. Blackwell Science, Oxford.
7. Black, J. G. 2013. *Microbiology: Principles and Explorations*. 6th Edition, John Wiley and Sons, Inc.

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MAHATMA GANDHI UNIVERSITY

SBS M P C 03: CELL BIOLOGY, GENETICS & EVOLUTION

SchoolName	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**CELL BIOLOGY, GENETICS & EVOLUTION**					
Type of Course	Core					
Course Code	**SBS M P C 03**					
Names of Academic Staff & Qualifications	**Dr J G Ray, Dr Keerthi T R, Dr Jayachandran K, Dr. Linu Mathew**					
Course Summary & Justification	is course on Cell Biology and Genetics deals with the frontier areas of basic biology e objective of the course content is to create a sound awareness about the current developments taking place in different fields of cell biology and genetics The course content is designed with a view to augment CSIR/UGC syllabus					
Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of cell biology and genetics**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**uild a perspective on current developments in the fields of cell biology, genetics and evolution and the cellular level organization of organisms**	E	
2	**mpare and analyze the processes of cell cycle, cell division, cell differentiation and cell death and analyze the relationship between cell cycle, ageing, cell death and cancer**	U/ An	
3	**Explain the processes, laws, and theories related to inheritance and evolution**	R	
4	**Perform genetic mapping based on data supplied**	S	
5	**aluate the behavior of genotypes and alleles in natural populations**	E	
6	**Communicate effectively about a given topic in cellbiology/ genetics/ evolution both verbally and in writing**	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Cell and its constituents: Cell constituents -** Mitochondria, Chloroplast, Endoplasmic Reticulum Golgi complex, Peroxisomes, Lysosome, Ribosome, Nucleus, Nucleolus, Chromosomes, Nucleosomes, Histones, Genome, Genomics, Proteomics. **Cell cycle and Cancer: Cell cycle -** Different stages, variations, checkpoints, regulations of cell cycle, maturation Promoting factor, cells, cyclins , ubiquitin, protein ligases, Anaphase Promoting complex, inhibitors of CdK, growth factors and D cyclins. Rb protein and E2F transcription factors. **Cancer -** Stages in cancer development, causes, properties of cancerous cells, tumor Viruses, oncogenes, functions of oncogene products, oncogene and signal Transduction , oncogene and G proteins, oncogene and cell survival, Tumor Suppressor gene, functions of tumor suppressor gene products, Diagnosis ,prevention and treatment of cancer	0.5	10
2	**Cell Differentiation**-Stages of development, regulation of development, cascade control/ Differentiation in Drosophila, maternal, Segmentation and homeotic Genes, Genetic control of embryonic development, Bi thorax mutant, Antennapediac mutant ,Hemeobox **Aging** Process of aging, theories of aging, Arking's contribution Oxidative stress, Telomere problem, DNA repair defects. **Cell Death** Necrosis and Apoptosis, Differences between necrosis and Apoptosis, stages in Apoptosis, mitochondrial damage DNA ladders, transglutaminase activity, programmed cell death in *Ceanorhabdtis elegans* CED 3, CED 4, CED 9 and their roles in Apoptosis Bax, Bid, Bcl2 protein	0.5	10
3.	**Classical Genetics:** Genetics, the evolution of the subject through pre mendelian, Mendelian and post Mendelian Peroids. Mendelism – the basis principles of inheritance, gene interactions – allelic and no allelic. Environment and gene expression, penetrance and expressivity. Multiple alleles and polygenic inheritance, Heritability and genetic advance **Evolution:** Origin of the universe and origin of life; concept of Oparin, Miller-Urey Experiments; Evolution of Prokaryotes - origin of eukaryotic cells - Margulis Endosymbiotic theory; **Geological Timescale:** Tools and techniques in estimating evolutionary time scale; **Theories of evolution of life**: Pre-Darwinian concepts – Lamarkism, Darwinism – major concepts - variation, adaptation, struggle, fitness and natural selection, Neo-Darwinian theories – theories of speciation – allopatric and sympatric speciation - Rose Mary and Peter Grant (Molecular evolution in Darwinian finches) - Neutral Theory of Molecular Evolution.	1.0	20
4	**Chromasome genetic mapping ,Organelle Genetics and Population Genetics:** Linkage and linked genes with special reference to inheritance, Chromosome mapping with three - point test crosses. Organelle Genetics and cytoplasmic inheritance. Population Genetics – types of gene variations, Measuring genetic variations, Hardy Weinberg principle and its deviations. Medical genetics - an	0.5	10

	introduction		
5	**Genetic System in Microbe, Yeast and Neurospora:** Plasmids & bacterial sex. Types of plasmids. Plasmids copy number and incompatibility, Replication of plasmid. Plasmid a cloning vector. Episomes. Transposable element-IS element and transposon, Integrons and Antibiotic resistance cassettes, Multiple antibiotic resistant bacteria, Mu-virus. Gene mapping in Bacteria. Bacteriophage genetics-Plaque formation & phage mutants, genetic recombination in lytic cycle. Genetic system in Yeast & Neurospora.	0.5	10
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Jonathan B (2016) Principles of Evolution, Garland Science, Taylor and Francis
2. Strickberger M W (2015) Genetics 3rd Edition, Pearson
3. Genetics a conceptual approach. 6th edition. Benjamin Pierce, Macmillan Learning, New York
4. The Cell-A Molecular approach, Fifth edition, Geoffrey M Cooper and Robert E .Hausman. , ASM Press ,Washington DC

Further Reading:

1. Principles of Genetics, Snustad, Simmons and Jenkins, John Wiley And Sons Inc
2. Genetics, Robert Weaver and Philip Hendricks, WH.C. Brown Publishers, Iowa
3. Introduction to Genetic Analysis, Griffiths, Wessler, Lewontin, Gelbart,Suzuki and Miller, Freeman's and Co, New York
4. REA's Problem Solvers in Genetics, Research Education Association,61, Ethel Roadwest, New Jersey
5. Cell and Molecular Biology by Gerald Karp,7th Edition,
6. Cell and Molecular Biology by De Robertis E.D.P, 8th Edition

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	**MAHATMA GANDHI UNIVERSITY** **SBS M P C 04: BIOPHYSICS AND BIOSTATISTICS**

SchoolName	**School of Biosciences**
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**
Course Name	**BIOPHYSICS AND BIOSTATISTICS**

Type of Course	Core					
Course Code	**SBS M P C 04**					
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S**					
Course Summary & Justification	This course is to introduce interdisciplinary Biophysics area, its scope and its importance The objective of the course is to give an insight into the basic concepts of thermodynamics, importance of basic biophysical phenomena, conformation and conformational changes, interaction of protein with other molecules and basic knowledge about radiation, its interaction with matter and its applications. The course content is to familiarize the basic concepts of biostatistics and its importance in research area of Life sciences The course content is designed with a view to augment CSIR/UGC syllabus					
Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basics of Biophysics and Biostatistics					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.

1	plain the scope and importance of biophysics	E	
2	scribe the concepts of thermodynamics and applications of basic biophysical phenomena.	U/ An	
3	Narrate the conformation and interaction of proteins and nucleic acids	R	
4	Explain the electromagnetic radiation, its interaction with matter and applications.	S	
5	form the retrieval of biological information by using structural and sequence databases	E	
6	Explain the basic concept of biostatistics and analyze, interpret statistical softwares and to do statistical design for their research	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Biophysical phenomena and Thermodynamics of biomolecular interactions**: Scope and definition of Biophysics, Principle and biological importance of Osmosis, Electroosmosis, osmotic pressure, osmotic equilibrium, Donnan equilibrium, Diffusion, Sedimentation, Filtration, Surface tension, Dialysis, Adsorption and Colloids. Laws of thermodynamics, Enthalpy, Entropy, Free energy, Redox reactions, Redox potential and its calculation by Nernst equation, examples of redox reactions in biological system.	0.5	10
2	**Structural Biophysics and computational biology**: The molecular interactions between proteins and nucleic acids: DNA- protein interaction and RNA- protein interactions, DNA-binding motifs: Helix-turn-Helix motif, Zn fingers, Helix-loop helix motifs and Leucine zippers. Molecular forces: Hydrogen bonding, hydrophobic interactions, Dipole interactions: charge-dipole interactions, induced dipoles, steric repulsion, Vander waals force in biomolecules, Structural and Sequence databases, Alignment algorithms; Retrieval of biological information from widely used resources: NCBI and PDB, Molecular modelling and Structure based drug designing.	0.5	10

3.	**Radiation Biophysics**: Electromagnetic spectrum, Ionizing and non ionizing radiation. Properties and biological effects of ultraviolet radiation, infrared and microwave radiations. Radioactivity, Interaction of radiation with matter. Units of Radiation. Biological effects of radiation. Applications of ionizing and non-ionising radiations in industry, agriculture and research. Radiation hazards.	1.0	20
4	**Introduction to Biostatistics:** Scope of Biostatistics, probability and probability distribution analysis. Variables in biology- collection, classification and tabulation of data- graphical and diagrammatic representation- scatter diagrams, histograms- frequency polygon- frequency curve-logarithmic curves. Descriptive statistics- measures of central tendency, Arithmetic mean, median, mode, geometric mean, harmonic mean. Measures of dispersion, standard deviation, standard error, variance, coefficient of variation. Correlation and Regression	0.5	10
5	**Test of significance:** Basic idea of significance test- hypothesis testing, levels of significance. Testing of single mean, double mean, single proportion, double proportion in large sample. Testing of single mean, double mean and Paired- t in small sample. ANOVA- One way and Two way; Chi-square test of goodness of fit and Chi-square test of independence, comparison of means of two samples, three or more samples. Fundamentals of field experiments- randomization, replication and local control. CRD and RBD. Statistical packages	0.5	10
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Proteins, Structure and molecular properties, Thomas E Creighton
2. Fundamentals of Biostatistics: Irfan.A. khan, Atiya Khanum, Ukaaz publications
3. Principles of Biostatistics: Marcello Pagano, Kimberlee Gauvreau, Duxbury Press
4. Biochemistry: Donald Voet and Judith G Voet, Wiley Publications

Further Reading:

5. Biophysics-Hoope W etal
6. Biophysics-Volkenstain M.V
7. Molecular Biophysics- Volkenstain M.V
8. Introduction to thermodynamics of irreversible process-John Wiley
9. Statistical methods in Biology- Briley N.J.T
10. Introduction to Biophysics-Sokal R.R & Rohl F.J
11. Biostatistics: Pardeep.K.Jasra, Gurdeep Raj, Krishna prakashan Media.(P) Ltd
12. Bloomfield, V. (2009) Computer Simulation and Data Analysis in Molecular Biology and Biophysics. Springer

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MAHATMA GANDHI UNIVERSITY

SBS M P C 05: PHYSIOLOGY

SchoolName	**School of Biosciences**
Programme	**MSc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**
Course Name	**PHYSIOLOGY**
Type of Course	Core
Course Code	**SBS M P C 05**

<table>
<tr><td>Names of Academic Staff & Qualifications</td><td colspan="6">Dr. R. Harikumaran Nair
MSc, PhD</td></tr>
<tr><td>Course Summary & Justification</td><td colspan="6">is course is designed to provide an overview of human physiology. Course topics will include the various systems of the body, functions of each system, and interrelationships to maintain the internal environment. The course also provides inputs to physiological stress and adaptive strategies to overcome stress</td></tr>
<tr><td>Semester</td><td colspan="6">First</td></tr>
<tr><td rowspan="2">Total StudentLearning Time (SLT)</td><td>Learning Approach</td><td>Lecture</td><td>Tutorial</td><td>Practical</td><td>Others</td><td>Total LearningHours</td></tr>
<tr><td>Authentic learning
Collaborative learning
Independent learning</td><td>60</td><td>20</td><td>0</td><td>40</td><td>120</td></tr>
<tr><td>Pre-requisite</td><td colspan="6">Basics Knowledge in Biology</td></tr>
</table>

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	udents should be capable of effectively communicating how the human body works	U	
2	udents should be able to explain interrelationships among molecular, cellular, tissue, and organ functions in each system	E	
3	Students should be able to describe the interdependency and interactions of the systems	A	
4	Students should be able to explain contributions of organs and systems to the maintenance of homeostasis	A	
5	udents should be able to identify causes and effects of homeostatic imbalances	E	
6	Able to gain the approaches used to study various functional systems of the human body and physiologic adaptation	I	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	The system as a basic unit in physiology: different systems in physiological process, interaction of different systems in normal and stress conditions, homeostasis, Neuro-Musculo-Skeletal systems: brain and peripheral nervous systems, neurotransmitters, synapse, neuro-muscular junction, musculoskeletal systems	0.5	10

2	Cardio-Pulmonary & Renal Physiology: Anatomy and general function of heart, blood and hemodynamic, blood pressure, heart rate, cardiac cycle, cardiac output, electrocardiography, echocardiography; anatomy of the respiratory system, principles of respiratory mechanisms, respiratory rate, lung volumes, oxygen uptake, lung function tests, gas transport; anatomy of the excretory system, nephron, glomerular filtration rate, urine formation, renal clearance test, renal regulation of electrolytes, dialysis	1	20
3.	Principles of endocrinology: Role of hormones for maintenance of the internal environment, hormone transport in blood, mechanism of hormone action, hormone metabolism and excretion, types of endocrine disorders, hypothalamus and pituitary, thyroid, adrenal glands, endocrine control of growth, sex hormones, pancreatic hormones, neurohormones	0.5	10
4	Gastrointestinal Physiology & Nutrition: Gastrointestinal structure, food digestion, and absorption, gastrointestinal hormones, central control of gastrointestinal functions, pathological situations of gastrointestinal functions. role of liver and bile in gastrointestinal functions.	0.5	10
5	Stress physiology: Stress-responses, the role of the hypothalamic-hypophyseal-adrenal axis, oxidative stress and mechanism, effect of stress-inducing and anti-stress agents, cardio-respiratory responses during high altitude acclimatization, stress-induced diseases, and remedy, Human tolerances to stresses in space including space flight: Physiological adaptation to space flight, physiology in deep-sea diving and other high-pressure operations	0.5	10
	Total Credits of the Course	3	60

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading

1. Vander's Human Physiology- The mechanism of body function. Widmaier, Raff & Strang

2. Textbook of Medical Physiology. Arthur.C. Guyton& John.E. Hall

3. Physiological basis of Medical Practice. John.B. West 4. Endocrinology- Mac E Hadley
Further Reading: Review of Medical Physiology- Ganong, William F Biochemistry and Physiology of the cell. An introductory text second edition- Edwards, N. A Hassall, K.A Notebook of medical physiology: endocrinology, with aspects of maternal, fetal and neonatal physiology- Hawker, Ross Wilson Human Physiology: an integrated approach- Silverthorn, Dee Unglaub Principles of anatomy and physiology- Tortora, Gerald J Derrickson, Bryan 6.Textbook of Endocrine Physiology- Griffin, James E; Ed. Ojeda, Sergio R;Ed

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	**MAHATMA GANDHI UNIVERSITY**
	SBS M P C 06: LABORATORY COURSE 1

SchoolName	**School of Biosciences**
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**
Course Name	**LABORATORY COURSE 1**
Type of Course	Core
Course Code	**SBS M P C 06**
Names of Academic Staff & Qualifications	1. **Dr. Anie Y – M. Sc (Biochemistry), PhD**
Course Summary & Justification	The course is designed to develop in students the essential skills to perform the basic biochemical assays, qualitative analysis of biomolecules and techniques for the separation of biomolecules. This will enhance the practical abilities of the students to carry out the analysis of biomolecules.

Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg. Authentic learning Collaborative learning Independent learning	10	20	150	30	210
Pre-requisite	General idea on reagents and solvents					

COURSE OUTCOMES (CO)

CO No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**
1	To prepare reagents, buffers and other solutions in required concentrations and required pH.	Ap	

2	To extract and estimate different bio-molecules (sugar, cholesterol, and proteins) in biological samples	Ap/S	
3	To identify the different components in a mixture of carbohydrates	S	
4	To detect the presence of albumin, casein and gelatin in biological samples	S	
5	To perform separation by Paper and Thin layer chromatography	S	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hours
1	Preparation of solutions: Percentage solutions, Molar solutions, Normal solutions, Dilution of Stock solutions, Preparation of buffers using the Henderson Hasselbach equation	0.5	10
2	Spectrophotometric experiments: Verification of Beer Lambert's law, Determination of UV-Visible spectrum of compounds, Determination of Concentration of molecules from Molar Extinction Coefficient values Extraction of Polysaccharides (Starch/Glycogen), Proteins, and Lipids from appropriate sources and their estimations. Estimations: Estimation of reducing sugars by Dinitrosalicylic acid method, Estimation of proteins (Biuret and Lowry's methods), Estimation of Methionine by Nitroprusside method, Estimation of Cholesterol by Zak's method.	1	60
3.	Qualitative analysis of Carbohydrate mixtures (a combination of polysaccharide, disaccharide and monosaccharide) following systematic scheme for analysis. (Starch, dextrin, glycogen, glucose, fructose, xylose, galactose, sucrose, maltose, lactose) Qualitative analysis of proteins- Albumin, casein, gelatin	1	80
4	Chromatographic techniques: Separation of amino acids by Paper chromatography (Descending or Ascending), Separation of Plant pigments by Thin layer chromatography	0.5	30

Total Credits of the Course	3	

Books for Reference

Compulsory Reading:

1. Introductory Practical biochemistry, S. K. Sawhney & Randhir Singh (eds) Narosa Publishing House, New Delhi, ISBN 81-7319-302-9, p 195 – 303
2. Standard Methods of Biochemical Analysis, S. K. Thimmaiah (ed), Kalyani Publishers, Ludhiana ISBN 81-7663-067-5, p 12 - 182.

Further Reading:

3. Hawk's Physiological Chemistry, Bernard L. Oser (ed) TATA McGRAW Hill Publishing Company LTD, New Delhi, p 60 – 127, 1317- 1334
4. Experimental Biochemistry: A Student Companion, Beedu Sasidhar Rao & Vijay Deshpande (ed), I.K International Pvt. LTD, New Delhi ISBN 81-88237-41-8, p 13- 17, p 49 - 72
5. Practical Biochemistry, R.C. Gupta & S. Bhargava (eds) CBS Publishers and Distributors, New Delhi, ISBN 81-239-0124-0 p 9 – 27
6. Practical Clinical Chemistry, Harold Varley, CBS Publishers and Distributors, New Delhi,

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Explicit Teaching, interactive Instruction: Active co-operative learning and skill development, Demonstrations, Group Assignments, Authentic learning, Library work and Group discussion, Preparation of experiment design and reports
Assessment Types	**Mode of Assessment** **C. Continuous Internal Assessment (CIA)** Assessment of the performance of student in the lab- 10 marks Internal Test -20 marks Project report (student needs to perform experiments on a specific project and report should be prepared)– 10 marks **D. Semester End examination – 60 marks**

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MAHATMA GANDHI UNIVERSITY

SBS M P C 07 LABORATORY COURSE-2

SchoolName	**School of Biosciences**
Programme	**MSc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**
Course Name	**LABORATORY COURSE-2**
Type of Course	Core
Course Code	**SBS M P C 07**
Names of Academic Staff & Qualifications	**Dr. R. Harikumaran Nair** MSc, PhD
Course Summary & Justification	e purpose of this laboratory course is to provide the student with the opportunity to observe many physiological principles. The course is

	designed to understand the mechanisms related to cardiovascular and respiratory functions.					
Semester	First					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	5	5	50		60
Pre-requisite	**Basics Knowledge in Biology**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	pply appropriate safety standards in laboratory	A	
2	quire laboratory skills in haematology, cardiovascular and respiratory physiology	S	
3	Appropriately utilize laboratory equipment, such as microscopes, dissection tools, general labware, physiology data acquisition systems	S	
4	mmunicate results of scientific investigations, analyse data, and formulate conclusions	C	
5	udents should be able to identify cell structure	U	
6	Work collaboratively to perform experiments	I	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Haematology** i) Determination of haemoglobin concentration ii) Enumeration of formed elements- red blood cells & white blood cells iii) Study of blood smear for the differential count and cell morphology iv) Erythrocyte sedimentation rate v) Determination of the bleeding time vi) Determination of clotting time	1	20
2	**Respiratory physiology- Pulmonary function testing** i) Demonstration on the recording of tidal volume ii) Demonstration on the recording of vital capacities iii) Demonstration on the recording of inspiratory & expiratory flow rates	1	20

3.	**Cardiovascular physiology- Electrocardiography** i) Demonstration on ECG recording- human or animal model ii) Identification of ECG waves iii) Calculation of heart rate from ECG	1	20
	Total Credits of the Course	3	60

Teachingand LearningApproach	**Laboratory Procedure (Mode of transaction)** Direct Instruction: lecture, Explicit Teaching, Demonstration, Hands on experimental sections, Skill acquisition by laboratory training
Assessment Types	**Mode of Assessment** E. Continuous Internal Assessment (CIA) 1. Internal Laboratory Skill Tests of maximum 20 marks 2. Seminar Presentation – Laboratory material and methods Maximum marks 10 3. Write a detailed report on instrumentation – 10 marks F. **Semester End Practical examination – 60 marks**

REFERENCES

1. Medical Laboratory Technology-A Procedure Manual for Routine Diagnostic Tests- Kanai L Mukherjee
2. Pocket Guide to Spirometry- David P Johns and Rob Pierce
3. Spirometry in Practice- A practical guide to using spirometry in primary care- Dr. David Bellamy, British Thoracic Society COPD consortium.
4. ECGs made easy- Barbara J Aehlert

Further Reading:

ECG Assessment and Interpretation- Cascio, Toni

ntroduction to medical laboratory technology- Baker, F J Silverton, R E Practical haematology- Dacie, John V Lewis, S.M

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SECOND SEMESTER

MAHATMA GANDHI UNIVERSITY

SBS M P C 08 IMMUNOLOGY

SchoolName	School of Biosciences					
Programme	M.Sc. Microbiology/Biotechnology/Biochemistry/Biophysics					
Course Name	**IMMUNOLOGY**					
Type of Course	Core					
Course Code	**SBS M P C 08**					
Names of Academic Staff & Qualifications	Dr.Radhakrishnan E.K. M.Sc., Ph.D					
Course Summary & Justification	is course on Immunology deals with various mechanisms and processes involved in the defense responses. This course is an important branch of life science. Human body has different lines of defense to fight against pathogens. The content in this course has been designed with an objective to provide detailed understanding on the process and mechanisms involved in the defense responses. Understanding on the functioning of immune system is highly essential for a student to explore its theoretical and practical aspects for the benefit of society.					
Semester	Second					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding on defense responses Knowledge in any branch of Life science					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1.	Students will able to understand and explain basic principles of immunology	R/U	
2.	Students will able to learn the recent advances in	R/U	

	immunology		
3.	Students will able to analyse the clinical importance of immunological reactions	U/ An	
4.	Students will become able to identify the correlation between immunological abnormalities and health status of humans	U/An	
5.	Students will get theoretical and technical know-how for the laboratory diagnosis of infectious diseases	C/S	
6.	Students can apply the knowledge and skills for clinical and diagnostic applications	A/S	
**Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Infection, Source and methods of transmission, Immunity- Types of immunity. Mechanisms of innate immunity, PAMPs, pattern recognition receptors, types, scavenger receptors and toll – like receptors, Phagocytes and Phagocytosis, Organs and cells with immune functions. Lymphocytes and lymphocyte maturation. PAMPs and PRRs in plants	0.5	10
2	Antigens, Epitopes and paratopes, B-cell and T-cell epitope, Antigenicity and Immunogenicity, Antibodies, Immunoglobulin – structure, classes and functions. Genetic basis of antibody diversity, Organization and Expression of Immunoglobulin Genes, V(D)J rearrangements; recombination signal sequences and their role, somatic hypermutation and affinity maturation Antigen-antibody reactions, Agglutination, Precipitation,	1.0	20

	Immunoflourescence, Complement fixation, Radioimmuno assay, ELISA, Western blotting		
3.	Immune response- Humoral and cell mediated, Receptors on T and B cells for antigens, MHC, TCR- mediated signalling, Signal transduction pathways associated with T-cell activation, Signal transduction by activated B- cell receptor, Antibody production, Primary and secondary immune response, Factors influencing antibody production, Clonal selection theory, Monoclonal antibodies – production and application, Antibody engineering. Complement system, Complement activation, Biological effects of complements, Antigen processing and presentation, Activation of T-cells, T cell function, Cytokines. Human microbiome and immunity	0.5	10
4	Immunology of organ and tissue transplantation, Allograft reaction and GVH reaction, Factors influencing allograft survival, Immunology of malignancy, Tumor antigens, Immune response in malignancy, Immunotherapy of cancer, Immunohematology, ABO and Rh blood group system, Immunology of blood transfusion, Hemolytic disease of new born	0.5	10
5	Immunological Tolerance, Autoimmunity, Mechanisms of autoimmunization, Autoimmune diseases. Inflammation, Hypersensitivity – immediate and delayed reactions, Clinical types of hypersensitivity, Immunodeficiency diseases, Immunoprophylaxis, Vaccines –types of vaccines, DNA vaccine, recent trends in vaccine development.	0.5	10
	Total Credits	**3**	

Teaching And Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction, Active co-operative learning, Seminar, Group Assignments Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar - Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. Semester End examination – 60 marks

REFERENCES

Compulsory Reading:

1. Immunology - Thomas J. Kindt, Barbara A. Osborne, Richard A. Goldsby, and Janis Kuby, W H Freeman and Co., 2013
2. Immunobiology - Charles A. Janeway Jr.,Paul Travers, Mark Walport and Mark J. Shlomchik, Garland Publishing., 2016

Further Reading:

3. Essential Immunology - Ivan M. Roitt and Peter J delves, Blackwell Publishing, 2016

4. Essential Clinical Immunology – Helen Chappel and Mansel Haeney, ELBS/Blackwell Scientific Publications, 2014

5. Introduction to Immunology – John W, Kimball Maxwell, Mac Millan International Edition, 1990

6.Text book of Microbiology – R. Ananthanarayanan and C K Jayaram Panicker. Orient Longman,2013

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School Name	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**Molecular Biology and Genetic Engineering**					
Type of Course	Core					
Course Code	**SBS M P C 09**					
Names of Academic Staff & Qualifications	**Dr Keerthi T R, Dr Jayachandran K, Dr. Linu Mathew**					
Course Summary & Justification	1. Molecular Biology and Genetic Engineering is one of the most dynamic and attractive courses in all branches of applied life sciences 2. The syllabus content in this paper is designed with an objective to train the students in both theoretical and practical aspects of the subject 3. This will also enable the students to get an idea about the latest developments taking place in this subject					
Semester	second					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basics of cell and molecular biology, Basics of tools and techniques of genetic engineering					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	completing this course the students will be able to plain the processes of replication, transcription and translation and analyse the importance of these processes in health and disease	E	
2	plain the concepts of gene regulation in prokaryotes and RNA world	R/ E	
3	nalyse the use of different tools and techniques of gene cloning in E coli and explain the applications of DNA technology	U	
4	ility to develop a protocol for cloning a gene from a selected organism	A	
5	ility to explain verbally and orally the concepts of molecular biology and genetic engineering	E	
6	Ability to write a research proposal based on the concepts discussed in the course	An/ C	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Replication–** Process of DNA replication, Semiconservative, discontinuous uni and bidirectional , Okazaki fragments, DNA polymerases in eukaryotes and prokaryotes , Klenov fragment, modes of replication, theta, rolling circle, d-loop replication, Primasome, SSB, Helicase, Ligase, methylation and control, repetitive DNA sequences, minisatellite, microsatellite, DNA protein interation DNA Linking number and topoisomerase, Inhibition of replication.	0.5	10
2	**Transcription**. Process of transcription, stages in transcription, RNA polymerases in prokaryotes and enkaryotes, sigmafactor in prokaryotes, Rho dependant and Rho independent termination. Enhancers, Transcription factors in Eukaryotes, Differences in transcription between prokaryotes and Eukaryotes, post transcriptional modifications-Polyadenylation, capping, r-RNA processing, Splicing-Spliceosome, lariat structure, Group 1, II and III Introns Rihozyme, Importance of ribozyme, properties, application, RNase P, RNAse III, RNAse H. monocistonic and polysistronic m-RNA, Joint transcript of r-RNA and t-RNA in prokaryotes and their processing, Transplicing, alternate splicing, inhibitors of Transcription. Molecular mechanism of gene regulation in prokaryotes-Transcriptional regulation in prokaryotes; Inducible & repressible system,+ & -ve regulation; Operon concept, structure of operon, Lac, Trp, Arc operon, Catabolic repression, Atteunation. Role of Hormones in gene regulation. RNA World, RNA based technology- Molecular mechanism of Ribozyme, Antisense RNA, SiRNA, MicroRNA, Ribozwitches & their applications; Telomerase structure and function, Nucleic acid as therapeutic agent	0.75	15
3.	**Translation:** Process of translation. Stages in translation, genetic code, properties, wobble hypothesis, eukaryotes and prokaryotes ribosomes, m-RNAs, t-RNAs, aminoacyl t-RNA synthatases, protein factors initiation complex, peptidyl transferase, releasing factors, differences between prokaryotic and eukaryotic systems, inhibition of translation. Post translation modification by cleavage, self assembly assisted self assembly chaperones, acylation, phosphorylation, acetylation and glycosylation, Histone acetylation and deacetylases, chromosome remodeling complex. Intein splicing. Protein targeting, cotranslational import, post translational import, SRP- structure and function, Blobel's concept, Lysosome targeting, M6P address Glycosylation core glycosylation terminal glycosylation, Dolichol phosphate.	0.5	10
4	**Tools and techniques for genetic Engineering:** History of rDNA Technology ,Cohen And Boyer Patents, Isolation of DNA and RNA from different sources, enzymes used in genetic engineering with special reference to restriction enzymes, ligases, and other DNA modifying enzymes. End modification of restriction fragments, vaccinia topoisomerases mediated ligation of DNA, TA cloning, and homopolymer tailing Vectors for E coli with special reference to plasmid vectors (pSC101, pBR322,pUC,their	1	20

	development, features and selection procedures),direct selection plasmid vectors, low copy number plasmid vectors, runaway plasmid vectors, Bacteriophages (λ and M13) with special reference to Charon phages, λEMBL, λWES λB', λ ZAP- their development, features, selection procedures, *in vitro* packaging mechanisms for phage vectors, cosmids, features, advantages and cosmid cloning schemes, phagemids with special reference to pEMBL, pBluescript, pGEM3Z , pSP64, pcDNA, pLITMUS Construction of genomic libraries and cDNA libraries, procedures for recombinant selection and library screening, PCR enzymes, types of PCR, primer design, real time PCR, RTPCR, Nested PCR, Inverse PCR, Assymmetric PCR, applications of PCR Cloning, Chemical synthesis of DNA, DNA sequencing:- plus and minus sequencing, Sangers dideoxy sequencing, Maxam and Gilberts method. Advanced sequencing procedures: – pyrosequencing, Illumina, ABI / SOLiD and their applications		
5	**Appications of Genetic Engineering:** Applications of transgenic Technology Improving quality, quantity and storage life of fruits and vegetables. Plants with novel features, Engineering metabolic pathways, Pharming. Animal cloning, Ethics of cloning. Applications of Molecular Biology in forensic sciences, medical science, archeology and paleontology	0.25	5
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** G. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks H. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Principles of gene manipulation – Old and Primrose, Blackwell Scientific publishers, Edn.5th

2. Cell and Molecular Biology by Cooper

Further Reading:

7. Principles of gene manipulation – Old and Primrose, Blackwell Scientific publishers, Edn.5th
8. Principles of gene manipulation – Old, Primrose, and Twyman, Blackwell Scientific publishers, Edn. 6th
9. Principles of gene manipulation – Old, Primrose, and Twyman Blackwell Scientific publishers, Edn 7th
10. Molecular biotechnology, Principles and Applications of Recombinant DNA, Glick Pasternak and Patten, 4th edition ISBN 978-1-55581-498-4 Wiley International Publishers
11. From gene to genomes – Concepts and applications of DNA technology Jeromy W Dale and Malcom von Shantz , John Wiley and sons
12. Principles of plant biotechnology: An introduction to genetic engineering in plants – SH Mantell
13. Cell and Molecular Biology by Gerald Karp, Academic Press
14. Cell Biology by DeRobertis
15. Genes-Benjamin Lewin

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	MAHATMA GANDHI UNIVERSITY
	SBS M P C 10: METABOLISM AND BIOENERGETICS

SchoolName	**School of Biosciences**
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**
Course Name	**METABOLISM AND BIOENERGETICS**
Type of Course	Core
Course Code	**SBS M P C 10**
Names of Academic Staff & Qualifications	2. **Prof. M S Latha – M. Sc (Biochemistry), PhD** 3. **Dr. Anie Y – M. Sc (Biochemistry), PhD** 4. **Guest faculty – M. Sc (Biochemistry), PhD**
Course Summary & Justification	The course is designed to get a deep knowledge of metabolic processes taking place in the biological systems and their regulation, which is needed to understand the more specialised areas of Biochemistry.

Semester	Second					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg. Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding of chemical groups and bonding; basics of cell biology and physiology					

COURSE OUTCOMES (CO)

CO No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**
1	be able to categorize, differentiate and predict the fates of different biomolecules via the metabolic pathways.	U/A	
2	To draw conclusions on the energetics of the metabolic pathways and to find out the variations in ATP generation during physiological and pathological conditions	A	
3	analyse different methods of regulation of the metabolic pathways.	A/An	
4	Describe the different steps involved and the importance of metabolomics in toxicity analysis and health management	A	
5	To describe the structure and functions of vitamins and hormones	U	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	**Module Content**	**Credits**	**Hours**
1	**Metabolic Pathways:** Detailed study on the catabolic pathways & anabolic Pathways -Carbohydrate, Protein, Amino acid and Nucleic acid metabolic pathways.	1	17

2	**Bioenergetics:** Functional significance of the mitochondrial respiratory chain and oxidative phosphorylation, Electron transport chain: structural components of the chain, complexes, free elements; Structure and functional properties of cytochromes, ferro-sulphurated proteins and CoQ; Generation of the electrochemical proton gradient: Chemiosmosis ATP synthesis- Proton flow through ATP synthase, Rotational catalysis. Inhibitors and uncouplers	0.75	13
3.	**Regulation of metabolism:** Hormonal and Allosteric regulation of pathways in carbohydrate, lipid, nucleotide, amino acid and protein metabolism; Coordinated regulation of opposing metabolic pathways; Regulation of mitochondrial electron transport and oxidative phosphorylation.	0.5	11
4	**Signal Transduction:** intracellular receptor and cell surface receptors signaling: Cyclic AMP-dependent protein kinase; Cyclic GMP-dependent protein kinase; Protein kinase C; Ca^{2+} -calmodulin-dependent protein kinases ; AMP-dependent protein kinase ; Receptor tyrosine kinases; Protein kinase B; Cytokine activation of the JAK'/STAT pathway; Cell cycle control; Receptor serine/threonine kinases; Other protein kinases ; Phosphoprotein phosphatases; Cancer Pathways: MAPK, P13K, TP53 network, NF*k*B pathways; Signalling by TGF β factor , STAT factor	0.5	11
5	**Metabolomics:** Introduction to origins of metabolomics; define terms: Metabolite, Metabolome, Metabonomics; Analytical techniques in study of Metabolomics (Principle & Methodolgy): Separation methods: Gas Chromatography, HPLC, Capillary Electrophoresis; Detection Methods: Mass spectroscopy, NMR. Applications of Metabolomics in toxicity assessment/ toxicology, diagnostics and health Screening	0.25	8
	Total Credits of the Course	3	

Books for Reference

Compulsory Reading:

1. Principles Of Biochemistry, 4/e (2006) by Robert Horton H , Laurence A Moran, Gray Scrimgeour K **Publisher:** Pearsarson **ISBN:** 0131977369, **ISBN-13:**9780131977365, 978-0131977365
2. Biochemistry 6th Edition (2007) by Jeremy M.berg John L.tymoczko Lubert Stryer **Publisher:** B.i.publicationsPvt.Ltd **ISBN:**071676766X **ISBN-13:** 9780716767664, 978-716767664
3. Lehninger Principles of Biochemistry, Fourth Edition by David L. Nelson Michael M. Cox
13. Publisher: W. H. Freeman; Fourth Edition edition (April 23, 2004) ISBN-10: 0716743396 ISBN-13: 978-0716743392

Further Reading:

- **E.S. West, W.R. Todd, H.S. Mason and J.T. van Bruggen,** A **Text Book of Biochemistry**, Oxford and IBH Publishing Co., New Delhi, 1974
- Biochemistry [with Cdrom] (2004) by Donald Voet, Judith G. Voet **Publisher:** John Wiley & Sons Inc **ISBN:** 047119350X **ISBN-13:** 9780471193500, 978-0471193500

- Principles Of Biochemistry (1995) by Geoffrey L Zubay, William W Parson, Dennis E Vance **Publisher:** Mcgraw-hill Book Company – Koga **ISBN:**0697142752 **ISBN-13:** 9780697142757, 978-0697142757
- Biochemistry (2008) by Rastogi **Publisher:** Mcgraw Hill **ISBN:**0070527954 **ISBN-13:** 9780070527959, 978-0070527959

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** **I. Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks **J. Semester End examination – 60 marks**

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	MAHATMA GANDHI UNIVERSITY
	SBS M P C 11 BIOPHYSICAL TECHNIQUES AND BIOINSTRUMENTATION

SchoolName	**School of Biosciences**					
Programme	**MSc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**BIOPHYSICAL TECHNIQUES AND BIOINSTRUMENTATION**					
Type of Course	Core					
Course Code	**SBS M P C 11**					
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S**					
Course Summary & Justification	This course is designed to introduce different techniques used in life sciences This course gives knowledge of the principle of operation and design of scientific instruments It attempts to render a broad and modern account of scientific instruments					
Semester	Second					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basics of Biophysics and Biostatistics					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To explain the methods used for gaining information about biological systems on an atomic or molecular level.	E	

2	To describe different spectroscopic techniques	U/ An	
3	To perform various biophysical fractionation and separation of biomolecules	R	
4	To describe how to perform electrophoretic techniques	S	
5	describe the procedures and applications of hydrodynamic techniques	E	
6	To perform different microscopic techniques	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Spectroscopic techniques**: Basic principles, nature of electromagnetic radiation, Interaction of light with matter, Absorption and emission of radiation; Atomic & Molecular Energy levels, Electronic, vibrational and Rotational spectroscopy of molecules, transition and selection rules; Atomic & Molecular spectra. Principle, Instrument Design, Methods & Applications of UV-Visible spectroscopy, Infrared spectroscopy, Raman Spectroscopy, Fluorescence spectroscopy, Nuclear magnetic Resonance Spectroscopy.	0.5	10
2	**Physicochemical Fractionation techniques**: Principle, Instrument Design, methods and Applications of all types of Adsorption and Partition Chromatography- Paper chromatography, Thin layer chromatography, High Performance Thin layer Chromatography, Gel filtration chromatography, Affinity chromatography, Ion-exchange chromatography, High Pressure Liquid Chromatography. Reversed phase chromatography, Hydrophobic interaction chromatography, Chiral chromatography, Counter current chromatography, Fast protein liquid chromatography, Two dimensional chromatography.	0.5	10

3.	**Electro analytical techniques and Hydrodynamic Techniques**: Principle, Electrophoretic mobility (EPM) estimation, factors affecting EPM, Instrument design & set-up, Methodology & Applications of Free and zone Electrophoresis – Paper electrophoresis, Gel electrophoresis, Poly Acrylamide gel electrophoresis, SDS PAGE, Capillary electrophoresis, Isoelectric focusing, Potentiometry, pH meter, Conductometry. Centrifugation & Ultracentrifugation-Basic principles, Forces involved, RCF Centrifugation, techniques- principles, types and applications. Viscometry-General features of fluid flow and nature of viscous drag for streamlined motion	1.0	20
4	**Optical & Diffraction Techniques**. Principle, Instrument Design, Methods & Applications of Polarimetry, Refractometry, Circular Dichroism and optical rotatory dispersion: Plain, circular and elliptical polarization of light, Relation between CD and ORD, application of ORD in conformation and interactions of biomolecules. Flow cytometry	0.5	10
5	**Microscopic techniques**: Principle and working of Compound microscope, Phase contrast microscope, Interference microscope , Fluorescence microscope , Polarizing microscope , Scanning and Transmission Electron Microscopy, CCD camera, Introduction to Atomic force microscopy, Confocal microscopy.	0.5	10
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1.Principles and techniques of practical biochemistry: Keith Wilson and John walker, Cambridge

2. Modern Experimental Biochemistry. Rodney F Boyer. Nenjamin/ Cummings publishing company Inc. Redwood city, California

Further Reading:

1. Practical Biochemistry- Principles and techniques. Keith Wilson and John walker (Eds), University press, Cambridge UK.

2. Principles and Techniques of electron microscopy- Biological applications. M.A Hayat., Mac Millan Press, London UK.

3. Biophysical Chemistry: UpadhyayUpadhyay and Nath, Himalaya Publishing House

4. Chromatographic methods. A Braithwate and F J Smith. Chapman and hall, NewYork.

5. Gel Electrophoresis of Nucleic acids- A Practical approach. Rickwood D and BD Hames. IRL Press, New York. 53

6. Spectrophotometry and Spectrofluorimetry: A Practical Approach. Harris DA and CL Bashford (Ed.) IRL Press, Oxford.

7. Introduction to Spectroscopy. Donald L. Pavia Gary M Lipman, George S Kriz. Harcourt brace College Publishers, Orlands, Florida

8. Gradwohls Clinical Laboratory Techniques. Stanley s. Raphael. W.E. Company, London, UK

9. Fundamentals of molecular Spectroscopy: C N Banwell, Tata Mc Graw hill publishing Company Ltd.

10. Spectroscopic methods and analyses: Christopher Jones, Barbara Mulloy Adrian H.Thomas.

11. Methods in Modern Biophysics: Bengt Nolting, Springer.

12. Bio separations Science and Engineering: Roger G Harrison, Paul Todd, Scott .R. Rudge, Oxford University Press.

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MAHATMA GANDHI UNIVERSITY

SBS M P C 12 LABORATORY COURSE-3

SchoolName	**School of Biosciences**
Programme	**MSc.Microbiology/Biochemistry/Biophysics/Biotechnology**
Course Name	**LABORATORY COURSE-3**
Type of Course	Core
Course Code	**SBS M P C 12**
Names of Academic Staff & Qualifications	**Dr. Jyothis Mathew** **MSc, PhD, Dr.Jisha M.S MSc, PhD and Dr.Radhakrishnan EK MSc, PhD**
Course Summary & Justification	e course includes training on sterilization and disinfection techniques, morphological, cultural and biochemical study of microbes and antibiotic sensitivity tests. The content of the course also include serological techniques. The technical knowhow of basic microbiological and serological methods is essential for post graduate programmes in all branches of Biosciences.
Semester	Second

Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	5	5	50		60
Pre-requisites	Theoretical knowledge in Microbiology and Immunology Basic laboratory skills					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	ıdents will acquire skills on practice of sterile and safety precautions in a Microbiology laboratory.	A	
2	ıdents will be able to prepare and sterilize media and to culture bacteria and fungi in laboratory	S	
3	Students will be able to examine morphological, physiological and biochemical properties of bacteria	S/E	
4	Students will be able to perform and interpret antibiotic sensitivity tests	S/E	
5	ıdents will be able to test and analyse the efficacy of disinfectants	S/An	
6	Students will be able to perform and interpret the various serological tests in a diagnostic laboratory	S/E	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1 1 2	Microscopic examination of bacteria in living conditions Testing of motility Staining procedures	0.5	30
2 2	Sterilisation methods Cultivation of bacteria and fungi Study of cultural characteristics and biochemical reactions of bacteria Testing of disinfectants Antibiotic sensitivity tests	0.75	45

3 3	Serological tests for the diagnosis of microbial infecdtions Agglutination and precipitation tests Immunodiffusion in gel ELISA	0.75	45
4 4 4	PAGE- Protein separation Native PAGE-Reagent preparation, Apparatus handling, gel casting, electrophoresis, and staining	0.5	30
5 5	DNA isolation Estimation of DNA RNA isolation Estimation of RNA Separation of DNA and RNA by Agarose gel electrophoresis Selective PCR amplification of a desired fragment	1.5	90
	Total Credits of the Course	4	240

Teachingand LearningApproach	**Laboratory Procedure (Mode of transaction)** Direct Instruction: lecture, Explicit Teaching, Demonstration, Hands on experimental sections, Skill acquisition by laboratory training
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Laboratory Skill Tests of maximum 20 marks 2. Seminar Presentation – Laboratory material and methods Maximum marks 10 3. Write a detailed report on instrumentation – 10 marks B. **Semester End Practical examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Medical Laboratory Manual for Tropical Countries Vol.2 Monica Cheesbrough ELBS, 2009
2.Mackie& McCartney Practical Medical Microbiology Churchil Livingstone, 1996
3. Molecular cloning by Sambrook, Fritsch and Maniatis, Cold Spring harbour laboratories
4. Biochemical Methods Sadasivam and Manickam
5. Gel electrophoresis of proteins: A practical approach (second edition) B D HAmes and Rickwood D(eds) Oxford University press

Further Reading:

1. Clinical Laboratory Methods Vol.2 Gradwohl The C.V.Mosby Company, 1981
2. London Practical Microbiology Dubey R.C.and Mahaswari D.K. S.Chand & Company Ltd. New Delhi, 2002
3. Experiments in Microbiology, Plant pathology and Biotechnology, K.R.Aneja, New Age International (P) Limited, New Delhi, 2003
4. Practical skills in Biomolecular Sciences, Weyers Jonathan, Reed Rob, Jones Allen, Holmes A D, Pearson publications

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MAHATMA GANDHI UNIVERSITY

SBS M P E 13: MICROBIAL TECHNOLOGY

School Name	**School of Biosciences**					
Programme	**MSc Biotechnology**					
Course Name	**Microbial Technology**					
Type of Course	**Elective**					
Course Code	**SBS M P E 13**					
Names of Academic Staff & Qualifications	**Dr. Keerthi TR**					
Course Summary & Justification	1. The course describe the application of microbes in various sectors 2. The course content explains the role of microbes and its utilization/application in various sectors especially in industrial & pharmaceutical area. 3. The course content also illustrates the various methods & process for production of bioactive compounds & products using microbes.					
Semester	Second					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisites	Basics of Microbiology					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**On completing this course, the student will be able to** Explain the methods for studying microbial genome and describe how metabolic & protein engineering help to enhance the production of microbial metabolites	**U/A**	

2.	Describe the methods , process & production of various microbial based food and dairy products also students have able to explain microbes are food for animal and human	**U/An**	
3.	Students should explain the role of microbes as biofertilizer, biopesticide, fungicide, and herbicide and also able to describe the various plant microbe interactions	**U/A**	
4.	Students have able to explain the methods and mechanism of microbes apply to protect various environmental sector.	**An/A**	
5.	Illustrate the utilization of microbes in the production of industrial and pharmaceutical products	**S/C**	
6.	Communicate effectively about a chosen topic in microbial technology both verbally and orally		

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Microbial Genomics:** Introduction to Microbial genomics, Structural Genomics, Functional genomics, Comparative Genomics, Meta Genomics - Genome analysis of extremophiles, Metabolic engineering and protein engineering for optimization of microbial products	**0.5**	**10**
2.	**Microbes in food & dairy industry:** Fermented foods-Introduction, Role & Advantages of fermented foods. Production of cheese, yoghurt, koji & Idli. Knowledge of other fermented dairy products. Single cell proteins-algae, bacteria, fungi, yeast & actinomycetes. Alcoholic beverages-Distilled and non distilled, Production of beer, wine & ethanol. Microbe as animal feed additives. Probiotics, Prebiotic & Synbiotics	**0.75**	**15**
3.	**Microbes in Agriculture**: Nitrogen fixation; Symbiotic & Non symbiotic Mechanism;Biofertilizers-Rhizobium, Azolla, Azospirillum, Algal Biofertilizers; Phosphate solubilizing microorganisms; Microbial biopesticide, biofungicide and herbicide; Micorrhiza; Plant –Microbe Interactions. Mushroom cultivation	**0.5**	**10**
4	**Microbes & Environment**: Biotechnology and	**0.5**	**10**

	pollution control; Use of immobilized microbial cell & enzyme in waste water treatment. Microbial biotransformation-Steroid, Microbial degradation of Herbicides, Insecticides & Pesticides; Bioremediation & Bioleaching		
5.	**Industrial &Pharmaceutical Applications**: Methanogens & Biogas Production; Microbial Hydrogen production; Microbes in plastic industry - Bioplastics; Microbial biosensors- Micro oxygen electrode. Biochips; Biofilm; Bioactive compounds from microbes. Bioethanol & biodieseal production. Microorganism for Bioassay & as Bio weapon	**0.75**	**15**
Total Credits of the Course		**3**	

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** K. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks L. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:
1. Biotechnology Fundamentals and Applications, S.S. Purohit and S.S. Mathur; Agro Botanical Publishers India.
2. Microbial Biotechnology, Alexander N Glazer & Hiroshi Nikaido Cambridge University Press.
3.Microbial Biotechnology, Farshad Darvishi harzevili Hongzhang Chen.CRC Press.
4. Microbial Biotechnology Principle & Applications Lee Yuan Kein.World Scientific Press.

Further Reading: 1. Microbial Technology-Fermentation Technology Vol 1 & 11 Peppler Perinas Elsiver. 2. Biofertilizers in Agriculture, N.S.Subha Rao;Oxford & IBH Publishing Co.Pvt.Ltd New Delhi. 3. Essentials of Biotechnology, R.C.Sobti & Suparna.S.Pachauri. Ane Books Pvt.Ltd. 4. Fermentation Technology Vol I&II. 5. Soil Microbiology – N.S. Subha Rao, 1999 6. Agriculture Microbiology – Rangaswamy 7. Microbial control and pest Management – S. Jayaraj. 8. Food Microbiology – Frazier W.C and Westhoff D.C., Tata Mc Graw-Hill 9. Food Microbiology – Rose A.H. in Economic Microbiology, Academic Pr

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	MAHATMA GANDHI UNIVERSITY
	SBS M PE 14 **ECOLOGY AND ENVIRONMENT**

School Name	**School of Biosciences**

Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**ECOLOGY AND ENVIRONMENT**					
Type of Course	Elective					
Course Code	**SBS M PE 14**					
Names of Academic Staff	**Dr J G RAY**					
Course Summary & Justification	The course is designed to equip students in perceiving, understanding and analyzing environmental problems from an ecological perspective, and a critical analysis of the existing control measures from a holistic perspective.					
Semester	second					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg: Authentic learning Collaborative learning Independent learning	54	18	0	28	100
Pre-requisite	**Knowledge in Biology at Graduate level**					

No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**
1	Students will be able to understand and communicate the sustenance of natural biological systems on the earth effectively	R/U/A	
2	They will acquire skills in explaining all kinds of interrelationships in natural biological systems	U/A	
3	Students will be able to explain environmental degradation and pollution as outcomes of ignorant and irresponsible human actions	U/An/Ap	
4	Students will be able to understand the significance of biodiversity and its conservation in the sustenance of natural ecosystems	An/Ap	

5	Overall, students will be skilful in analyzing as well as designing and maintaining of environmental sustainability of all kinds of developmental activities	R/U/A/An /Ap	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

Module No	Module Content	Credits	Hours
1	**Introduction to Ecology and different ecological objects**: Basic concept of the environment – components of the environment, the definition of ecology, ecological things. Autecological and Synecological concepts: **A.Population Ecology (Autecological concepts)**: **(a)** Characteristics of populations **(b)** Genecology - ecads, ecotypes, ecospecies, coenospecies; k-selection and r-selection populations **B. Synecological concepts(a)** Ecological processes of community formation, ecotone, edge effect. Classification of communities - criteria of classification, dynamic system of classification by Clement **(b)** Special plant communities - quantitative, qualitative and synthetic characteristics of plant communities**, (c)** Dynamic community characteristics - cyclic replacement changes and cyclic no-replacement changes	0.5	20 hrs
2	**Ecological succession -(a)** The concept – autogenic and allogenic succession, primary and secondary, autotrophic and heterotrophic **(b)** Retrogressive changes or the concept of degradation, concept of climax or stable communities, resilience of communities, ecological balance and survival thresholds **Biosphere and Ecosystem - (a)** Significance of habitat, biodiversity, ecological niche, trophic level, primary and secondary productivity, food chains, food webs, ecological pyramids, energy flow and nutrient cycles **(b**) Comparative study of the significant world ecosystems: Different aquatic and terrestrial ecosystems concerning their productivity, 0.5 57 biodiversity, energy flow, food chains and trophic levels	0.5	20 hrs
3.	**Natural Resources:** Soil, water and air Resources – soils and parent materials – ecology of soil fertility; Fresh water and marine resources – global distribution of water resources – surface and groundwater resources – water conservation – prevention of marine pollution – conservation of marine	0.5	10 hrs

	resources; Atmospheric resources – the structure of atmosphere – climate and weather – climatic factors – precipitation, wind temperature, aerosols		
4	**Environmental pollution**: **(a)** Definition and classification **(b)** Water pollution: Water quality parameters and standards, different types of pollutants and their consequences. Types of water pollution, prevention and control - watershed management, different kinds of wastewater treatments; Phyto and bioremediation **(c)** Air pollution: Air quality standards and index, ambient air monitoring using high volume air sampler, types and sources of air pollutants, air pollution and human health hazards, control of air pollution **(d)** Noise pollution **(e)** Radioactive and thermal pollution: Causes and hazardous effects, effective management **(f)** Concept of solid wastes **(g)** Pollution Control - Bioremediation, Phytoremediation, bioaugmentation, biofilms, biofilters, bioscrubbers and trickling filters. Use of bioreactors in waste management	1.0	40 hrs
5	**Climate Change and other Global Environmental issues** - Factors responsible for climate change, Climate change mitigation – global conventions and protocols on climate change - El-Nino and La Nina phenomenon and its consequences; Environmental laws, environmental monitoring and bioindicators, environmental safety provisions in the Indian constitution, major ecological laws in free India; UNEP and its role in climate change control– IPCC, UNFCC, annual environment summits – 1973 Stockholm conference to 2015 Paris Conference – new developments of annual UNFCC meetings in the coming years - Future Earth Programme	0.5	10 hrs
	Total Credits of the Course	**3**	**100 hrs**
Books for Reference			
Compulsory Reading: 1. MC Dash (1993) Fundamentals of Ecology, Tata McGraw Hills 2. Odum EP 3rd Edition (1991) Fundamentals of ecology, Saunders and Com			
Optional Further Reading 1. Barbour MD et al. (1980) Terrestrial plant ecology. The Benjamin-Cummings Pub. Com 2. 2. Benton AH and Werner WE (1976) Field biology and Ecology, Tata McGraw Hill 3. Blanco-Canqui and Humberto LR (2008) Principles of Soil Conservation and Management, Springer			

4. Molles MC (2012) Ecology – Concepts and applications, 6th Edition, Mc Graw Hill	
Course evaluation: **Assignments & Seminar (10 marks each); Two internal test papers (20 effects) end semester examination (60 marks)**	

MAHATMA GANDHI UNIVERSITY

SBS M PE 15: NEUROBIOLOGY

SchoolName	**School of Biosciences**					
Programme	**MSc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**NEUROBIOLOGY**					
Type of Course	Elective					
Course Code	**SBS M PE** 15					
Course Offered By	**Dr Harikumaran Nair R**					
Course Summary & Justification	This course is designed to provide an overview of Neurobiology. Stress will be placed on methods and concepts rather than facts alone. The course will proceed from the basic biophysical properties of neurons and glia to the physiological basis of learning, memory, and sensory processing					
Semester	Second					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics Knowledge in Physiology**					

COURSE OUTCOMES (CO)

CO	Expected Course Outcome	Learning	PSO No.

No.		Domains	
1	Students should be capable of effectively communicating how neural system works	U	
2	Students should be able to explain electricity and the biophysics of cell	E	
3	Students should describe how do neurons talk to one-another	A	
4	Students should be able to explain how neural circuits organize information	A	
5	Students should be able to narrate how is information stored	E	
6	Lastly, students should gain a general understanding how is information collected and processed.	I	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Introduction to neurobiology, the structure and distinguishing features of neurons, how is a neuron recognized? The architecture of nervous systems. Neuronal model systems. Chemical/electrical synapses. Recording/monitoring techniques.	0.5	10
2	Ionic basis of the resting potential. Maintenance of resting membrane potential, passive and active mechanisms, channels and pumps, ionic permeability	0.5	10
3.	Action potentials and ion channels, Mechanism of nerve action potential: Characteristics of action potential, initiation and propagation of action potential, voltage dependent sodium channels, mechanism of action potential propagation, factors affecting the speed of action potential propagation,	1	20

	molecular properties of voltage sensitive sodium channels, molecular properties of voltage dependent potassium channels, calcium dependent action potentials, voltage- clamp analysis of action potentials		
4	Synaptic transmission: Chemical and electrical synapse, neurotransmitter release, synaptic potential, excitatory synaptic transmission between neurons, excitatory neurotransmitters, inhibitory synaptic transmission, inhibitory neurotransmitters, neurotransmitter gated ion channels, presynaptic inhibition and facilitation, neuronal integration, synaptic transmission at neuromuscular junction	0.5	10
5	Synaptic plasticity, language and cognition: Short term changes in synaptic strength, long term changes in synaptic strength, modification of synaptic strength in reflex circuits, learning, language function and cortical areas involved in language, cognition, dementia and loss of cognitive abilities	0.5	10
	Total Credits of the Course	3	60

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading

1. Basic Neurochemistry- Molecular, cellular and medical aspects. George J Siegel, Bernard W Agra noff R, Wayne Albers, Stephen K Fisher & Michael D Uhler
2. Neurobiology: Molecules, cells and systems. Gary G Mattews
3. From Neuron to Brain- John G Nicholls, A Robert Martin, Bruce G Wallace & Paul A Fuchs

Further Reading:

1. Neuroscience, edited by Purves, Augustine, Fitzpatrick, Hall, LaMantia, Mooney, Platt and White. Sinauer (2018) Sixth Edition.
2. Foundations of Neurobiology, Delcomyn, F. 1st edition W. H. Freeman and Company (1998)
3. Behavioral Neurobiology: An Integrative Approach, Zupanc, G. K. H. Oxford University Press. 2nd edition (2010)
4. Neurobiology: molecules cells and systems Gary G. Mathews 2nd edition. Blackwell Science Inc. (2001).
5. Neuroscience: exploring the brain. Bear, M., Connors, B.W. and Paradiso, M.A. 2nd edition Lippincott, Williams and Wilkins (2001)

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MAHATMA GANDHI UNIVERSITY

SBS M PE 16 ENVIRONMENT SCIENCE

SchoolName	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**ENVIRONMENT SCIENCE**					
Type of Course	Elective					
Course Code	**SBS M PE** 16					
Course Offered By	**Dr M S Jisha**					
Course Summary & Justification	This course on environmental Science deals with principles and scope of environment science. The objective of the course content is to create a sound awareness about the environment impact and its monitoring and Predict the consequences of human actions on the web of life, global economy and quality of human life The course content is designed with a view to augment CSIR/UGC syllabus					
Semester	second					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of Biological sciences**					

COURSE OUTCOMES (CO)

CO	Expected Course Outcome	Learning	PSO No.

No.		Domains	
1	Gain in-depth knowledge on natural processes that sustain life and govern economy.	U/A	
2	Able to describe the principles of ecology	U/ C	
3	Develop critical thinking for shaping strategies (scientific, social, economic and legal) for environmental protection and conservation of biodiversity, social equity and sustainable development.	R/An	
4	Acquire values and attitudes towards understanding complex environmental-economic social challenges	U/R	
5	Understand the current environmental problems and preventing the future ones.	U/R	
6	Create an insight to the strategies and methodologies of environmental impact assessment	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Definition, principles and scope of environmental science, Earth, Man and environment, ecosystem, pathways in ecosystem. Physic-Chemical and Biological factors in the environment Geographical classification and Zones. Structure and functions of ecosystem, Abiotic and biotic components, energy flows, food chains, Food, web, Ecological pyramids, types and diversity Terrestrial (Forest, grass land) and Aquatic (Fresh water, marine, eustarine) ecosystems. mineral cycling. Habitat and niche. Major terrestrial biomes. Impact of microorganisms on global ecology, microorganisms in extreme environment	0.5	10
2	Definition, Principles and scope of ecology, Human ecology and Human settlement, evolution, origin of	0.5	10

	life and speciation Population ecology characteristics and regulation. Community ecology structure and attributes. Levels of species diversity and its management, Edges and ecotones. Ecological succession. Concept of climax. Common Flora and fauna in India. Endangered and Threatened Species		
3.	Biodiversity status, monitoring and documentation Biodiversity management approaches. Conservation of biological diversity, methods and strategies for conservation. Natural resources, conservation and sustainable development. Hotspots of biodiversity, National parks and Sanctuaries	0.5	20
4	Environmental pollution- Air: Natural and anthropogenic source of pollution, Primary and Secondary pollutants , Methods of monitoring and control of air pollution, effects of pollutant on human beings, plants animals, material and on climate, Acid rain, Air Quality standards Water: types, Sources and consequences of water pollution, Physio-chemical and Bacteriological sampling and analysis of water quality, Soil: Physio-chemical and Bacteriological sampling as analysis of soil quality, Soil pollution- control, Industrial waste effluents, and heavy metals Their interaction with soil components, Noise: Sources of noise pollution, Noise control and battement measures. Impact of noise on human health, Radioactive and thermal Pollution. Bioremediation-	1.0	20

	Strategies for bioremediation, Biosensors, biological indicators of pollution and monitoring. Detoxification of hazardous chemicals, mycotoxins. Biological weapons		
5	Introduction to environmental impact analysis, Impact Assessment Methodologies Generalized approach to impact analysis, Guidelines for Environmental Audit Introduction to environmental Planning, Environmental priorities in India and Sustainable development, Environment protection-issues and problems, International and national efforts for environment Protection. Global environmental problems-Ozone depletion, global warming, climatic change, desertification, green movement, ecofeminism. Current environmental issues in India	0.5	10
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** M. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic

	based on research findings and literature search – 10 marks N. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Jonathan B (2016) Principles of Evolution, Garland Science, Taylor and Francis.
2. Odum E. P and Barret G W. Fundamentals of ecology. W. B Saunders company, Philadelphia
2. Chapman and Reiss, Ecology principles and applications. Cambridge University

Further Reading:

1. Jobes A. M., Environmental biology, Routledge, London.
2. Odum E. P. Basic ecology. Saunders College.
3. A textbook of environmental sciences, Arvind kumar.
4. Alleby M.Basics of environmental science. Routledge, Newyork
5. Cunningham, W. P and Siago, B. W, Environmental science.
6. Kewin T. P and Owen C. A., Introduction to global environmental issues. Routledge, London.
7. Chiras,D.D, Environmental science Cell and Molecular Biology by De Robertis E.D.P, 8th Edition

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MAHATMA GANDHI UNIVERSITY

SBS M PE 17 MOLECULAR MICROBIOLOGY

SchoolName	School of Biosciences					
Programme	M.Sc. Microbiology/Biotechnology/Biochemistry/Biophysics					
Course Name	MOLECULAR MICROBIOLOGY					
Type of Course	Elective					
Course Code	**SBS M PE** 17					
Course Offered by	**Dr Radhakrishnan E K**					
Course Summary & Justification	This course on Molecular Microbiology deals with the applications of various molecular biological techniques in Microbiology. This course is an important branch of Microbiology. Rapid identification of microorganisms is very important for the clinical, diagnostic and research purposes and the methods used for the same have developed significantly with the advances in Molecular biology. The content in this course has been designed with an objective to provide detailed understanding on the techniques, principle and applications of molecular biology for the microbial identification, production of recombinant proteins and also for studying the unculturable microorganisms through metagenomics. This will enable the students to identify the research, learning and job opportunities based on the latest developments in this subject.					
Semester	Second					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding on microorganisms and molecular biology Knowledge in any branch of Life science					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1.	Students will able to understand and explain molecular biological applications in microbiology	R/U	
2.	Students will able to learn rapid methods used for the microbial identification	R/U	
3.	Students will able to understand the functioning of human microbiome and its beneficial role	U/ An/E	
4.	Students will become able to understand molecular basis of microbial virulence	U/An/A	
5.	Students will able to apply the knowledge for advanced microbiological applications	C/S	
6.	Students will able to identify the research and technical opportunities in molecular microbiology	A/S	
**Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Molecular biology of Microbial evolution, rRNA sequence and cellular evolution, Signature sequence and phylogenetic probe. Identification and characterization of microorganisms, Molecular methods for microbial identification, Molecular typing methods: Bacterial strain typing, Pulsed Field Gel Electrophoresis, PCR-based microbial typing, Genotyping by Variable Number Tandem Repeats, Multilocus Sequence Typing, Automated Ribotyping	1	10
2	Unculturable bacteria and Metagenomics, Methods used in metagenomics, New generation sequencing technologies for metagenome study, Human microbiome, Importance of human microbiome in relation to human health and disease.	1.0	20
3.	Molecular basis of microbial virulence. Bacterial adherence: basic principles, effects of adhesion on bacteria and host cells. Bacterial invasion of host cells; mechanism. Bacterial toxins: classification based on molecular features, Molecular detection and characterisation of bacterial pathogens, detection of bioterrorism.	0..5	10

4	Microbial production of recombinant proteins: expression, purification and applications, Microbes in plant transformation, Agrobacterium tumefaciens T-DNA transfer process, Application of microorganisms for combinatorial and engineered biosynthesis, Engineering *E.coli* for the production of curcumin	0.5	10
	Total Credits of the Course	3	

Teaching and Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction, Active co-operative learning, Seminar, Group Assignments Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar - Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. Semester End examination – 60 marks

REFERENCES

Compulsory Reading:

1. Molecular Microbiology – Diagnostic Principles and Practice, David H. Persing, Fred C. Tenover, James Versalovic, Yi-Wei Tang, Elizabeth R. Unger, David A. Relman, Thomas J. , ASM Press., 2016
2. Brock Biology of Microorganisms- Michael T. Madigan and John M.Martinko,

Prentice Hall, 2015
Further Reading: 3. Microbial Physiology – Albert G. Moat, John W. Foster and Michael P. Spector , 2002 4. Metagenomics for Microbiology, Jacques Izard Maria Rivera , 1st edition, Academic Press Published Date: 12th November 2014 5. Production of Recombinant Proteins: Novel Microbial and Eukaryotic Expression Systems, Gerd Gellissen, May 2005Longman,2013

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MAHATMA GANDHI UNIVERSITY

SBS M PE 18
DEVELOPMENTAL BIOLOGY

School Name	**School of Biosciences**
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**
Course Name	**DEVELOPMENTAL BIOLOGY**
Type of Course	Elective
Course Code	**SBS M PE** 18
Names of Academic Staff & Qualifications	**Dr J G RAY**

Course Summary & Justification	The course is designed to equip students in perceiving, understanding, and analyzing reproductive and embryological developmental processes in plants to apply the principles towards increasing plant productivity through breeding.					
Semester	second					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	E.g., Authentic learning Collaborative learning Independent learning	54	18	0	28	100
Pre-requisite	**Knowledge in Botany at the Graduate level**					

No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**
1	Students will be able to understand and communicate the reproductive and developmental events in plants effectively	R/U/A	
2	They will acquire the skills to explain all kinds of reproductive parts and seed developmental processes, including seed storage in plants	U/A	
3	They will be able to explain how developmental processes initiates and proceeds in plants	U/An/Ap	
4	Students will be able to explain the specific developmental process and its ultimate impact on the productivity or successful completion of lifecycle in plants	An/Ap	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

Module No	**Module Content**	**Credits**	**Hours**
1	**Introduction:** Basic concepts of developmental Biology; An overview of plant and animal development, Potency, Commitment, Specification, Induction, Competence, Determination and Differentiation morphogenetic	1.0	40 hrs

	gradients; cell fate and cell lineages; stem cells; genomic equivalence and the cytoplasmic determinants; imprinting; mutants and transgenics in the analysis of development		
2	**Development in flowering plants**: **(a)** Angiosperm life cycle **(b)** Anther: Structure and development, microsporogenesis, male gametophyte development. Palynology: Pollen morphology, exine sculpturing, pollen kit, NPC formula. Applications of palynology- palynology concerning taxonomy. Viability of pollen grains Pollination, pollen germination, growth and nutrition of pollen tube**. (c)** Ovule: Structure, ontogeny and types. Megasporogenesis. Embryosac – development, classes, ultrastructure, and nutrition of embryosac. Female gametophyte development.	0.5	15 hrs
3.	**Fertilization in Plants:** Double fertilization; embryo development - different types. Endosperm development, types of endosperm, haustorial behaviour of endosperm. Xenia and metaxenia. Polyembryony – types and causes. Seed formation, dormancy and germination. Apomixis, Parthenogenesis.	0.5	15 hrs
4	**Morphogenesis and organogenesis in plants:** Shoot and root development; Leaf development and Phyllotaxy. Transition to flowering, floral meristems and floral development; Homeotic genes in plants; Senescence, programmed cell death and hypersensitive response in plants	1.0	30 hrs
	Total Credits of the Course	**3**	**100 hrs**
Books for Reference			
Compulsory Reading: 1. Maheswari P. 1950. An introduction to the embryology of Angiosperms. McGraw Hill 2. Wolpert L, C Tickle and AM Arias (2015) Principles of development			
Optional Further Reading 1. Krishnamurthy KV (2015) Growth and Development in Plants 2. Raghavan V (2000) Developmental Biology of Flowering Plants			

3. Gilbert SF (2000) Developmental Biology 4. Developmental Biology, 8th Ed, Gilbert 5. Developmental Biology Paperback – 2008 by Werner A. Muller	
Course evaluation: **Assignments, 1 Seminar, and one assignment (10 marks each) Two internal test papers (20 marks) end semester examination (60 marks)**	

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THIRD SEMESTER

	MAHATMA GANDHI UNIVERSITY
	SBS M P C 19 ENZYMOLOGY

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**BIOCHEMISTRY**					
Type of Course	Core					
Course Code	**SBS M P C 19**					
Names of Academic Staff & Qualifications	**Prof. M S Latha – M. Sc (Biochemistry), PhD**					
Course Summary & Justification	The course is designed to get a deep knowledge of the mechanisms by which cellular reactions are accelerated. The course builds a base for the students to understand and predict the metabolism of all living things and provide basics of drug development process related to enzyme targets and enzyme therapy					
Semester	Third					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHou

						rs
	Eg. Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic idea about protein structure and function					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To give details of different characteristics of enzymes, enzyme classification and nomenclature, different factors that affect their activity and their mechanisms of action.	U	
2	To contrast different modes of enzyme inhibition and regulation.	An/E	
3	To describe the structure and functions of vitamins and hormones	U	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hours
1	**Introduction to enzymes:** Holoenzyme, apoenzyme, and prosthetic group; Interaction between enzyme and substrate- lock and key model, induced fit model, Features of active site, activation energy. Enzyme specificity and types; Enzyme Commission system of classification and nomenclature of enzymes (Class and subclass with one example) **Mechanisms of enzyme action-** Rate Enhancement Through Transition State Stabilization, Acid-base catalysis, covalent catalysis, metal ion catalysis (eg: Serine Proteases: cystenyl protease, aspartyl protease, metalloprotease, Lysozyme). **Coenzymes and their functions** - NAD, $NADP_+$, FAD, FMN, lipoic acid, TPP, pyridoxal phosphate, biotin and cyanocobalamin **Isolation and characterization of enzymes:** Isolation of enzymes and the criteria of purity; Characterization of enzymes- active site mapping, Measurement and expression of enzyme activity, enzyme assays. Definition of IU, katals, enzyme turnover number and specific activity.	1.0	15

2	**Enzyme kinetics:** Factors affecting the velocity of enzyme catalyzed reaction- enzyme concentration, temperature, pH, substrate concentration, inhibitors and activators (explanation with graphical representation). Derivation of Michaelis -Menten equation and Km value determination and its significance, Definition of V_{max} value of enzyme and its significance, Lineweaver- Burk plot, Eadie-Hofstee and Hanes plots.Bi-substrate reactions: Classification, Reaction mechanisms.	1.0	10
3.	**Enzyme inhibition:** Reversible and irreversible – examples. Reversible-competitive, noncompetitive, uncompetitive inhibition and mixed inhibition, Irreversible inhibition- mechanism based inactivators, affinity labels, group specific inhibitors; Graphic Determination of Inhibitor Type; Dose—Response Curves of Enzyme Inhibition; Mutually Exclusive Binding of Two Inhibitors; Structure—Activity Relationships and Inhibitor Design; Tight Binding Inhibitors: Identifying Tight Binding Inhibition, examples; Time-Dependent Inhibition: examples; Distinguishing between modes of inhibitor interaction with enzyme	1.0	15
4	**Regulation of Enzyme activity: Different c**ovalent modifications,; Zymogen form of enzyme and zymogen activation; Multienzyme complexes and their role in regulation of metabolic pathways; Allosteric regulation: example Aspartate trascarbamoylase, Sigmoidal kinetics of allosteric enzymes, Models of Allosteric Behavior, Effects of Cooperativity on Velocity Curves. Isoenzymes- Lactate dehydrogenase and creatine phosphokinase.	0.5	10
5	**Application of enzymes:** Applications of enzymes in industry (eg: in food industry, paper and leather industry, detergent industry and waste management). Diagnostic and therapeutic enzymes; Applications of enzymes in life science research, Ribozymes, Abzymes, Immobilised enzymes, Biosensors, synthetic enzymes, Enzyme engineering	0.5	10
	Total Credits of the Course	4	
Books for Reference			

Compulsory Reading:

1. Fundamentals of Enzymology: The Cell and Molecular Biology of Catalytic Proteins by Nicholas C. Price, Lewis Stevens, and Lewis Stevens (2000) Publisher: Oxford University Press, USA ISBN: 019850229X ISBN-13: 9780198502296, 978-0198502296
2. Enzyme Kinetics: A Modern Approach Book: Enzyme Kinetics: A Modern Approach by Alejandro G. Marangoni (2003) Publisher: Wiley-Interscience ISBN: 0471159859 ISBN-13: 9780471159858, 978-0471159858
3. Principles Of Biochemistry, 4/e (2006) by Robert Horton H , Laurence A Moran, Gray Scrimgeour K **Publisher:** Pearsarson **ISBN:** 0131977369, **ISBN-13:**9780131977365, 978-0131977365
4. Biochemistry 6th Edition (2007) by Jeremy M.berg John L.tymoczko Lubert Stryer **Publisher:** B.i.publicationsPvt.Ltd **ISBN:**071676766X **ISBN-13:** 9780716767664, 978-716767664
5. Lehninger Principles of Biochemistry, Fourth Edition by David L. Nelson David L. Nelson (Author)

Further Reading:

- Enzyme Kinetics and Mechanisms by Taylor Publisher: Spring ISBN: 8184890478 ISBN-13: 9788184890471, 978-8184890471
- Enzyme Mechanism by P.K. Shivraj Kumar (2007) Publisher: RBSA Publishers ISBN: 8176114235 ISBN-13: 9788176114233, 978-8176114233
- Enzymes and Enzyme Technology by Kumar (2009) Anshan Pub ISBN: 1905740875, ISBN-13: 9781905740871, 978-1905740871
- Enzymes in Industry: Production And Applications by Aehle W (2007) Publisher: John Wiley & Sons Inc ISBN: 3527316892 ISBN-13: 9783527316892, 978-3527316892
- Enzymes: Biochemistry, Biotechnology, Clinical Chemistry (second Edition) by Trevor Palmer, Philip Bonner (2007) Publisher: Horwood Publishing Limited ISBN: 1904275273 ISBN-13: 9781904275275, 978-1904275275

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** **O. Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks **P. Semester End examination – 60 marks**

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	MAHATMA GANDHI UNIVERSITY
	SBS M P C 31: MOLECULAR BIOPHYSICS

SchoolName	**School of Biosciences**
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**

Course Name	**MOLECULAR BIOPHYSICS**					
Type of Course	Core					
Course Code	**SBS M P C 31**					
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S**					
Course Summary & Justification	This course is designed to provide an overview of chemical structure of various macromolecules involved in propagation of life. This course comprehends the influence of macromolecular three dimensional structure on their function Appreciate the relevance of bioinformatics to the function of biological macromolecules					
Semester	Third					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of** Molecular biophysical techniques and bioinformatics					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To describe and discuss the relationship between the structure and function of biological macromolecules	E	
2	To determine the role of biological receptors	U/ An	
3	To narrate the cell membrane structure and function	R	
4	To explain how to use bioinformatics tools and software's	S	

5	To know different molecular biology databases and formats in which data is stored	E	
6	To describe the concepts of computer aided drug discovery	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Structure and conformations of proteins**: Structure stabilizing forces, Super secondary structural domains, Domain and motifs, Classification and role of reverse turn, Hydrogen bonding in globular proteins. Main chain and side chain conformation of globular proteins. Globular and fibrous proteins. Conformation, organization and interactions of structural proteins such as collagen, alpha-keratin, silk fibroin, actin and myosin. Examination of 3D structure of chymotrypsin and Rubisco. Folded conformation of globular proteins- lysozyme and cytochromes. **Protein evolution**: Concept of protein evolution, Protein speciation, phylogeny, phylogenetic tree- cladogram, phylogenetic study of cytochrome c. Homologies and analogies- convergent, divergent and parallel evolution **Structure and conformation of polysaccharide** - Amylase, Cellulose, Chitin.	1.0	10
2	**Biological receptors**: Drug-receptor interaction. Protein ligand interaction: Ligand- binding sites of Immunoglobulins, substrate- binding sites of Seriene proteases, Haem-binding sites, Nucleotide- binding sites, Binding sites for phosphoryl groups, Interaction of proteins with other macromolecules- lipoprotein, Glycoprotein and Nucleoprotein.	0.5	10

3.	**Membrane biophysics**: Various membrane models, Carbohydrate, Lipids & Proteins components of cell membrane, Electrical properties of lipids and proteins, Principles of membrane organization & stability, Molecular motion in membrane & membrane fluidity, Structure of membrane proteins- Integral membrane proteins, Peripheral membrane proteins, Polypeptide toxins, Membrane protein complex, Transport across membrane, Selective permeability of biomembrane, Selectivity & ion specificity of biomembrane, Ion channel structure and gating function, Ion channel types and characterization, Role of carriers in ion transport.	1.0	20
4	**Bioinformatics:** Nature and scope of bioinformatics, Biological databases and various file formats, Sequence retrieval and submission, Overview of available Bioinformatics resource on the web- NCBI, EBI, EXPASY etc, Sequence alignment – BLAST and FASTA, Pairwise sequence alignment:- global and local alignment, multiple sequence alignment, Clustal W, Clustal X, Pattern and profile databases – PROSITE and BLOCKS, Structure databases – understanding structures from Protein Data Bank, Metabolic databases – post translational modification databases, Tools for viewing and interpreting macromolecular structures- Swiss pdb viewer, Webmol, Rasmol, PyMol, Chimera.	1.0	10
5	**Application of Bioinformatics in Drug discovery**: Role of bioinformatics in drug discovery, Different methods of target identification and validation, Ways of lead identification and optimization, *In-silico* prediction of ADMET properties of drug molecules, Molecular docking and prediction of drug quality. Bioinformatics companies.	0.5	10
	Total Credits of the Course	4	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Introduction to Proteomics Tools for the New Biology: Daniel C. Liebler, Humana press

2. Proteins, Structure and molecular properties, Thomas E Creighton

Further Reading:

1. Molecular Biophysics- Volkenstain M.V

2. Biopolymers, AP- Watson,A.G & Blackwell,J

3. Principles of protein structure Schulz, G.E. & Schimmer, R.H..

4. Structure and Molecular Principles, W.H. Freeman & Co.Creighton, T.E.

5. Saenger,W., Principles of Nucleic Acid Structure, Springer.

6. Protein-Protein Interactions: Erica Golemis, CSHL Press.

7. Protein Architecture A practical Approach: A.M.Lesk,

8. Molecular and Cellular biophysics, Jack A. Tuszynski, Chapman & Hall/ CRC

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	MAHATMA GANDHI UNIVERSITY
	SBS M P C 32: ELECTROPHYSIOLOGY

SchoolName	**School of Biosciences**
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**
Course Name	**ELECTROPHYSIOLOGY**
Type of Course	Core
Course Code	**SBS M P C 32**

Names of Academic Staff & Qualifications	**Dr. R. Harikumaran Nair**					
Course Summary & Justification	The course will deal with the methodological aspects of electrophysiology The course will cover the main methods used for human electrophysiology The course will empower the students to carry out and interpret the various neurophysiological techniques					
Semester	Third					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of** Biophysics and Biostatistics					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Estimate the feasibility of recording and stimulating any electrophysiological signal from first principles of biophysics	E	
2	Describe the working principles of all currently available medical devices for therapeutic modulation of neural signals	U/ An	
3	Identify technological and biological limitations in the treatment of clinical disorders of the heart, motor control and special senses	R	
4	Record and analyze common electrophysiological signals, including ECG, EMG and EEG	An/ C	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Basic principles of electricity, overview of electrophysiological instrumentation, types of electrophysiological recordings, current voltage relationship and the membrane potential	0.5	10
2	Electrophysiology of heart, electrocardiogram (ECG), source of ECG voltage-dipole theory, vector analysis of ECG, Brain Potentials, electroencephalogram (EEG), source and mechanism of formation of rhythmic pattern of EEG, characteristics of EEG waves	0.5	10
3.	Event related potential (evoked potential)-types, characteristics, and significance electromyogram (EMG) – motor unit potential, physiological significance and analysis of EMG	1.0	20
4	Structure of retina, electrical response of the photoreceptors to light, molecular mechanism of phototransduction process, auditory system in brain, mechanoreceptors of vibration sense, cochlear mechanics	0.5	10
5	Chemical sense, structure of olfactory epithelium, chemo transduction in olfactory receptor cells, processing of olfactory information in brain, structure of the taste buds, chemo transduction in taste receptor cells, processing of taste information in brain	0.5	10
	Total Credits of the Course	3	

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** Q. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified

	to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks R. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. From Neuron to Brain. -John G. Nicholls, A. Robert Martin, Bruce G. Wallace & Paul A. Fuchs.

2. Ion channels. Molecules in Action-David J. Aidley & Peter R. Stanfield

3. The Physiology of Excitable Cells- David J. Aidley

Neurobiology: Molecules, cells and systems. Gary G Mattews

Further Reading:

1. Principles of Neuroscience.Kandel, Schwartz and Jessell, ed., McGraw-Hill, 4th ed., 2000.

2. Glover, BM, Brugada, P. eds. Clinical Handbook of Cardiac Electrophysiology. New York, NY: Springer; 2016.

Sra, JS, Akhtar, M. eds. Practical Electrophysiology. Minneapolis, MN: Cardiotext Publishing; 2014.

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MAHATMA GANDHI UNIVERSITY

	SBS M P C 33 LABCOURSE 4-Biophysics

SchoolName	**School of Biosciences**					
Programme	**MSc Biophysics**					
Course Name	**LABCOURSE-4**					
Type of Course	Core					
Course Code	**SBS M P C 33**					
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S** **Dr.Anie Y – M. Sc (Biochemistry), PhD** **Guest faculty– M. Sc (Biochemistry), PhD**					
Course Summary & Justification	To familiarize the student to various Biophysical and Molecular techniques. To develop laboratory experience on spectrometry, chromatography, electrophoresis, To develops practical skills about Isolation of DNA and PCR amplification					
Semester	Third					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of** molecular biophysical methods					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.

1	To explain and characterize UV absorption spectrum	E	
2	To extract and purify enzymes from different sources and to examine their kinetic behavior	U/ An	
3	To perform purification of compounds by column chromatography and HPLC	R	
4	To perform the structural elucidation of purified compounds	S	
5	To perform the DNA isolation and PCR amplification	E	
6	To perform various electrophoretic techniques	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Absorption spectra analysis** • To study the characteristics of UV absorption spectra of Aromatic Amino Acids • To study the characteristics of UV absorption spectra of Proteins • To study the characteristics of absorption spectra of Nucleic Acids and Nucleotides	0.5	10
2	**Enzyme Kinetics** • Effect of Substrate Concentration on velocity of Enzyme catalyzed reaction: Determination of K_M and V_{max} using Line weaver- Burk plot • Effect of Temperature on velocity of Enzyme catalyzed reaction:	1.0	10

	Determination of Q_{10} • Effect of pH on velocity of Enzyme catalyzed reaction: • Effect of activators on velocity of Enzyme catalyzed reaction: • Determination of type of inhibition using Line-weaver Burk plot • Docking of Enzymes with ligand molecules using docking softwares • Determine the drug likeliness of ligand molecules • Determining Binding energies of ligand with receptors • Determining Ki values		
3.	**Chromatography** • Paper chromatography of Plant Pigments • Paper chromatography of aminoacids • Paper chromatography of sugars. • Thin layer chromatography of aminoacids. • HPTLC separation of plant pigments • HPTLC of Amino acids & sugars • To isolate the proteins- Casein from milk and Hb from RBC • To analyse the Erythrocytes membrane lipids by TLC • Fractionation of Sugars from fruit juice using TLC/HPTLC • Column Chromatography separation of plant derived compounds. • UV Visible spectrum of the separated compounds	1.0	20

	• To perform the separation of column fractions using HPLC • GCMS analysis of purified compounds • LCMS analysis of purified compounds • FTIR analysis of purified compounds		
4	**Electrophoresis** • Paper Electrophoresis of Amino acids • Cellulose acetate strip Electrophoresis of Amino acids • Polyacrylamide Gel Electrophoresis (PAGE). • SDS- Polyacrylamide Gel Electrophoresis (PAGE) • DNA isolation from bacteria and plants. • Agarose gel electrophoresis of DNA • PCR amplication of 16SrDNA	0.75	10
5	**Bioinformatics** • Literature databases: PubMed, PMC and PLOS. • Nucleic acid sequence databases: NCBI, EMBL and DDBJ. • Protein sequence databases: Uniprot and TrEMBL. • Protein structure databases: PDB and SCOP. • Metabolic pathway databases: KEGG and Reactome. • Protein interaction databases: STRING and BioGRID. • Homologous sequence search by BLAST and FASTA.	0.75	10

	• Multiple sequence alignment and tree construction		
	Total Credits of the Course	4	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** S. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks T. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Introductory Practical biochemistry, S. K. Sawhney &Randhir Singh (eds) Narosa Publishing House, New Delhi, ISBN 81-7319-302-9, p 195 – 303
2. Standard Methods of Biochemical Analysis, S. K. Thimmaiah (ed), Kalyani Publishers, Ludhiana ISBN 81-7663-067-5, p 12 - 182.
3. Biochemical methods by S Sadasivan and A Manickam. New Age international publishers
4. Biotechnology: A laboratory course by Becker J.M.
5. Bioinformatics Practical Manual by Mohammed Iftekhar and Mohammed Rukunuddin Ghalib

Further Reading:

1. An introduction to practical Biochemistry by D. T. Plummer, Mc Graw Hill.

2. Laboratory manual in Biochemistry by J. Jayaraman, Wiley Eastern Limited.
3. Introductory Practical Biochemistry by S. K. Sawhney and Randhir Singh, Narosa
4. Experimental Biochemistry: A student companion by Beedu Sashidhar Rao and Vijay Deshpande, I.K. International Pvt. Ltd., New Delhi.
5. Lab manual in Biochemistry by J. Jayaraman, Wiley Eastern Limited
6. An introduction to practical Biochemistry by D. T. Plummer, Mc Graw Hill.
7. Introductory Practical Biochemistry by S. K. Sawhney and Randhir Singh, Narosa
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MAHATMA GANDHI UNIVERSITY

SBS M P E 42: QUALITY CONTROL IN HERBAL DRUGS

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**QUALITY CONTROL IN HERBAL DRUGS**					
Type of Course	Elective`					
Course Code	**SBS M P E 42**					
Course Summary & Justification	The course is designed to get a clear idea on quality control approaches in natural herbs and products and modern analytical techniques for the analysis of the herbal drugs.					
Semester	Third					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Eg. Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding of plant-based drugs					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To estimate the quality assurance of herbal materials.	C	
2	To isolate, purify and characterize the photochemical from medicinal plants.	A	
3	To interpret the structure of natural products	U/E	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hours
1	WHO Guidelines for Quality Control of herbal raw materials. Determination of pesticide residue, arsenic and heavy metals, afflatoxins and microbial contaminants	0.5	10
2	Definition, principle of the various extraction techniques like maceration, percolation, hot continuous extraction, pilot scale extraction, microwave assisted extraction and supercritical fluid extraction. GMP for the production of quality botanicals.	1	15
3.	General methods for isolation and purification of active principles from medicinal plants. Application of chromatographic techniques in isolation & characterisation of phytochemical constituents viz., paper chromatography, thin layer chromatography, column chromatography, gas chromatography (GC), high performance liquid chromatography (HPLC) and high performance thin layer chromatography(HPTLC).	0.5	10
4	Role of chemical and biological markers in standardization of herbal products	0.5	10
5	General methods for structural elucidation of natural products, Application of spectroscopy for characterization of phytoconstituents	0.5	15

Total Credits of the Course	3	

Books for Reference

Compulsory Reading:

1. Herbal Drug Technology, S. S. Agrawal, M. Paridhavi, Publisher Universities Press, 2007, ISBN 8173715793, 9788173715792

Further Reading:

2. Pharmaceutical Analysis Hiquchi, Bechmman,Hassan.
3. Methods of Drug Analysis Gearien,Graboski.
4. Text Book of BioPharmaceutic Analysis Robert Smith and JamesStewart.
5. Pharmaceutical Analysis Modern methods Part A and B Munson James.W.
6. Quantitative Analysis of DrugsGarrot.
7. Quantitative Analysis of Drugs in Pharmaceutical Formulations P. D.Sethi.

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** **U. Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks

	V. Semester End examination – 60 marks

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MAHATMA GANDHI UNIVERSITY

SBS M P E 43: IPR AND PATENTING

School Name	**School of Biosciences**					
Programme	**Msc Biotechnology/ Biochemistry/ Biophysics/ Microbiology**					
Course Name	**IPR AND PATENTING**					
Type of Course	Elective					
Course Code	**SBS M P E 43**					
Course Summary & Justification	To introduce students the concept of intellectual property and IPR					
Course Offered by	Dr Linu Mathew					
Semester	Third					
Total StudentLearningT	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning

ime (SLT)						Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisites	None					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**On completing this course, the student will be able to** **Define different international agreement on IPR**	u	
2	**Analyse the patentability of an invention and laws on plant variety protection**	An	
3	**Compare the patentability of biological entities**	U	
4	**File a patent**	S	
5	**Communicate effectively about a patent related topic both verbally and in writing**	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Introducti Introduction. Definitions General Agreement on Trade and Tariff (GATT) and World Trade Organizations Establishment and functions of GATT, WTO, and WIPO. WTO Guidelines and Summits. Physical and Intellectual Property	0.5	10
2	TRIPS Different types of intellectual property rights (IPR) - Patents, Trade mark, Trade secret, copyright and Geographical indications Requirement of patentability, Biotechnological examples of patents, trademark, trade secret and copy right	0.5	10
3.	Patenting research tools and the law: Patents as a Strategy for Protection of Intellectual Property,	0.5	10

	Benefits and Costs of Patents, Requirements for Patent Protection, patentable subjects and protection in biotechnology, international convention for the protection of new varieties – Strasbourg convention, UPOV convention. Experimental Use Exemption		
4	Patent filing and Infringement Patent application-forms and guidelines, fee structure, time frames; Types of patent applications: provisional and complete specifications; PCT and convention patent applications; International patenting-requirement, procedures, and costs; financial assistance for patenting-introduction to existing schemes; Indian Patent Act, 1970 and recent amendments Publication of patents in India Status of patenting in Europe and US. Patenting by research students, lecturers, and scientists University/organizational rules in India and abroad, credit sharing by workers, financial incentives, Patent infringement- meaning, scope, litigation, case studies and examples	1	20
5	The patentability of microorganisms, legal protection for plants and other higher organisms, new plant varieties by rights, tissue culture protocols, transfer of technology. Patentability of vectors. Licensing - Flavr Savr™ tomato as a model case, Biopiracy and case studies on patents (Basmati rice, Turmeric, and Neem)	0.5	10
	Total Credits of the Course	3	

Teaching and Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** W. Continuous Internal Assessment (CIA)

	1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks X. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Patents (2003), N.Subbaram, Pharma Book Syndicate, Hyderabad.
2. WIPO Hand book on Intellectual Property
3. IPR , Biosafety, and Bioethics Deepa Goel and Shomoni Parashar

Further Reading:

1. Revised guidelines for research in Transgenic plants (August 1998), Department of Biotechnology, Ministry of Science & Technology, Government of India, New Delhi.
2. Intellectual Property, W.R. Cornish, Sweet and Maxwell publishers, London

Web resources

1. https:// worldwide. espacenet.com
2. https:// patentscope. wipo. int
3. https:// ipindiaservices.gov.in

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MAHATMA GANDHI UNIVERSITY

SBS M P E 44: ADVANCED TECHNIQUES IN DIAGNOSTIC MICROBIOLOGY

SBS M IV E 1770 ADVANCED TECHNIQUES IN DIAGNOSTIC MICROBIOLOGY

SchoolName	**School of Biosciences**					
Programme	M.Sc. Microbiology/Biotechnology/Biochemistry/Biophysics					
Course Name	ADVANCED TECHNIQUES IN DIAGNOSTIC MICROBIOLOGY					
Type of Course	Elective					
Course Code	**SBS M P E 44**					
Course Summary & Justification	Different methods are used to detect the diseases caused by microorganisms. The syllabus content in this course has been designed with an objective to provide the basic principle and applications of various methods used in diagnostic microbiology. This will enable the students to learn the basic and advanced methods in diagnostic microbiology which will enable them to identify the research and job opportunities based on the latest developments in this subject					
Semester	Third					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent	60	20	0	40	120

	learning					
Pre-requisite	Basic understanding on diseases caused by microorganisms, different methods used to detect the diseases					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1.	Students will able to understand the process and methods in medical microbiology lab	R/U	
2.	Students will able to understand various clinical samples used for diagnostic applications	R/U	
3.	Students will able to explain the principles of methods used in medical microbiology	U/ An/E	
4.	Students will get exposed to both the conventional and rapid methods used for the microbial identification	U/An/A	
5.	Students will able to identify research and job opportunities in diagnostic microbiology	C/S	
6.	Students will able to analyze scope of technological advancement for rapid microbial identification	S/I	
**Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Introduction to diagnostic microbiology, laboratory safety, hospital epidemiology. Lab methods in Medical Microbiology, basic virology, basic mycology, Clinical material - collection and transport. Etiological agents recovered from different clinical materials	1.0	10

2	Biochemical profile based microbial identification systems, Urea breath test, Rapid antigen tests, Enzyme-Linked Immunoassay, Western blot, Advanced antibody detection, Bacterial antimicrobial susceptibility tests	1.0	20
3.	Polymerase chain reaction, Principle, applications and types of PCR in medical diagnostic field, Microbial Identification Based on PCR amplification of 16S rDNA, Sequence analysis, Application of Real Time PCR in Diagnostic Microbiology, Microbial Strain Typing Using Repetitive Sequences Advances in the Diagnosis of *Mycobacterium tuberculosis* and methicillin resistant *Staphylococcus aureus.*	0.5	10
4	Probe-Based Microbial Detection and Identification, Southern Blot Hybridization, Microarray- Based Microbial Identification and Characterization, Recent advances in medical microbiology	0.5	10
	Total Credits of the Course	3	

TeachingAnd Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction, Active co-operative learning, Seminar, Group Assignments Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar - Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. Semester End examination – 60 marks

REFERENCES

Compulsory Reading:

1. Bailey and Scott's Diagnostic Microbiology Publisher: Elsevier Health, 28 Jun 2013

2. Advanced Techniques in Diagnostic Microbiology Editors: Wu, Shangwei, Stratton,

Charles, 2012
Further Reading: 3.Textbook of Diagnostic Microbiology Hardcover, by Mahon (Author), Publisher: Elsevier Health - US; 5 edition (18 February 2014) 4.Koneman's Color Atlas and Textbook of Diagnostic Microbiology 7th Edition by Gary W. Procop MD MS , Elmer W. Koneman, Publisher: LWW; 7 edition (June 15, 2016)

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MAHATMA GANDHI UNIVERSITY

SBS M P E 45: RADIATION BIOPHYSICS

SchoolName	**School of Biosciences**
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**
Course Name	**RADIATION BIOPHYSICS**
Type of Course	Elective
Course Code	**SBS M P E 45**
Course Summary & Justification	To introduce the student to an important division of Biophysics-Radiation Biophysics To familiarize the topics of Radiation and Radioactivity, its interactions, biological effects, dosimetry, hazards, protection and application in medicine, industry and agriculture The course is designed to provide an overview of different imaging

	techniques used in medical field					
Semester	Third					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of** Radiation biophysics					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To describe various kinds of radiation and radiation units	E	
2	To explain the various biological effects of radiation	U/ An	
3	To narrate how to detect and measure radiation	R	
4	To explain how to protect from radiation exposure	S	
5	To describe the use of radioisotopes in medicine, industry and agriculture	E	
6	To discuss about the biomedical imaging techniques	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Radioactivity:** Laws of radioactivity, α, β, γ rays. Properties of electromagnetic radiation. Radiation units; Exposure and Dose, Dose equivalent unit, KERMA, Absorbed dose and Derived Units- Equivalent Dose and Effective dose, Dose rate.	0.5	10

	Interaction of radiation with matter- Bremsstrahlung, Photoelectric effect, Compton effect, Ion pair production. Interaction, absorption and scattering of electron. Heavy charged particles and Neutrons. Attenuation coefficient and absorption coefficient. HVL, Mean free path, Absorption edges, LET, Relative biological effectiveness (RBE)		
2	**Biological effects of radiation:** Radiolysis of water, Production of free radicals & their interactions, Competition kinetics, Diffusion kinetics & Physicochemical effects, Role of scavengers, G-value, Genetic Effect of radiolysis, Chromosomal breakage and Aberrations Direct and Indirect action, Oxygen and temperature effect, OER. Target theory, Single hit & Multi hit theory, Multi target theory, Calculation of target, Mass, Volume & Molecular weight, Effect of radiation on Nucleic acids, Proteins, Enzymes & Carbohydrates, Somatic and genetic effects of radiation, Stochastic and deterministic effects, early and late effects, Radiation sickness, Radiation syndrome, Haemopoietic syndrome, G.I syndrome, CNS syndrome, Acute radiation damage, Early and late effects of radiation, Effect of chronic exposure to radiation. Acute radiation damage, LD-50, Dose effect relationship. Cell recovery and modification of Radiation damage	0.5	10
3.	**Radiation dosimetry**: Principles of radiation detection and measurement- Dosimetry- General requirements of Dosimeters, Radiation sources, Telegamma Unit (Cobalt unit), Gamma chamber, Nuclear reactors, Thermal & fast neutron sources. Dosimeters- Basic principles, Design & Working of physical dosimeters- Ionization chamber, Proportional counters, GM- Counter, Concepts of Gas amplification, Resolving time & Dead time, Scintillation Detectors, Thermolumeniscent	1.0	20

	Dosimeter, Semiconductor, Surface barrier & Lithium detectors, Area survey meter & Pocket dosimeter, Film badge, General principle of chemical dosimetry, Salient Features of Chemical dosimeter, Dose evaluation formula for chemical dosimetry, Principles of radiolytic reaction, Experimental methods- Influencing factors of Fricke dosimeter methyl orange, FBX dosimeter, Free radical dosimeter, Ceric sulphate dosimeter, PMMA, PVC, chlorobenzene dosimeter, High & low dose indicators		
4	**Radiation Hazards and Protection**: Natural and man-made radiation exposures, maximum possible dose, Radiation hazards- external and internal radiation hazards. Radiation protection measurement in industrial establishment, Radioisotope labs, diagnostic and therapeutic installation and during the transportation of radioactive substances, Disposal of radioactive wastes.	0.5	10
5	**Applications of radiation-** Radioisotopes in Biology, Agriculture, Plant breeding, Plant Physiology, Medicine. Internally administered isotopes. Radioiodine in thyroid function analysis. Renal, liver and lung function analysis. Radio Immuno Assay, Radiotracer techniques. Auto radiography. Specialized radio isotopic applications in industries **Biomedical imaging techniques-** Principle of analogue and digital imaging, Ultra sound imaging, Nuclear resonance imaging, X-ray imaging and CT scan, Principle of tomographic techniques, Computerised tomography, positron emission tomography, application and interpretation of image	0.5	10
	Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** Y. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks Z. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Glenn.F. Knoll., Radiation detection and Measurement; III Edition,John Wiley & Sons, Inc.
2. Edward L. Alphen., Radiation Biophysics©, Prentice Hall

Further Reading:

1. Frank.H. Attix., Introduction to Radiological Physics & Radiation dosimetry
2. Wagner, Szabo, Buchanan., Principles of Nuclear medicine.
3. Orton, C.G., Radiation Dosimetry: Physical and Biological aspects.
4. Girish Lahari- Nuclear Physics,Mohit Books International.
5. S.P.Yarmonenko;Radiobiology, Mir Publishers.
6. JozsefKonya.Noemi M. Nagy; Nuclear and Radiochemistry,Elsevier insight

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MAHATMA GANDHI UNIVERSITY

SBS MP E 46: ALGAL BIOFUEL TECHNOLOGY

School Name	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**ALGAL BIOFUEL TECHNOLOGY**					
Type of Course	Elective					
Course Code	**SBS MP E 46**					
Names of Academic Staff & Qualifications	**Dr J G RAY**					
Course Summary & Justification	The course is to introduce the nature and principle of different form of biomass energy, familiarize the principle in the selection of suitable biomass fuels for different bioenergy applications and explain the advantages and limitations of biofuels over traditional fuels such as coal and oil.					
Semester	Third					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	E.g., Authentic learning Collaborative learning Independent learning	54	18	0	28	100
Pre-requisite	**Knowledge in Botany at the Graduate level**					

No.	Expected Course Outcome	Learning Domains	PSO No.
1	Develop a critical knowledge of the concept of biomass energy	R/U/A	
2	Identify the ecological importance of algae as a biomass resource	U/A	
3	Acquire the basic skills of utilization of algae as a biofuel resource	U/An/Ap	
4	Understand the importance of algae as a sustainable energy resource	U/An	
5	Understand the importance of algae as a sustainable energy resource	U/ An	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

Module No	Module content	Credits offered
1	Introduction – Current situation and overview on different energy sources – petroleum fuels – scope, limitations and challenges; various alternative energy forms – wind, geothermal, solar – limitations and scope; Basic characteristics of biomass as source of energy – 1st generation, 2nd generation and 3rd generation biofuels – scope and limitations Significance of algal biomass resource – classification of algae – basic morphology and taxonomy of algae- morphological characteristics of the three major groups – cyanobacteria – green algae and diatoms – micro algae and macro algae in biomass production.	1.0
2	Basic characteristics of algal feed stocks – cultivation – Photoautotrophic vs. Heterotrophic - Open vs. Closed Systems - Scale-Up Challenges, ProcessDevelopment-Scale and Integrated Bio-refinery Stability of Large-Scale Cultures, Scalable System Designs: Maintaining Productivity, Nutrient Sources, Sustainability, and Management, Water Management, Conservation, and Sustainability, fermentation tanks – closed bioreactors – open ponds – scope and limitations of each kind - Harvesting/ dewatering of biomass – Ultrasonic Harvesting, Filtration, Flocculation and Sedimentation, Flocculation and Dissolved Air Flotation, Centrifugation, Other Harvesting Techniques; Drying, Microalgae Drying Methods	1.0
3	Extraction - Lipid Separations and Extractions from Algae, Physical Methods of Extraction and/or Cellular Biomass, Pre-treatment, Microwave Assisted, Pulsed Electric Field, Ultrasonic , Catalytic Methods, of Extraction and/or Cellular Biomass Pre-treatment, Acid/Base Hydrolysis,	0.5

	Solvent-Based Extraction of Lipids Solvent Extraction Accelerated Solvent Extraction, Mixed Solvent Extraction, Supercritical Fluid Extraction, Switchable Solvents, Comparison of Extraction Methods, Lipid Extraction Challenges, Presence of Water Associated with the Biomass, Separation of Desired Extracts from Solvent Stream	
4	Production of Biofuels from Algae through Heterotrophic Fermentation or by Direct Secretion, Alcohols , Alkanes, Processing of Whole Algae, Pyrolysis, Gasification, Anaerobic Digestion of Whole Algae, Supercritical Processing, Hydrothermal Processing, Conversion of Extracted Algae , Chemical Transesterification, Direct Transesterification of Lipids into Fatty Acid, Methyl Esters, Carbohydrate and Protein Fermentation, Biochemical (Enzymatic) Conversion, Catalytic Transesterification, Conversion to Renewable Diesel, Gasoline, and Jet Fuel, Processing of Algal Residuals after Extraction	0.5
	Total Credits of the course	3

Books for References

Compulsory Reading:

1. Carney, Laura T., and Todd W. Lane. 2014. "Parasites in algae mass culture." Frontiers in Microbiology 5, Article 278. doi:10.3389/fmicb.2014.00278 2. Chisti, Yusuf. 2007.
2. "Biodiesel from microalgae." Biotechnology Advances 25 (3): 294– 306. doi:10.1016/j. biotechadv.2007.02.001.
3. Bracmort, K. 2014. Algae's Potential as a Transportation Biofuel. Congressional Research Service Report 7-5700. https://www.fas.org/sgp/crs/misc/R42122.pdf
4. Darzins, A., P. Pienkos, and L. Edye. 2010. Current Status and Potential of Algal Biofuels Production. IEA Bioenergy Task 39. Report T39-T2.
5. http://www.fao.org/uploads/media/1008_IEA_Bioenergy_-_Current_status_and_potential_for_algal_biofuels_production.pdf

Further Reading:

6. H.Verachtert et al.: Ethanol production by immobilized microorganisms, 1984, Katholieke Universitiet te Leuven, p.21104 6
7. Blanken, W, P. R. Postma, L. de Winter, R. H. Wijffels, and M. Janssen. 2016. "Predicting microalgae growth." Algal Research 14: 28–38. doi:10.1016/j.algal.2015.12.020.
8. Coons, J. E., D. M. Kalb, T. Dale, and B. L. Marrone. 2014. "Getting to low-cost algal biofuels: A monograph on conventional and cutting-edge harvesting and extraction technologies." Algal Research 6 (B): 250–70. doi:10.1016/j.algal.2014.08.005

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OPEN COURSES OFFERED BY SCHOOL OF BIOSCIENCES

MAHATMA GANDHI UNIVERSITY

SBS M PO 34: BIOTECHNOLOGY AND SOCIETY

SchoolName	**School of Biosciences**					
Programme	**MSc/ MA/ MBA (offered for schools other than School of Biosciences)**					
Course Name	**Biotechnology and Society**					
Type of Course	Open					
Course Code	**SBS M PO 34**					
Names of Academic Staff & Qualifications	**Dr Jayachandran K and Dr. Linu Mathew**					
Course Summary & Justification	1. This course is meant for PG students of MG University other than the students of School of Biosciences. 2. The course deals with the applications of Biotechnology in a societal perspective; the learner has a previous knowledge about biotechnology through mass media and their secondary school education 3. n this course they will develop a scientific understanding about biotechnology and how it benefits the society					
Semester	Third					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	80	20	0	40	140
Pre-requisite	**None**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**scribe the applications of Biotechnology in a societal perspective**	E	
2	**itically evaluate the benefits of biotechnology to society**	U/ An	

3	**nalyse the ethical and social issues related to biotechnology and intellectual property**	An	
4	**mmunicate effectively about a given topic in biotechnology and society both verbally and in writing**	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module Content	Credits	Hrs
Introduction to biotechnology: Biotechnology – a boon or a bane, Biotechnology- an interdisciplinary pursuit, public perception of biotechnology, biotechnology and the developing world, biotechnology – Indian scenario	0.5	10
Industrial and environmental biotechnology: Bioprocess and fermentation technology, enzyme technology; food and beverage biotechnology; biological fuel generation and single cell protein, GM food and controversies associated, Biosensors and biochips, Biotechnology for profit making	1	20
Genetics and biotechnology: Protoplast and cell fusion techniques, genetic engineering, whole genome sequencing, Animal cloning - ethics and applications, genetic engineering - social, moral and ethical considerations, mitochondrial evolution – tracing your routes, DNA Fingerprinting - concept and applications	1.0	20
Biotechnology in agriculture and medicine: Creation and applications of transgenic animals and plants, applications of plant and animal cell culture; gene therapy- techniques and applications	0.75	15
Protection and safety of biotechnological inventions: Patents trade secrets and plant breeders' rights, biological and physical containment, and problems of organism pathogenesis and biologically active biotechnology products, Bioterrorism, Biopiracy	0.75	15
Total Credits of the Course	4	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction: Active co-operative learning, Seminar, Group Assignments Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative

Assessment Types	**Mode of Assessment** AA. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks BB. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Biotechnology, John E Smith, Cambridge low price editions; Cambridge University press ISBN 0-521-58694
2. An introduction to genetic engineering, Desmond. T. Nicholl. Cambridge University press ISBN 81-7596-101-5

Further Reading:

1. Gene cloning and DNA analysis an introduction, T A Brown, Blackwell science publishers ISBN 0-632-05901-X
2. Molecular biotechnology, Principles and Applications of Recombinant DNA, Glick Pasternakand Patten, 4th edition ISBN 978-1-55581-498-4 Wiley International Publishers

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MAHATMA GANDHI UNIVERSITY

SBS M PO 35: MICROBIOLOGY IN EVERYDAY LIFE

SchoolName	School of Biosciences
Programme	M.Sc./M.A. in any subject
Course Name	MICROBIOLOGY IN EVERYDAY LIFE

Type of Course	Open Course					
Course Code	**SBS M PO 35**					
Names of Academic Staff & Qualifications	Dr.Radhakrishnan E.K. M.Sc.,Ph.D					
Course Summary & Justification	Microorganisms have important role to support the human life. The syllabus content in this course has been designed with an objective to provide overall understanding on importance of beneficial microorganisms and the challenges with microbial pathogens to humans. This will enable the students to identify the importance of microorganisms. With the emerging health challenges a better understanding on microorganisms will be highly beneficial for the students.					
Semester	Third					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	80	20	0	40	140
Pre-requisite	Basic interest in microbiology, understanding on importance of microorganisms and its relation with humans					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1.	Students will able to understand the importance of microbiology in various processes of daily life	R/U/I	
2.	Students will able to understand the methods to study microorganisms	R/U	
3.	Students will get exposed to the techniques used in microbiology	U/ E	
4.	Students will able to explain the role of microorganisms in relation to health and disease	U/An/A	
5.	Students will able to understand disease progression and mechanisms involved	C/S	

6.	Students will able to apply the knowledge to for better management of microorganisms for healthy life	A/S	
**Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	History and Developments in Microbiology: Prokaryotic and eukaryotic cell, Contributions from Leuwenhoek, Louis Pasteur, and Robert Koch. Microbiome, An overview of microorganisms, the bacteria and the archea. General characteristics, morphology, Structure of bacteria. Virion, viroids and prions, Eukaryotic Microorganisms., A brief introduction to microscopy, Staining of bacteria and fungi, Cultivation of bacteria and fungi, culture media and methods	1.0	20
2	Methods to control Microorganisms: Disinfection, Sterilization, Sterilizing Agents, Antibiotics, Antibiotic Sensitivity tests, Antibiotic Resistance	1.0	20
3.	Microbes in relation to health and disease: Human microbiome, Infection, source of infection, method, of transmission, Immunity, Innate and adaptive immunity, Microorganisms involved in respiratory tract infection, Meningitis, Urinary tract infection, STD, Skin infection, Nosocomial infection, Tuberculosis, Typhoid fever, Dengue, AIDS, Hepatitis, Ebola and COVID-19	1.0	20
4	**Microbes in relation to food:** Microorganisms in preparation of food materials, lactic acid bacteria, role of microorganisms in preparation of curd, cheese and cultured dairy products, probiotics, and their importance, single cell protein, Microorganisms responsible for food borne infection and intoxication. Water borne diseases- prevention and control Fermented food, milk and milk products, role of food preservative **Industrially important microbial products**: Role of Microorganisms	1.0	20

	in 74 production of bread and beer. Microbial enzymes and their uses- detergent, enzymes, therapeutic enzyme Streptokinase		
	Total Credits	4	

Teaching And Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction, Active co-operative learning, Seminar, Group Assignments Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar - Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. Semester End examination – 60 marks

REFERENCES

Compulsory Reading:

1. Microbiology. Prescott, Harley and Klein wim C Brown publishers, 2014

2. Brock Biology of Microorganisms, Michael T. Madigan, John M. Martinko, David A. Stahl, David P. Clark,14th edition, 2015

Further Reading:

3. Principles and practice of disinfection, preservation and sterilization – Russel AD et al., Blackwell Scientific Publications, 2013

4.Microbiology Concepts and Applications. Pelczar Jr Chan. Creig. McGraw Hill Inc, 5 th edition, 2001 5.

5. Topley and wilson's Principles of Bacteriology, Virology and Immunology – Arnold – Heinemann, 1990

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MAHATMA GANDHI UNIVERSITY

SBS M PO 36 : ENVIRONMENT LEAD AUDITOR COURSE

School Name	**School of Biosciences**					
Programme	**III SEM Open Course**					
Course Name	**ENVIRONMENT LEAD AUDITOR COURSE**					
Type of Course	**Open**					
Course Code	**SBS M PO 36**					
Names of Academic Staff & Qualifications	**Dr J G RAY**					
Course Summary & Justification	**This course provides postgraduates with a thorough understanding of the basic principles of ecology and environment and introduces** the basic concept of ecological objects – population, community, and ecosystem- and then explain the details of the environmental auditing process. The course will equip them as environment auditors - 'Lead Auditor' as per ISO 14001 Standards. Environmental auditing is an essential process of all institutions and industrial processes to achieve sustainability in their activities, process, production and practice. It is an emerging career as well.					
Semester	Third					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg: Authentic learning Collaborative learning Independent learning	80	18	0	30	128

Pre-requisite	Students of arts/science/management/commerce at Graduate level

.	Expected Course Outcome	Learning Domains	PSO No.
	Upon completing this course, students will develop a critical knowledge of the basic principles of ecology and the environment.	R/U/A	
	They will be able to analyse environmental issues from a social perspective.	U/A	
	They will acquire the basic skills of environmental auditing. They will develop the skills of a lead auditor	U/An/Ap	
	They will develop the skills of an Environment lead auditor of ISO-14000 standard as per the British Standard Institution requirements	An/Ap	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

module No	Module content	Credits offered	hrs	
1	Introduction to Ecology and Environment science: the concept of the environment; Life as a system phenomenon- hierarchy in the system of life; Ecological objects- population, community and ecosystem; Ecology of humans, Concept of sustainable environmental quality. Environment Pollution – definition and classification; Water pollution – water quality parameters and standards, control of water pollution, wastewater treatments; Air pollution – primary and secondary pollutions, air pollution monitoring and control; Land pollution – solid waste management, recycling, reuse and recovery, problems of plastic waste	1.0	20	
2	Natural Resources and Biodiversity Conservation: classification of resources, resource depletion, preservation, conservation and restoration of resources; Concept of biodiversity – genetic, species and ecosystem diversities, principles of biodiversity	0.5	10	

	conservation, ex-situ and in-situ conservations; IUCN accounting of biodiversity – hot spots, red data book; Global environmental crisis - UNEP, UNFCC, One earth programme, globally crucial agricultural heritage (GIAH)			
3	Legal methods to sustain environment quality: environment laws – national and international environmental laws; Montreal protocol and its amendments, Kyoto protocol, constitutional provisions of environment quality in India, major environmental laws of India, environment protection act of 1986, National environment policy; Environment Impact Assessment	0.5	10	
4	Environment Audit: definition, types of audit, objectives of environmental audit, benefits of ecological audit, basic environment management philosophy, critical steps to environment audit – pre-audit, onsite audit and post-audit, step by step approach of auditing, action plan, auditor requirements; Environment Management Systems: ISO-14000-2004, model for this international standard, different clauses in ISO 14000 standard – scope, normative references, terms and definitions, EMS requirements – clauses 4.1 to 4.6	2.0	40	
	Total Credits of the course	4		
Books for References				
Compulsory Reading: 1. Ray J G (2010) Basic Principles of Ecology and Environment, Prathibha Publications, Kerala,India 2. Mehrotra A et al. (2001) A to Z of Environmental Audit,SOFEM Publ. New Delhi 3. Dash M C (1993) Fundamentals of Ecology, Tata McGraw Hills Publ. Co. New Delhi				
Further Reading: 4. Singer FD (2016) Ecology in Action, Cambridge University Press 5. Chapman JL and Reiss MJ (1998) Ecological Principles and Applications, Cambridge University Press, London 6. Trivedi RK (Ed) International Encyclopaedia of Ecology and Environment (Volumes 1-30), IIE, New Delhi 7. 7. Ramade F (1981) Ecology of Natural Resources, John Wiley and Sons, New York				

MAHATMA GANDHI UNIVERSITY

SBS M PO 3 : SYSTEM BIOLOGY

SchoolName	**School of Biosciences**					
Programme	**MSc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**SYSTEM BIOLOGY**					
Type of Course	Open course					
Course Code	**SBS M PO 37**					
Names of Academic Staff & Qualifications	**Dr. R. Harikumaran Nair** MSc, PhD					
Course Summary & Justification	is course is designed to provide an overview of human physiology. urse topics will include the various systems of the body, functions of each system, and interrelationships to maintain the internal environment. The course also provides inputs to physiological stress and adaptive strategies to overcome stress					
Semester	Third					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHou

						rs
	Authentic learning Collaborative learning Independent learning	80	20	0	40	140
Pre-requisite	**Basics Knowledge in Biology**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	udents should be capable of effectively communicating how the human body works	U	
2	udents should describe the interdependency and interactions of the systems	A	
3	students should be able to explain contributions of organs and systems to the maintenance of homeostasis	E	
4	The content of the course will elicit curiosity in functioning of human body	I	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module Content	Credits	Hrs
Body organization, cells, tissues, organ and organ systems, body fluid compartments, reflex, biological rhythms	0.25	5
Cell membrane, cell organelles, movement of molecules across cell membranes, diffusion, osmosis, endocytosis, exocytosis	0.25	5
Neuron, basic principles of electricity, neuronal potentials, neuronal communications, brain, spinal cord, different nervous systems, somatic sensation, vision, hearing, chemical sense, motivation, emotion, learning, memory	0.5	10
Muscles, muscle contraction, body movement, hormones, hormone disorders, heart, cardiac functions, blood, hypertension, respiration, gas transport between lungs and tissues, respiratory problems	1.5	30

Kidney, dialysis, digestion and absorption of food, diabetes mellitus, increased plasma cholesterol, body temperature, gametogenesis, male and female reproductive functions	1.5	30
Total Credits of the Course	4	80

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** CC. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks DD. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading

1. Systems Biology: Definitions and Perspectives. Alberghina, L. and Westerhoff, H,
2. Essentials of Medical Physiology. K Sembulingam&PremaSembulingam

3.Biochemistry and Physiology of the cell. An introductory text second edition- Edwards, N. A Hassall, K.A

Further Reading:

1.Vander's Human Physiology- The mechanism of body function. Widmaier, Raff & Strang
2.Textbook of Medical Physiology. Arthur.C. Guyton&John.E. Hall
3.Physiological basis of Medical Practice. John.B. West

4.Human Physiology: an integrated approach- Silverthorn, Dee Unglaub

5.Principles of anatomy and physiology- Tortora, Gerald J Derrickson, Bryan

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MAHATMA GANDHI UNIVERSITY

SBS M PO 38 : SUSTAINABLE AGRICULTURE

School Name	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**SUSTAINABLE AGRICULTURE**					
Type of Course	Open course					
Course Code	**SBS M PO 38**					
Names of Academic Staff & Qualifications	**Dr J G RAY**					
Course Summary & Justification	The course is to introduce the concept of sustainable agriculture, especially its principles of ecological sustainability. The course will equip students to understand the concept of organic farming. It will enable an understanding of plant nutrient management as well as pest management in sustainable agriculture. Organic farming is becoming an internationally significant agricultural practice, and the knowledge has global significance. Interdisciplinary biology students with a good understanding of organic farming will enable our students to find suitable job opportunities in such farming industries.					
Semester	Third					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning

						Hours
	E.g., Authentic learning Collaborative learning Independent learning	60	18	0	28	106
Pre-requisite	**None**					

No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**	
	Students will develop a critical knowledge of the basic principles of sustainable agriculture	R/U/A		
	They will be able to analyze environmental issues related to chemicalized agriculture	U/A		
	They will acquire the basic skills of sustainable organic agriculture	U/An/Ap		
	They will develop the skills to evaluate different kinds of farming	An/Ap		
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) nd Appreciation (Ap)				

Modul e No	Module content	Credits offered	hrs
	Introduction to Sustainable agriculture: Concept of ecological sustainability and sustainable agriculture-Natural, Ecological and organic farming – definition, concepts, and practices – management, principles, methods, merits and demerits.	1.0	20
	Challenges to Sustainable agriculture – Productivity vs sustainability; Soil organic matterdecomposition, C: N ratios, mineralization and immobilization processes, hummus, the role of organic matter in soil quality – natural way to prevent soil degradation and erosion, types and control measures. Soil related water pollution- sources, different pollutants in soils and their managements Plant nutrient management in sustainable agriculture: Bio-availability of nutrients in soils, deficiency symptoms on plants, nutrient interactions and chelated micronutrients.Bio-fertilizers – benefits - classifications, production - maintenance and application	1.0	20
	Organic Manures – bulky and concentrated – FYM – Biocomposting, Compost – rural, urban, vermicompost and coirpith; Panchagavya preparation and other organic nutrients application - Enrichment of	1.0	20

	organic manures; Sewage and sludge; Green manures – potentials and limitations; Quality parameters of organic manures and specifications – Biofertilizers -		
-	Biopesticides and biological control agents: Types of biocontrol agentsbiological agents and pheromones, control of weeds, diseases and insect pests and field sanitation - competition, predation, antibiosis and fungistatic Efficacy of traditional biopesticides - Botanical insecticides- beneficial insects like the honeybee, lac insect, silkworm and pollinators Biological control - concepts and potentialities for managing soil-borne pathogens. Types of biological interactions, competition, 1.078 mycoparasitism; Mycorrhizal associations, Biodynamic products, Biodynamic composting, Liquid manure, Influence of Bio-dynamic products on crop production. Visit Organic Farms	1.0	20
	Total Credits of the course	4	
	Books for References		
Compulsory Reading: 1. Dahama AK (2007). Organic Farming for Sustainable Agriculture. 2nd Edn. Published by AGROBIOS (India) Jodhpur 2. National Standards Programme for Organic Production and Organic Products (2000) Department of Commerce, Ministry of Commerce and Industry, Govt. of India			
Further Reading: 3. Gehlot D (2005). Organic Farming: Standards, Accreditation, Certification and Inspection, AGROBIOS (India) Jodhpur 4. Gupta PK (2007). Soil, Plant, Water and Fertilizer Analysis Published by AGROBIOS (India), Jodhpur 5. Sadasivam S and Manickam A (1992). Biochemical Methods for Agricultural Sciences Wiley Eastern Limited and Tamil Nadu Agricultural University, Coimbatore			

MAHATMA GANDHI UNIVERSITY

SBS M PO 39: ECOLOGY AND SOIL FERTILITY

School Name	**School of Biosciences**

Programme	**III Sem Open Course**					
Course Name	**Ecology of Soil Fertility**					
Type of Course	Open					
Course Code	**SBS M PO 39**					
Names of Academic Staff & Qualifications	**Dr J G RAY, Ph D in Soil Ecology**					
Course Summary & Justification	The course is designed to help postgraduates of both arts and science know what soils are and how they form and sustain them. The course will help them understand the nature and importance of soil fertilityand its natural biological maintenance. They will also know the role and importance of soil biodiversity in soil fertility and its sustenance. Since farming has become an important activity to ensure ecosystem sustainability, it has become imperative for every educated person to understand the scientific principles of soil fertility and its natural maintenance. Organic farming is also becoming an essential career for educated youth.					
Semester	Third					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	E.g., Authentic learning Collaborative learning Independent learning	80	18	0	30	128
Pre-requisite	**none**					

No.	**Expected Course Outcome**	
	Students will develop a critical knowledge of the concept of soil fertility	
	They will be able to understand the vitalnatural components of soil fertility	
	They will learn to account for various soil biodiversity components and their significance	
	They will know sustainable management of soil fertility	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)		

Module No	**Module content**	**Credits offered**	

1	Concept of soils – soil and parent materials– soil formation – role of climate and vegetation in soil formation – soil profile, soil taxonomy; water relations of soils – hygroscopic, capillary and field water content – run-off water – factors affecting percolation	1.0	20
2	Soil Physics and Chemistry; soil physical properties - colour and texture, soil structure - aggregate formation, aggregate stability, capillarity, porosity; Soil chemistry, pH, carbon in soils –humus – its chemistry and role in soil, exchangeable and soluble cations and anions in soils soil as a buffer system; soil amendments – problems of tillage – problems of irrigation – problems of chemical fertilizers and liming	1.0	20
3	Soil biology – the role of soil biota – soil fauna and flora - soil ecological processes and microbial function - decomposition - introduction, overview, fragmentation, factors controlling decomposition, carbon, nitrogen and phosphorus cycles in soils, transformations of nitrogen nitrification-immobilization-volatilization, denitrification, soil ecosystem management and soil biota	1.0	20
4	Agriculture – traditional versus modern – problems of chemicalized agriculturecontrol of soil degradation - desertificatication of soils – soil reclamation – soil conservation –prevention of soil erosion - mulching, contour bunds – sustainable soil fertility - ecology of soil fertility – principles of ecological and organic farming - climate change, global warming and soil ecology	1.0	20
	Total Credits of the course	4	

Books for References

Compulsory Reading:

8. Nyle C Brady (1984) Nature and properties of Soil, Mc Milan Publishers
9. Ray J G (2010) Basic Principles of Ecology and Environment, PrathibhaPubli., Kerala, India

Further Reading:

10. Colemn DC et al. (2003) Fundamentals of soil ecology, Elsevier
11. Christian Ditchfield (2003) Soils, Children's Press, Dublin
12. James BN (2003) The world beneath our feet: A guide to life in the soil, Oxford University Press

MAHATMA GANDHI UNIVERSITY

SBS M PO 40 : INFECTIOUS DISEASE MANAGEMENT

SchoolName	School of Biosciences					
Programme	M.Sc./M.A. in any subject					
Course Name	INFECTIOUS DISEASE MANAGEMENT					
Type of Course	Open Course					
Course Code	**SBS M PO 40**					
Names of Academic Staff & Qualifications	Dr.Radhakrishnan E.K. M.Sc.,Ph.D					
Course Summary & Justification	Infectious diseases cause significant threat to the existence of humans. The syllabus of this course has been designed to introduce the importance and relevance of infectious diseases. This also include the mode of transmission of various infectious diseases and the diagnostic methods used for various infectious diseases This will enable the students to identify the importance of infectious diseases and the microbial basis of the same. With the emerging health challenges a better understanding on infectious diseases will be highly beneficial for the students.					
Semester	Third					
Total Student Learning	Learning Approach	Lectur	Tutori	Practi	Other	Total

Time (SLT)		e	al	cal	s	LearningHours
	Authentic learning Collaborative learning Independent learning	80	20	0	40	140
Pre-requisite	Basic interest in infectious diseases and microbiology					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1.	Students will able to understand the importance of infectious diseases	R/U	
2.	Students will able to understand the types of organisms causing infectious diseases	R/I/ U	
3.	Students will learn the mode of transmission of infectious diseases	U/ E	
4.	Students will able to explain the infectious diseases and its basis	U/I/A	
5.	Students will able to understand diagnosis of infectious diseases	C/S	
6.	Students will able to apply the knowledge on infectious diseases for better its management	S/C	

**Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)*

COURSE CONTENT

Module No	**Module Content**	**Credits**	**Hrs**
1	Infectious disease, etiological agents-bacteria, fungi, viruses, prions, protozoan. Special focus on COVID-19, Reservoir- human, environment. Carriers- incubatory, inapparent infection, convalescent and chronic carrriers. Mode of transmission- direct and indirect. Portal of entry- respiratory, genitourinary, alimentary, skin and transplacental	1.0	20
2	Infectious disease- acute respiratory infections, diarrheal diseases, hepatitis, HIV, tuberculosis, sexually transmitted diseases, malaria,	1	20

	and other vector-borne diseases		
3.	Lab diagnosis of infectious disease, sample collection, sample processing microscopy, culture, immunological methods, nucleic acid based identification methods and non-nucleic acid based identification methods	1	20
4	Infectious disease management, treatment, antibiotics- types of antibiotics, mode of action, antibiotic resistance, antiviral, antifungal, and antibacterial agents, immunization and infectious diseases, vaccination against major infectious diseases, types of vaccines	1.0	20
	Total Credits	4	

Teaching And Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction, Active co-operative learning, Seminar, Group Assignments Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar - Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. Semester End examination – 60 marks

REFERENCES

1. Bailey and Scott's Diagnostic Microbiology Publisher: Elsevier Health, 28 Jun 2013

2. CURRENT Diagnosis & Treatment in Infectious Diseases, Walter R. Wilson and Merle A. Sande 3. Fundamentals of Molecular Diagnostics (1st Edition) By David Bruns Edward Ashwood Carl Burtis : Elsevier. 2007

Further Reading:

3. Textbook of Diagnostic Microbiology Hardcover, by Mahon (Author), Publisher: Elsevier Health - US; 5 edition (18 February 2014)
4. Koneman'sColor Atlas and Textbook of Diagnostic Microbiology 7th Edition by Gary W. Procop MD MS, Elmer W. Koneman, Publisher: LWW; 7 edition (June 15, 2016).
5. Advanced techniques in Diagnostic microbiology. Yi-wei Ting, Charles W. Stratton: Springer
7. Sherris Medical Microbiology (5th edition) by Kenneth J. Ryan, C. George Ray
5. Infectious Disease: Pathogenesis, Prevention and Case Studies By Nandini Shetty, Julian W Tang, Julie. Wiley- Blackwell (April, 2009).

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MAHATMA GANDHI UNIVERSITY

SBS M PO 41: PROBIOTICS AND NUTRACEUTICALS

School Name	**School of Biosciences**
Programme	**MSc/ MA/ MBA (offered for schools other than School of Biosciences)**
Course Name	**Probiotics& Nutraceuticals**
Type of Course	**Open Course**
Course Code	**SBS M PO 41**
Names of	**Dr.Keerthi TR**

Academic Staff & Qualifications						
Course Summary & Justification	1.The cover concept of nutraceuticals/functional food - extra health benefits in addition to the basic nutritional value of food. 2.Enable students to recognize the link between nutrition, health and diseases 3.Identify major types of health foods and nutraceutical products in the market. Role of Probiotics & Prebiotics to maintain health. 4.Students get exposure towards the market opportunity of nutraceuticals and the nutraceutical industry					
Semester	Third					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	80	20	0	40	140
Pre-requisites	Basics of Health and Nutrition.					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**On completing this course, the student will be able to Explain the classification and types of nutraceuticals/functional foods. Describe the role of nutraceuticals in lifestyle diseases.**	**U/E**	
2.	**Describe the nutraceuticals from plant origin including algal nutraceuticals & their health benefits**.	**U/R**	
3.	**Explain various nutraceuticals of animal origin & their therapeutic applications**.	**An/U**	
4.	**Illustrate the health benefits & mechanism of probiotics & prebiotics. Describe the various probiotics & prebiotics available in the market & their production & specific applications**	**U/A**	
5.	**Communicate effectively about a chosen topic in Probiotics & Nutraceuticals both verbally and orally**	**An/A**	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	**Concept of Functional Foods/Nutraceuticals**: Definition and classification of nutraceuticals, dietary supplements, fortified foods, functional foods and Phyto- nutraceuticals. Scope	**1.0**	**20**

	involved in the industry, Indian and global scenario. Relation of functional foods/ Nutraceutical (FFN) to foods & drugs. Applications of herbs to functional foods. Concept of free radicals and antioxidants; Nutritive and Non-nutritive food components with potential health effects. Role of nutraceuticals in the prevention and treatment with special reference to diabetes mellitus, hypertension, hypercholesterolemia. Concept of antioxidants - use of antioxidants as dietary supplements in prevention and treatment of cancer, obesity and stress		
2.	**Nutraceuticals of plant origin:** Nutraceuticals in Fruits and Vegetables and their Health Benefits; Sources and role of Isoprenoids, Isoflavones, Flavonoids, carotenoids, Tocotrienols, polyunsaturated fatty acids, sphingolipids, lecithin, choline. terpenoids. Vegetables, Cereals, milk and dairy products as Functional foods. Health effects of common beans, *Capsicum annum*, mustards, , garlic, grape, citrus fruits. **Algal nutraceuticals:** Micro & macro algae as nutraceuticals. Algae as source of omega - 3 fatty acids, antioxidants and minerals - extraction and enrichment	**1.0**	**20**
3.	**Nutraceuticals of animal origin:** Animal metabolites - Sources and extraction of nutraceuticals of animal origin. Examples: chitin, chitosan, glucosamine, chondroitin sulphate and other polysaccharides of animal origin, uses and applications in preventive medicine and treatment. fish oils, and sea foods	**1.0**	**20**
4	**Probiotic & Prebiotic:** Concept of prebiotics and probiotics - principle, mechanism, production and technology involveddifferent forms available in the market. Benefits &applications - examples of bacteria used as probiotics, Types & use of prebiotics in maintaining the useful microflora & other health benefits .Other biotic approaches for maintaining good health. Market opportunities of nutraceuticals	**1.0**	**20**
Total Credits of the Course		**4**	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** EE.Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10

	3. Write a detailed report on a given topic based on research findings and literature search – 10 marks FF. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Shi, J. Asian Functional Foods CRC Press 2005
2. Webb, G.P. Dietary Supplement and Functional Foods Blackwell 2006 .
3. Shibamoto T. Functional food and health, Oxford University Press, 2008

Further Reading:

1. Shi, J. Functional Food Ingredients and Nutraceuticals: Processing Technologies CRC Press 2007
2. Bagchi D. Nutraceutical and functional food regulations in the United States
3. and around the world, Elsevier/Academic Press, 2008.
4. Guo M. Functional foods: principles and technology, CRC Press, 2009.
5. J.Paulo Sousa e Silva.,Ana.C.Freiles.Probiotic Bacteria .Pan slanford publishing Pte.Ltd
6. FaizelBux. Biotechnological Application of Microalgae.CRC Press.
7. Wayne.RBidlack.,Roymond L Rodringuez.NutritionalGenomics.CRC Press.
8. Frances Sizer., Elecener Whitney Nutrition concept and contraversesisWordswerth Publishers.
9. Thomas J Mont Ville.,Kart R Matthews 7 Kalmia E Kniel Food Microbiology, ASM Press.

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FOURTH SEMESTER

MAHATMA GANDHI UNIVERSITY

SBS M P C 56: BIOPHYSICAL CHEMISTRY

SchoolName	**School of Biosciences**
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**
Course Name	**BIOPHYSICAL CHEMISTRY**
Type of Course	Core
Course Code	**SBS M P C 56**
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S**
Course Summary & Justification	To understand the student about the link between Physics, Chemistry and Biology To perceive the knowledge about ligand binding, models of ligand binding, light scattering, calorimetry, x-ray crystallography The course also provides information about mass spectrometry and nanotechnology
Semester	Fourth

Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120

Pre-requisite	**Basics of** biophysical chemistry methods

CO No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**

COURSE OUTCOMES (CO)

1	To explain covalent and coordinate bond formation	E	
2	To describe the process of ligand binding interaction and various models	U/ An	
3	To narrate the bio-molecular interactions and their role in modulation of biological processes	R	
4	To describe Isothermal titration and Differential scanning calorimetry in the study of proteins and carbohydrates	S	
5	To explain the mechanism of X-ray Crystallography	E	
6	To determine the instrumentation and mechanism of different type of mass spectrometers	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module Content	Credits	Hrs
Atoms and chemical bonds: Electron theory of valance, Chemical bonding and interaction, synthesis and cleavage of covalent bonds, Reactive species-electrophiles, nucleophiles and radicals, Structure, bonding and special properties of water. Coordinate bond, coordinate bond formation in transition metals Bonding of iron in haemoglobin, cobalt in Vitamin B12, magnesium m in chlorophyll,	0.5	10
Ligand binding interaction: Ligand interaction at equilibrium, identical and independent sites model, Scatchard plot, multiple classes of independent sites, interaction between binding sites, Allosterism, MWC model, sequential model, oxygenhaemoglobin binding, binding of two different ligands, cooperative binding, anti-cooperative binding and excluded site binding, energetics & dynamics of binding, binding of immunoglobulin and DNA binding protein. Static and dynamic light scattering, Surface Plasmon resonance, isothermal titration calorimetry and differential scanning calorimetry	1.0	10

X-Ray Crystallography-Crystals, Molecular crystal symmetry, Miller indices, reciprocal Lattice, Ewalds Construction, X ray diffraction by crystals, Bragg's Law & Bragg's diffraction equation, laue powder and rotation methods & Laue's equations, diffraction methods-Laue's method, Weissenberg diffraction camera and powder method. Protein X-ray crystallography- production of suitable crystals, Acquisition of the diffraction pattern, Determination of phases, Heavy atom replacement method, Calculation of the electron density and refinement. Phase problem in crystallography. Neutron diffraction, Electron diffraction, Synchrotron diffraction, Fibre diffraction and its application	1.0	20
Mass spectrometry: Principles of operation and instrumentation, ion formation and types; molecular ions, meta stable ions, fragmentation processes, fragmentation patterns and fragment characteristics in relation to parent structure and functional groups, mass spectrum; its characteristics, presentation and interpretation. Sector Mass spectrometer, Quadrupole Mass spectrometer, Ion trap Mass spectrometer, Time-of flight Mass spectrometer, Fourier transform Mass spectrometer. Combination with chromatographic methods, Biological application	1.0	10
Introduction to Nanotechnology: Orientation and introduction towards Nanotechnology, Classification and nomenclature of Nanomaterials, synthesis and Characterizationof Nanomaterials, Structural and functional principles of Bionanotechnology, protein and DNA based Nanostructures, Nanoparticles in agricultural and food diagnostics, Applications of Nanotechnology	0.5	10
Total Credits of the Course	4	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** GG. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks HH. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Biophysical chemistry, Principles and Techniques- Himalaya Publ.House
2. X-ray structure determination, George.H. Stout, Lyle H Jensen, John Wiley & Sons.
3. Methods in Modern Biophysics, Bengt nolting, II edition

Further Reading:

1. Biophysical Chemistry, P.R.Bergethon, E.R.Simons
2. Principles of Instrumental Analysis, Skoog, Holler, Nieman
3. Biophysics, VasanthaPattabhi, N.Gautham
4. Biophysical Chemistry, Techniques for the study of biological structure and function, CantorSchimmel part II
5. Bioseparations Science and Engineering: Roger G Harrison, Paul Todd, Scott R. Rudge, Oxford University Press.
6. Methods in Modern Biophysics, Bengt Nolting.
7. Practical Protein crystalloigraphy, Duncan.E, Mc Rec
8. Biomaterials: A Nano Approach,S Ramakrishna, M Ramalingam, T.S. Sampath Kumar, Winston O. Soboyejo,Published by CRC Press
9. Bionanotechnology: Lessons from Nature, D S. Goodsell, by John Wiley & Sons, Inc
10. Klabunde, K.J. (Ed.), "Nanoscale Materials in Chemistry", John Wiley & Sons Inc. 2001

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MAHATMA GANDHI UNIVERSITY

SBS M P C 57: Laboratory Course 5- Biophysics

SchoolName	**School of Biosciences**					
Programme	**Msc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**Lab Course 7**					
Type of Course	Core					
Course Code	**SBS M P C 57**					
Names of Academic Staff & Qualifications	**Mrs. Resmi S. S** Prof (Dr) Hari kumaran nair Guest faculty					
Course Summary & Justification	To familiarize the student to various Biophysical, Physiological, Radiation and Bioinformatics techniques To develop laboratory skills on Separation techniques, Physiological experiments, effects of radiation and Bioinformatics tools					
Semester	Fourth					
Total StudentLearning	Learning Approach	Lectur	Tutori	Practi	Other	Total

Time (SLT)		e	al	cal	s	LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of** physiological and biophysical methods					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To describe the UV analysis of nucleic acids and proteins	E	
2	To perform IR spectroscopy and Ultracentrifuge	U/ An	
3	To perform molecular docking of target versus inhibitors	R	
4	To isolate RNA and plasmids	S	
5	To determine spirometry analysis	E	
6	Molecular Dynamic Simulation	An/ C	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module Content	Credits	Hrs
Analytical Biophysics • To study the Erythrocytes Membrane Permeability and Transport effects of Hypotonic & Hypertonic shock • To determine the partial characteristics of Membrane Protein by SDS-PAGE. • To determine the effects of UV on E. coli and elucidation of cell survival curve. • Determination of effects of UV on cell membrane	0.5	10

Spectroscopic analysis • Determination of effects of UV on cell membrane. • To study the renal stone using Infra-Red (IR) Spectroscopy. • To determine the oil content of oil seeds using Nondestructive IR Spectrophotometry. • Denaturation & Renaturation of DNA. • To isolate the chloroplast and characterize the chloroplast membrane protein • 6. To determine the molecular weight of biomolecules using ultracentrifuge	0.5	10
Molecular Docking • Identification of Nucleic Acid Binding Proteins Using Nondenaturing Sodium DecylSulfate Polyacrylamide Gel Electrophoresis (SDecS-Page) • Multiple sequence alignment and Conserved Amino acid residues. • Analysis and study of sequence using different types of BLAST tool • Molecular docking by using AutoDock4 software	1.0	20
Molecular Biophysics • Conformation of Nucleic acid by Spectral study. • To isolate RNA • To isolate and characterize Plasmid DNA. • To hydrolyze the t-RNA and separation of Nucleotides by TLC and paper chromatography • Restriction digestion and agarose gel electrophoresis of DNA • Demonstration of various advanced microscopic techniques	0.5	10

Physiological Biophysics • ECG, (demonstration). • Spirometry, Body temperature, pulse sensors, • Breath holding time, Measure the pulse rates, Heart beat rate, BP measurement	0.5	10
Total Credits of the Course	3	

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** II. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks JJ. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

3. Biochemical methods by S Sadasivan and A Manickam. New Age international publishers
4. Biotechnology: A laboratory course by Becker J.M.
5. Bioinformatics Practical Manual by Mohammed Iftekhar and Mohammed Rukunuddin Ghalib

Further Reading:

7. An introduction to practical Biochemistry by D. T. Plummer, Mc Graw Hill.
8. Laboratory manual in Biochemistry by J. Jayaraman, Wiley Eastern Limited.
9. Introductory Practical Biochemistry by S. K. Sawhney and Randhir Singh, Narosa

10. Experimental Biochemistry: A student companion by BeeduSashidhar Rao and Vijay Deshpande, I.K. International Pvt. Ltd., New Delhi.
11. Lab manual in Biochemistry by J. Jayaraman, Wiley Eastern Limited
12. An introduction to practical Biochemistry by D. T. Plummer, Mc Graw Hill.
13. Introductory Practical Biochemistry by S. K. Sawhney and Randhir Singh, Narosa

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School Name	**School of Biosciences**
Programme	**Msc Biotechnology/ Biochemistry/ Biophysics/ Microbiology**
Course Name	**SBS M P E 59: ENVIRONMENT BIOTECHNOLOGY**
Type of Course	Elective
Course Code	**SBS M P E 59**
Course Summary & Justification	Environmental Biotechnology is offered to train the students both in the theoretical and practical aspects of identifying environmental problem where a solution is possible through Biotechnological methods Enabling students in formulating ideal solution to environment problems based on green chemistry concept is the need of this time . Students have to earn a sense of Environmental concern and to get experience in the applications of Biotechnological methods for environmental protection.This course is also introduced as a part of the

	national policy effort to incorporate environmental education into the curriculum of all P.G Programme of all universities in India.					
Semester	Fourth					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisites	None					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**On completing this course, the student will be able to** Understand the effect of a specific environmental problem identified	u	
2	**Analyse** Apply the most suitable biological method for the effective treatment of the pollutant	An	
3	**Compare** Explore into the possibility of applying the developed method in the field.	U	
4		S	
5	**Communicate**	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Introducti I**ndustrial pollution causes, problems:** Air, Soil and Water pollutants, Types of pollutants characterization, Persistence and Biomagnification of Xenobiotics, recalcitrant molecules, nitroaromatic polychlorinated, biphenyls and dioxans, synthetic polymers, alkylbenzyl sulphonates, Hydrocarbons, Pesticides, Phenolics, Anilines, Inorganic pollutants, Heavy metals. Detection and Quantification of pollutants. Environmental laws	0.5	10
2	**Biodegradation, Process and application:** Microbial infallibility, types of biodegradation, factors affecting biodegradation, enzymes involved in biodegradation, catabolic plasmids, Molecular Approaches, Biogeochemical cycles, Bioleaching. Biodegradation of Hydrocarbons, cellulose, lignin, Phenoland pesticides. Application of TOC, FT/IR, GC-MS analysis in biodegradation studies	0.5	10
3.	**Industrial wastewater:** Types of industrial effluents, characterization of the wastewater. Chemical Oxygen Demand, Biological Oxygen Demand, Total organic carbon, Nitrogen contents, Suspended solids. Total heterotrophic bacterial population. Bacteriological analysis of drinking water, Presumptive, completed, and confirmed test. Treatment strategies primary, Secondary and tertiary treatment Physical, Chemical and Biological treatment. Floc based and film based strategies, aerobic and anaerobic methods	1	20
4	**Biological treatment of industrial wastewater:** Activated sludge process, different stages, Types. Oxic/Anoxic, Extended aeration methods, Nitrification and denitrification. Trickling filter process, Different stages Types, Biofilm applications, Rotating Biological contactor,UASB, Submerged aerobic filters, Fluidized Bed Reactor, Packed bed reactor, Oxidation lagoons. Bioreactors for wastewater treatment. Advanced treatment strategies Teritiary treatment methods, Disinfection, Chlorination, Chlorination dosage chlorination derived byproducts	0.5	10
5	**Solid waste management:** Solid waste, Types, Problems, Characterization and sorting of wastes. Municipal and industrial waste management, Land fills composting, stages in composting, Types of composting vermicomposting. Methanogenesis, stages in anaerobic digestion, methanogens Anaerobic reactors Biogas generation, Household treatment strategies, Present problem and Possible remedies	0.5	10
	Total Credits of the Course	3	

Teaching and Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** KK. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks LL. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1.Microbial Ecoology, Atlas and Bartha, Pearson Publication

2.Comprehensive Biotechnology—2 nd Edition,Murray Moo Young ISBN-9780444533524,Pergman

3.Industrial Microbiology, Samuel Cate Prescot and Cecil Gordan Dunn,Third edition Mac Graw-Hill

4.Waste water microbiology, Gabriel Bitton,Third edition, Wiley, ISBN-9780471717966

Further Reading:

1. Environmental Biotechnology -Theory and application , Gareth m Evans and Judith C Furlong , Wiley 2003

2. Envoronmental Chemistry-Anilkumae DE,

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MAHATMA GANDHI UNIVERSITY

SBS M P E 60: OMICS IN BIOTECHNOLGY

School Name	**School of Biosciences**
Programme	**MSc Biotechnology**
Course Name	**Omics in Biotechnology**
Type of Course	**Elective**
Course Code	**SBS M P E 60**
Course Summary & Justification	1.The course describes new approach, the concept of "OMICS" in various levels. It is a multi-disciplinary emerging field that encompasses genomics, epigenomics, transcriptomics, proteomics, and

	metabolomics. 2.The course content explain the high-quality techniques, methods & analysis from genome level will help in the complete understanding of a biological process. These approaches are targeted towards understanding complex systems more thoroughly at the molecular level.					
Semester	Fourth					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisites	Basics of Molecular Biology					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**On completing this course, the student will be able to Explain genome and types of genomics, tool and methods in genomic study, as well as Genome structure of selected organisms.**	**U/E**	
2.	**Explain the Proteomics, Transcriptomics & Metabolomics & Describe the tool and methods employed to study. Students have able to explain the various application of Proteomics, Transcriptomics & Metabolomics study**	**An/A**	
3.	**Students have able to illustrate the techniques employed for metagenomic analysis and application of metagenomic study**	**S/I**	
4.	**Describe the classification and types of databases & applications of data bases**	**U/R**	
5.	**Communicate effectively about a chosen topic in Omics in Biotechnology both practically and theoratically.**	**C/S**	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hrs

1	**Genome & Genomics**: Definition of Genome & Genomics.Types of genomics,, Functional Genomics.Structural genomics&Comparative genomics, Tools in Genomics,Structural genomics:-Classical ways of genome analysis, large fragment genomic libraries; Physical & Genetic mapping of genomes; Genome sequencing, sequence assembly, annotation& bioinformatics.Functional genomics:-DNA chips and their use in transcriptome analysis; Mutants and RNAi in functional genomicsNext generation sequencing methods; Structure of genomes: bacteria, yeast, nematode, Arabidopsis, rice, zebra fish, mouse and man.Applications of genomics	**1.0**	**20**
2.	**Proteomics,Transcriptomics & Metabolomics:** Basic concepts , Introduction to transcriptomics,proteomics and metabolomicsTools of proteomics- SDS PAGE, 2D PAGE , Liquid chromatography , Mass Spectrometry (ESI and MALDI) ,Protein identification by peptide mass fingerprinting ,Applications of proteomics-. Protein identity based on composition, Motifs and patterns, Analysis and characterization of proteins and metabolites:. Proteomics approaches to the analysis of protein-protein interactions, and metabolic profiling through emerging metabolomic techniques like 2D gel electrophoresis and Mass spectrometric and computational techniques.Applications of proteomics in agriculture, human health and industry	**0.5**	**10**
3.	**Metagenomics:** Definition of metagenomics**,**Techniques in metagenomics-Isolating DNA from an environmental sampleClone DNA,Insert into plasmid,Develop sample library,Screen or sequence,Analysis of metagenomic data.Application of metagenomics	**0.5**	**10**
4	**Biological data bases:** Classification databases. Biological databases-primary sequence databases- Composite sequence databases- Secondary databases-composite protein pattern databases,Pattern and profile databases Genome Information Resources: DNA sequence databases-specialized genomic resources, GRAIL, GENSCANProteome databases Protein sequence databases - SWISS-PROT and TrEMBL — PROSITE and BLOCKS - 2D PAGE databases – Structure databases - PDB- Metabolic databases – post translational modification databases	**1.0**	**20**
	Total Credits of the Course	**3**	

Teaching and Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** MM. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks NN. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Introduction to proteomics, Daniel. C. Libeler, Humana Press 2002
2. Thompson, J.D., Schaeffer-Reiss, C., and Ueffing, M. 2008. *Functional Proteomics. Methods andProtocols*. Humana Press, New York.
3. Metabolomics- Methods and Protocols by Wolfram Weckwerth, Humana Press.
4. Aurthur M Lesk Introduction to Bioinformatics .Oxford University press.

Further Reading:

1. Bostjan Koba.,Mitchell Guss & Thomas Habs Structural Proteomics.Humana Press.
2. Twyman, R.M. 2004. *Principles of Proteomics*. Taylor & Francis
3. Mass Spectrometry for Biotechnology by Gary Siuzdak, Academic Press.
4. Proteomics for Biological Discovery by Timothy Veenstra and John Yates, Wiley.
5. Lipidomics- Technologies and Applications by Kim Ekroos, Wiley-VCH.
6. Web/Journal Resources.
7. Transcriptomics: Expression Pattern Analysis, Virendra Gomase, Somnath Tagore; VDM Publishing, 2009 – Science
8. Brown TA. 2007. Genome III. Garland Science Publ.
9. Campbell AM & Heyer L. 2004. Discovery Genomics, Proteomics and Bioinformatics. Pearson Education.
10. Jollès P & Jörnvall H. 2000. Proteomics in Functional Genomics: Protein Structure Analysis.
11. Kamp RM. 2004. Methods in Proteome and Protein Analysis. Springer.
12. Primrose SB & Twyman RM. 2007. Principles of Genome Analysis and Genomics
13. Blackwell. Sensen CW. 2005. Handbook of Genome Research. Vols. I, II. Wiley CVH.

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MAHATMA GANDHI UNIVERSITY

SBS M P E 61: MOLECULAR PHYLOGENY

School Name	**School of Biosciences**
Programme	**Msc Biotechnology/ Biochemistry/ Biophysics/ Microbiology**
Course Name	**MOLECULAR PHYLOGENY**
Type of Course	Elective
Course Code	**SBS M P E 61**
Course Summary & Justification	1. This elective course deals with the tools and techniques of Molecular phylogeny. The course has a theoretical and a practical dimension 2. The learner will develop an understanding about models of nucleic acid substitution, tree building algorithms, data mining tools and submission tools for nucleic acid data and applications of Molecular phylogeny
Semester	Fourth

Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120

Pre-requisites	Basics of genome organisation and organic evolution, concepts of biological classification

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	**On completing this course, the students will be able to** **Compare and narrate the models of nucleic acid substitution, tree building algorithms, data mining tools, and submission tools for nucleic acid data**	An	
2	**Deposit nucleic acid sequences in databases and able to perform data mining**	S	
3	**Perform sequence alignment and editing**	S	

4	**Analyse sequence alignments by suitable software and perform phylogenetic analysis**	S	
5	**Carry out a phylogenetic analysis from raw sequence data up to final conclusions**	S	
6	**Communicate effectively about a phylogenetic problem both verbally and in writing.**	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Basic concepts of molecular evolution: Genetic information, population dynamics, evolution and speciation, data used for molecular phylogenetics, phylogenetic tree, methods for inferring phylogenetic trees, networking, RNA world	0.75	15
2	Sequence databases and data base searches: Sequence databases, composite databases, database mirroring, and search tools, data base searching by sequence similarity – BLAST and FASTA, multiple sequence alignments CLUSTAL, MUSCLE, T-COFFEE	0.5	10
3.	Phylogenetic inference: Genetic distances and nuclear substitution models, phylogenetic inference based on distance methods- UPGMA, Neighbour Joining, Minimum Evolution, Least square	0.5	10
4	Phylogenetic inference: Maximum Likelihood and Bayesian phylogenetic analysis, phylogenetic analysis based on parsimony, phylogenetic analysis using protein sequences, testing tree reliability – Bootstrapping and jackknifing	0.5	10
5	Testing models and trees: Models of evolution and phylogeny reconstruction, model fit, likelihood ratio tests, Practising MEGA, Paup*, RaxML, Mr Bayes, J Model Test, Sequence submission tools- SEQUIN and BankIt	0.75	15
	Total Credits of the Course	3	

Teaching and Learning Approach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative

Assessment Types	**Mode of Assessment** OO. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks PP. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Molecular evolution And Phylogenetics, Masatoshi Nei and Sudhir Kumar, Oxford University Press, ISBN 0195135857
2. Baldauf, SL (2003) "Phylogeny for the faint of heart: a tutorial." Trends in Genetics; 19(6):345-351.

Further Reading:

3. The phylogenetic Hand book, 2nd Edition, Philippe Lemey, Marco Salemi, Anne –Mieke Vandamme, Cambridge University Press, ISBN-13 978-0-511-71963-9
4. Hall, BG. (2004) Phylogenetic Trees Made Easy: A How-To Manual, 2nded. Sinauer Associates, Inc.: Sunderland, M A. I*SBN: 978-0-87893-606-9*
5. Hartwell, LH, L Hood, ML Goldberg, AE Reynolds, LM Silver, RC Veres (2008) Genetics: From Genes to Genomes, 3rd Ed. McGraw-Hill: New York ISBN-13: 978-0073525266 ISBN-10: 007352526X

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MAHATMA GANDHI UNIVERSITY

	SBS M P E 62: PLANT MICROBE INTERACTIONS

SchoolName	**School of Biosciences**					
Programme	M.Sc. Microbiology					
Course Name	**PLANT-MICROBE INTERACTIONS**					
Type of Course	Elective					
Course Code	**SBS M P E 62**					
Course Summary & Justification	This course develops concepts in plant- microbe interaction The major objective of this paper is to give an insight into the consequences, on population and ecosystem level, of compatible and incompatible interactions, to understand infection process and control measures and to familiarize with the microbial production of plant metabolites.					
Semester	Fourth					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics of agricultural microbiology**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Comprehensively discuss interactions between plants and microbes as well as the defense reactions of the host plant	U/R/ An	
2	Gain insight into genetics of host-pathogen interactions and resistance mechanism in plants.	C/ I/An	
3	Comprehend various methods to analyse plant diseases and biological methods of disease control	S/An/A	
4	Analyse why plants and microbes react in certain ways in pathogenic and symbiotic interactions	U/R/An	

5	Understands the role of microbes in developing plant immunity	U/R	
6	Have an in-depth knowledge on biopesticides and their role in pest control	An/ C	
Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)			

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Different interfaces of interactions -soil-plant-microbe interactions leading to symbiotic (rhizobial and mycorrhizal), associative, endophytic and pathogenic interactions	0.5	10
2	General concepts of plant immunity. PAMP-triggered immunity (PTI) and Effector triggered immunity (ETI). Outer membrane vesicles (OMVs) and their involvement in plant immunity. The type III secretion system. Hypersensitive response. Genetic basis of plant defences. Quorum-sensing in bacteria and their role in plant defence mechanisms. Phytohormones and antibiotics as plant therapeutics.	1.0	20
3.	Plant pathogens and molecular basis of pathogenesis .Genetics of host-pathogen interactions, resistance genes, resistance mechanisms in plants. basal and induced defence mechanisms. Systemic Acquired Resistance (SAR) and Induced Systemic Resistance (ISR), Recognition mechanism and signal transduction during plant - pathogen interaction. Virulence determinants of plant pathogenic bacteria-Enzymes, Toxins, pili, siderophores, secretion systems	1.0	20
4	Microbial pest control: Bacillus thuringiensis-mode of action, Biocontrol agents– uses and practical constraints Biofungicide and bioherbicides. Plant growth promoting rhizobacteria. Use of plant–microbe symbiosis for remediation of pollutants and carbon (C) sequestration	0.5	10
	Total Credits of the Course	3	

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction: Active co-operative learning, Seminar, Group Assignments Authentic learning: Library work and Group discussion, Presentation by individual student/ Group representative

Assessment Types	**Mode of Assessment** C. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks D. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Subba Rao, N.S. 2005. Soil Microorganisms and Plant Growth, Oxford and IBH Publishing Co.
2. B. Lugtenberg (ed). 2015.Principles of plant microbe interactions, Springer

Further Reading:

1. Microbial control and pest Management – S.Jayaraj.
2. Paul, E.A. 2007. Soil Microbiology, Ecology and Biochemistry, Academic Press.
3. M.Gillings and Holmes .2004.Plant microbiology-Bios Scientific publishers.
4. Kosuge T & Nester EW. 1989. Plant-Microbe Interactions: Molecular and Genetic Perspectives .Vols I-IV. McGraw Hill.
5. Verma DPS & Kohn TH. 1984. Genes Involved in Microbe-Plant Interactions. Springer Verlag.
6. Gary Stacey, Noel T. Keen, 1995. Plant-Microbe Interactions. Vols I-VI Springer Science & Business Media.
7. Jeng-Sheng Huang **2001.**Plant Pathogenesis and ResistanceBiochemistry and Physiologyof Plant-Microbe Interactions .Springer Verlag

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MAHATMA GANDHI UNIVERSITY

SBS M P E 63: HUMAN VIROLOGY

SBS M IV E 17 69 HUMAN VIROLOGY

SchoolName	**School of Biosciences**					
Programme	**M.Sc Microbiology/Biochemistry/Biotechnology/Biophysics**					
Course Name	**HUMAN VIROLOGY**					
Type of Course	Elective					
Course Code	**SBS M P E 63**					
Course Summary & Justification	This course on Human Virology deals with an important area of Medical Microbiology The objective of the course content is to create a sound awareness in human viruses and viral diseases. their The course will augment the student's knowledge in pathogenesis of viral diseases and their laboratory diagnosis and prophylaxis.					
Semester	Fourth					
Total StudentLearningTime (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total LearningHours
	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	Basic understanding on Human Anatomy, Physiology and Biochemistry Knowledge in Basic Virology, Molecular Biology and Immunology					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1.	On completing this course student will be able to analyse comparatively the structure and properties of important human viruses	**U/An**	
2.	Students will be able to understand and evaluate the mechanism of pathogenesis of viral diseases	**U/E**	
3.	Students will become aware of the methods applicable in viral diagnostics	**U/A**	
4.	Students will be able to analyse the various mechanisms of viral oncogenesis	An	
5,	Students will be able to understand and compare the mechanisms of action of various antiviral agents	U/An	
6	Students will be able to understand and evaluate the	U/E	

	methods of prophylaxis of viral diseases in humans		

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Study of properties of human DNA viruses viz. Pox, Herpes, Adeno, Papova, and Parvo viruses. Pathogenesis and laboratory diagnosis of diseases caused by these viruses	0.5	10
2	Study of properties of human RNA viruses viz. Picorna, Orthomyxo, Paramyxo, Rhabdo, and Rubella viruses	1.0	20
3.	Arboviruses and Hepatitis viruses - Properties. Pathogenesis and laboratory diagnosis of diseases caused by these viruses. Viral haemorrhagic fevers, SARS CoV-2, HIV,Properties, pathogenesis and laboratory diagnosis of Slow virus infections, Prion diseases	1.0	20
4	Viruses and cancer, Viral oncogenesis, Viruses implicated in the cancers of humans, Prophylaxis of viral diseases, Types of viral vaccines, antiviral agents and their mechanisms of action, Interferons	0.5	10
	Total Credits of the Course	3	

TeachingandLearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** E. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks

	F. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading:

1. Jawetz,Melnick& Adelberg's Medical Microbiology27 th Edition Carrol, Butel, Morse, Mietzner Mc Graw Hill

3. Ananthanarayan & Panicker's Text book of Microbiology.9th Edition Arti Kapil (Ed) University Press (India) Pvt.Ltd.

Further Reading:

Further Reading:

1. Human Virology Fourth Edition Leslie Collier, John Oxford & Paul Kellam University Press.

2. Fundamental Virology 5th Edition David M.Knipe& Lippincott Williams & Wilkin

3. Viruses Biology,Applications & Control

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MAHATMA GANDHI UNIVERSITY

SBS M P E 64: PHYSIOLOGICAL BIOPHYSICS

School Name	**School of Biosciences**
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**
Course Name	**PHYSIOLOGICAL BIOPHYSICS**
Type of Course	Elective
Course Code	**SBS M P E 64**
Names of Academic	**Dr Harikumaran Nair R**

Staff & Qualifications						
Course Summary & Justification	The course is designed to provide the fundamental principles of modern physiology, protein science and structural biology, and to prepare students for higher learning and answer questions like :-How do solutes transport across cell membranes? What is the ionic basis of the membrane potential? How does the cell membrane behave like an electrical circuit? What is the molecular physiology of muscle contraction? What are the mechanisms of hemodynamic? What is the biophysical property of lung mechanics?					
Semester	Fourth					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	E.g., Authentic learning Collaborative learning Independent learning	54	18	0	28	100
Pre-requisite	**Basic Knowledge in Biosciences**					

No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**
	Understand basic level of cell physiology3. 4.	U	
	Explain cell transport and communication in a cell	R	
	Explain how lung and cardiac dynamic property is important to sustain life.	R	
	Understand force generating capacity of muscles	U	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

Module No	Module content	Credits offered

	Chemical composition of body, movement of molecules across cell membranes, control of cells by chemical messengers	0.5
	Design of circulatory system, pressure, flow and resistance in circulatory system, physical characteristics of blood, haemo dynamics principles & equations, genesis & spread of cardiac impulse, cardio dynamics, regulation of blood pressure & blood volume, heartbeat coordination, mechanical events of cardiac cycle, cardiac output, cardiovascular responses to stress	1
	Lung mechanics, ventilation, gas exchange process, gas diffusion, gas transport, pulmonary circulation, neural generation of rhythmical breathing, control of respiration by partial pressure of gases and hydrogen ion concentration, ventilation in response to stress, pulmonary function tests	0.5
	Ionic composition & distribution of body fluids, division of labour in kidney tubules, concept of renal clearance, regulation of sodium, water and potassium balance, calcium regulation, hydrogen ion regulation, renal mechanics, acidosis, alkalosis, basic concepts of energy expenditure, regulation of total body energy stores, regulation of body temperature. Molecular mechanism of muscle contraction, mechanics of single-fiber contraction and whole muscle contraction, muscle energy metabolism, control of body movement, maintenance of upright posture and balance, walking, vestibular system and equilibrium, state of consciousness, motivation and emotion, cerebral dominance and language	1
	Total Credits of the course	3

Books for References

Compulsory Reading

1. Brobeck J.R, Best and Taylor's Physiological bases of medical practice
2. Basar E, Biophysical and physiological system analysis
3. Guyton A.C, textbook of Medical Physiology
4. Robert Glambos, Nerves and muscles

Further Reading:

1. Biophysics A Physiological Approach, Patric F Dillon (2012)
2. Comprehensive Biophysics, Volume I-IX, Edward H Egelman (2012)
3. Introduction to experimental Biophysics, Jay Nadeau (2012)
4. Physiology, Biophysics and Biomedical Engineering, Andrew W Wood (2012)

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MAHATMA GANDHI UNIVERSITY

SBS M P E 65 GOOD LABORATORY PRACTICES

SchoolName	**School of Biosciences**					
Programme	**MSc Biochemistry/ Microbiology/ Biotechnology/ Biophysics**					
Course Name	**GOOD LABORATORY PRACTICES**					
Type of Course	Elective					
Course Code	**SBS M P E 65**					
Course Summary & Justification	To equip the students with appropriate knowledge, skills to undertake general and quality management of laboratory practices and procedures. To adequately address quality issues and improve the overall delivery of clinical and public health laboratory services in their facilities/organizations. To sensitize the students with medical and public health ethics issues and to ensure its application in teaching and practice.					
Semester	Fourth					
Total StudentLearning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours

	Authentic learning Collaborative learning Independent learning	60	20	0	40	120
Pre-requisite	**Basics Knowledge in Biosciences**					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	Understand basic good laboratory practice	U	
2	Appreciate how to conduct research safely and efficiently	Ap	
3	Understand the requirements for safe working practices and risk assessment	U	
4	Apply experimental design and the need for controls	A	
5	Consider ways in which student can maximise research effort	C	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hrs
1	Introduction to good laboratory practices (GLP) and its application, history of GLP, fundamental points of GLP	0.5	10
2	Resources-personnel, Facilities - buildings and equipment, Characterization- test item, test system, rules for performing studies-the study plan or protocol, standard operating procedures (SOPs) raw data and data collection- records and recording, study report, archives and archiving, quality assurance, audit and inspections, implementation of GLP	1	20
3.	Applications of the GLP principles to field studies, applications of the GLP principles to short term studies, applications of the GLP principles to in vitro studies	0.5	10
4	Ethics in research- locating ethics in research, justice in research, science and society, ethical issues in biotechnology, ethical guidelines related to human experimentation, guidelines regarding animal use in research, institutional biosafety monitoring mechanisms.	1	20
	Total Credits of the Course	3	60

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction: Brain storming lecture, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments Authentic learning, , Library work and Group discussion, Presentation by individual student/ Group representative

Assessment Types	Mode of Assessment A. Continuous Internal Assessment (CIA) 1. Internal Tests of maximum 20 marks 2. Seminar Presentation – a theme is to be discussed and identified to prepare a paper and present in the seminar Maximum marks 10 3. Write a detailed report on a given topic based on research findings and literature search – 10 marks B. **Semester End examination – 60 marks**

REFERENCES

Compulsory Reading

1. Handbook on Good Laboratory Practice- World Health Organization
2. Ethical Guidelines for Biomedical Research on Human Participants- Indian Council of Medical Research
3. Guidelines on the regulation of scientific experiments on animals- Ministry of Environment and Forests, India
4. Textbook on Ethics in Research- European Commission, Publications Office of the European Union

Further Reading:

1. **Good Laboratory Practice Regulations, 4th edition edited By Sandy Weinberg- CRC Press, 2007**
2. **The Indispensable Guide to Good Laboratory Practice (GLP): Second Edition 2nd Edition- Mark Gregory Slomiany- Springer, 2009**

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MAHATMA GANDHI UNIVERSITY

SBS M PE 66: MEDICAL BIOPHYSICS

School Name	**School of Biosciences**
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**
Course Name	**MEDICAL BIOPHYSICS**

Type of Course	Elective					
Course Code	**SBS MP E 66**					
Names of Academic Staff & Qualifications	**Mrs. Resmi S S**					
Course Summary & Justification	The course is to introduce the student to important areas of medical Biophysics like Bioelectric signals, Laser, Medical imaging, Sonography, Fluoroscopy, Nuclear medicine, Radiation therapy and ergonomics. and to get an insight on how experimental methods and theoretical approaches from physics can give answers related to the structure and functions of biological system.					
Semester	Fourth					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	E.g., Authentic learning Collaborative learning Independent learning	54	18	0	28	100
Pre-requisite	**Basic Knowledge in Biosciences**					

No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**
	Explain bioelectric signals and its recording Describe types of electrodes, their design, properties and uses.	R	
2	Understand principle of operation of LASER and its applications	U/A	
3	Explain different types of imaging technique and applications of nuclear medicine	U/ An	
4	Describe the importance of radiotherapy	U	
5	Narrate different areas of ergonomics	U	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

Module No	Module content	Credits offered
	Origin and Characteristics of Bioelectric signals & recording, Electrodes, types Design and properties and Utility, Skin contact impedance of Electrodes, noise suppression techniques, recording system, Medical Display systems, Patient Monitoring systems, Biomedical Telemetry, Computer Applications in medical field, Patient Safety. Cardiac pace makers, Defibrillators, Hemodialysis machines, Short wave and Micro wave Diathermy, Ultrasonic Therapy, Pain relief through electrical stimulation, Surgical Diathermy, Laser, principle of operation, Types, Laser tissue interaction, Biomedical applications in surgery and therapy	1.0
	Principle, Working of Blood flow Meters, Pulmonary function analyzers, Blood gas analyzer, Oximeters, Audiometer. Medical-Imaging Techniques. Physical aspects of Medical-imaging, Principle, Practical System, Medical utility of X-ray imaging, Fluoroscopy, Xeroradiography, Computerized Axial Tomography, Mammography, Angiography, Myelography, Magnetic resonance imaging, Ultrasonography	1.0
	Basic principles of Nuclear Medicine, Diagnostic use of Radioisotopes Invivo& In-vitro procedures, (Single isotope, Double isotope methods) , Radio immunoassay counting system, General principles & procedures of organ scanning, Renal imaging, Cardiac imaging, Thyroid scanning, Blood volume determination by isotope method, Rectilinear scanners & Gamma scintillation camera, Positron emission Tomography (PET), Single Photon emission computer Tomography (SPECT), Radio pharmaceuticals & their Diagnostic applications	0.5
	Concepts of teletherapy & Brachytherapy, Co-60 Therapy, Basic principles & scope of radio therapy, Benign & Malignant tumors, Tissue tolerance dose &Tumor lethal dose, Medical dosometry, Dose fractionation, Palliative & Curative therapy, Treatment planning, Isodose distribution, Patient data, Correction & Setup, Field shapping, Skin dose and field separation, brachytherapy, Sources, Calibrations, Dose	0.5

	distribution implant dosimetry. LINAC (Linear accelators). Ergonomics, Muscle mechanics, Load velocity relation, Length tension relation, Entire State, Role of elastic components in muscle contraction, Ergonomic problems of computer users.	
	Total Credits of the course	3

Books for References

Compulsory Reading:

.Hand book of Biomedical Instrumentation: R.S Khandpur,Tata McGraw-Hill Publishing company Ltd

2. Biomedical Instrumentation and measurements:Leslie Cromwell, Fred.J. Weibell,Erich. A.Pfeiffer. Prentice-Hall of India Private Ltd

Further Reading:

1. Bioinstrumentation: John.G.Webster.Wiley-India
2. 2. Medical Physics: Martin Hollins.University of BATH

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MAHATMA GANDHI UNIVERSITY

SBS MP E 67: BIOFERTILIZERS AND BIOPESTICIDES

School Name	**School of Biosciences**
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**

Course Name	**BIOFERTILIZERS AND BIOPESTICIDES**					
Type of Course	Elective					
Course Code	**SBS MP E 67**					
Names of Academic Staff & Qualifications	**Dr J G RAY**					
Course Summary & Justification	The course is to introduce the concept of biofertilizers and biopesticides,familiarize different agriculturally important microorganisms which are being used as biofertilizers for maintaining the soil and plant health, and conventional biopesticides and the basic chemistry and action of the same					
Semester	Fourth					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	E.g., Authentic learning Collaborative learning Independent learning	54	18	0	28	100
Pre-requisite	**Basic knowledge in soil and farming**					

No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**
	Develop a critical knowledge on the concept of soil fertility, fertilizers and pesticides	R/U	
2	Analyse environmental significance of biofertilizers and biopesticides	An	
3	Understand the important soil microbes beneficial to soil fertility	U	
4	Develop the skills to prepare biopesticides biofertilizers of diverse kinds	C	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

Module No	Module content	Credits offered
	Different agriculturally important beneficial microorganisms – free living, symbiotic (rhizobial, actinorhizal), associative and endophytic nitrogen fixers including cyanobacteria, taxonomic classification, nodule formation, competitiveness and quantification of N2 fixed	1
2	Different agriculturally important beneficial microorganisms – phosphate solubilizing bacteria and fungi, including mycorrhiza; Different agriculturally important beneficial microorganisms – plant growth promoting rhizobacteria, Different agriculturally important beneficial microorganisms – Biocontrol microbial inoculants; Different agriculturally	2

	important beneficial microorganisms for recycling of organic waste and compositing, bioremediators and other related microbes	
	Different agriculturally important beneficial microorganisms - selection, establishment, competitiveness, crop productivity, soil & plant health, mass scale production and quality control of bio inoculants. Biofertilizer inoculation and microbial communities in the soil.	3
	Conventional natural insect control agents such as pyrethrins, rotenones, nicotine, ryanodine, isobutylamides, drimane sesquiterpenoids, withanolides, clerodanes, quassinoids and limonoids - sources, isolation, characterization, synthesis, application and mode of action Phytoalexins, stress metabolites: Sources such as Leguminosae, Solanaceae etc. Acetylene and polyacetylene phytoalexins Pesticides of microbial origin : Sources, chemistry and mode of action of tetranactin, avermectins, milbimycins and spinosad. Herbicides like biolaphos and phosphonothricin. Phytotoxins like Alternaria alternata toxin, tentoxin, cornexistin, hydantoxidin. Other microbials such 1.5105 as NPV based insecticides Allelochemicals and chemical ecology. Application of biotechnology in pest management (ex. Bt)	4
	Total Credits of the course	3

Books for References

Compulsory Reading:

13. Sylvia DM, Fuhrmann JJ, Hartlly PT & Zuberer D. 2005. Principles and Applications of Soil Microbiology. 2nd Ed. Pearson Prentice Hall Edu.
14. Copping LG. 1996. Crop Protection Agents from Nature: Natural Products and Analogues. Royal Soc. Chem., London

Further Reading:

15. van Elsas JD, Trevors JT & Wellington EMH. 1997. Modern Soil Microbiology. CRC Press
16. Bergerson FJ. 1980. Methods for Evaluating Biological Nitrogen Fixation. John Wiley & Sons

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SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**HEALTH AND NUTRITION**					
Type of Course	Elective					
Course Code	**SBS M P E 68**					
Course Summary & Justification	The course is designed to provide basic information on nutrition and its importance in providing health.					
Semester	Fourth					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	Eg. Authentic learning Collaborative learning Independent learning	50	30	0	40	120
Pre-requisite	Basic understanding of food and food ingredients					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To describe the basic principles of nutritional biochemistry and different methods of nutritional analysis.	R/U	
2	To identify and compare the different ingredients and nutritional value of food components	A	
3	To identify different diseases associated with nutritional deficiency and overnutrition	U	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hours
1	**Introduction to nutrition -** Food as source of nutrients, functions of food, definition of nutrition, nutrients & energy, adequate, optimum & good nutrition, malnutrition. Basics of energy metabolism, nutrition & dietetics - Unit of measuring energy, calorific value of food, BMR & factors affecting it, SDA of food, calculation of energy requirement, balanced diet, nutrition in health & disease. Nutritional disorders- Epidemiology, clinical features, prevention and dietary treatment for Protein Energy malnutrition, nutritional anaemias.	1	15

2	**Food sources:** Carbohydrates : Functions, classification, food sources, storage in body. Fats & oils : composition, saturated and unsaturated fatty acids, classification, food sources, function of fats. Proteins - composition, sources, essential & non-essential amino acids, functions, Protein deficiency	0.5	10
3.	**Water, Vitamins and minerals-** Water - as a nutrient, function, sources, requirement, water balance & effect. Minerals - macro & micronutrients. - functions, sources. Bioavailability and deficiency of Calcium, Iron, Iodine, Sodium & Potassium (very briefly). Vitamins (water & fat soluble) - definition, classification & functions. Effect of cooking & heat processing on the nutritive value of foods. Processed supplementary foods.	0.5	10
4	**Nutritional problems affecting the community-**Etiology, prevalence, clinical features and preventive strategies of-Undernutrition - Protein energy malnutrition: Nutritional Anaemias, Vitamin A Deficiency, Iodine Deficiency Disorders. Overnutrition – obesity, coronary heart disease, diabetes. Fluorosis	1	15
	Total Credits of the Course	3	

Books for Reference

Compulsory Reading:

1. Mudambi, SR and Rajagopal, MV. Fundame ntals of Foods, Nutrition and Diet Therapy; Fifth Ed; 2012; New Age International Publishers
2. Mudambi, SR, Rao SM and Rajagopal, MV . Food Science; Second Ed; 2006; New Age Publ.

Further Reading:

1. Srilakshmi B. Nutrition Science; 2012; New Age International (P) Ltd.
2. Swaminathan M. Handbook of Foods and Nu trition; Fifth Ed; 1986; BAPPCO.
3. Bamji MS, Krishnaswamy K and Brahmam GNV (Eds) (2009). Textbook of Human Nutrition, 3rd edition. Oxford and IBH Publishing Co. Pvt. Ltd. New Delhi.

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, Presentation by individual student/ Group representative

Assessment Types	**Mode of Assessment** **C. Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks **D. Semester End examination – 60 marks**

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MAHATMA GANDHI UNIVERSITY

SBS M P E 69: NEUTROPHIL BIOLOGY

SchoolName	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**NEUTROPHIL BIOLOGY**					
Type of Course	Elective					
Course Code	**SBS M P E 69**					
Course Summary & Justification	The course is designed to get a detailed idea about the functioning of neutrophils in providing immune response and the mechanisms behind it. This would be helpful for the students, in case they take up research in immunology, cell biology or cellular biochemistry.					
Semester	Fourth					
Total Student Learning Time	Learning Approach	Lecture	Tutorial	Practical	Others	Total

(SLT)						Learning Hours
	Eg. Authentic learning Collaborative learning Independent learning	50	20	10	40	120
Pre-requisite	Basic understanding of immunology and blood cells					

COURSE OUTCOMES (CO)

CO No.	Expected Course Outcome	Learning Domains	PSO No.
1	To describe the role of neutrophils in imparting and fine-tuning immune response	R/U	
2	To identify and compare different functions of neutrophils	U/A	
3	To identify different techniques to perform neutrophil functional analysis	S	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

COURSE CONTENT

Module No	Module Content	Credits	Hours
1	**Introduction to immune system-** innate and adaptive immune system, cells involved in immune system, humoral immunity, cytokines, antibodies, complement system. cell-mediated and humoral immune response	0.5	10
2	**Neutrophil Physiology**-Neutrophil structure, Granule types- azurophilic, specific, gelatinase, secretory vesicles, Antimicrobial peptides. Neutrophil Subpopulations. Neutrophil activation, apoptosis and clearance. Neutrophils in the resolution of inflammation. Neutrophil in immune cross-talk	0.5	10
3.	**Neutrophil defense mechanisms-** Chemotaxis, Phagocytosis, degranulation, ROS generation,NADPH oxidase, Neutrophil extracellular trap formation, NETosis vs. apoptosis and necrosis, Cytokine secretion. Diseases associated with altered neutrophil defence- Autoimmunity, cancers, thrombosis.	1	15
4	**Techniques to study neutrophils:** Neutrophil isolation and maintenance, Cell counting, Phagocytic assays, chemotactic assays, NBT assay, MTT assay, other assays of ROS production, Granule isolation, Neutrophil protein analysis, microscopic analysis of neutrophils and granules – Light and fluorescent microscopy, SEM and TEM	1	15
	Total Credits of the Course	3	

Books for Reference
Compulsory Reading: 1. Neutrophil Methods and Protocols, Quinn, Mark T., DeLeo, Frank R., Bokoch, Gary M. (Eds.). ISBN 978-1-59745-467-4. 2. Biochemistry and physiology of the neutrophil, Steven W Edwards, Cambridge university press Online ISBN-9780511608421 3. The Neutrophil, Murphy, Patrick , Springer, ISBN- ISBN 978-1-4684-7418-3
Further Reading: 1. Neutrophil function: Mechanisms to diseases. Borko Amulic, Christel Cazalet,Garret L. Hayes, Kathleen D. Metzlerand Arturo Zychlinsky; Annu. Rev. Immunol. 2012. 30:459–89. 2. Neutrophil biology: an update. Yoshiro Kobayashi, EXCLI J. 2015; 14: 220–227. doi: 10.17179/excli2015-102. 3. Advances in neutrophil biology: clinical implications. Cowburn AS, Condliffe AM, Farahi N, Summers C, Chilvers ER. Chest. 2008 Sep;134(3):606-12. doi: 10.1378/chest.08-0422. 4. The Neutrophils: New Outlook for Old Cells. 3rd Edition.Edited by: Dmitry Gabrilovich (H Lee Moffitt Cancer Center, USA & University of South Florida, USA). ISBN: 978-1-84816-836-7

Teachingand LearningApproach	**Classroom Procedure (Mode of transaction)** Direct Instruction, Explicit Teaching, E-learning, interactive Instruction:, Active co-operative learning, Seminar, Group Assignments, Authentic learning, Library work and Group discussion, demonstrations, Presentation by individual student/ Group representative
Assessment Types	**Mode of Assessment** **A. Continuous Internal Assessment (CIA)** Internal Test -20 marks Assignment – Every student needs to write an assignment on a given topic based on the available published literature – 10 marks Seminar Presentation – A topic needs to be presented and discussed with the class- 10 marks **B. Semester End examination – 60 marks**

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SBS MP E 70: MEDICINAL PLANTS

School Name	**School of Biosciences**					
Programme	**M.Sc. Biochemistry/Microbiology/Biotechnology/Biophysics**					
Course Name	**MEDICINAL PLANTS**					
Type of Course	Elective					
Course Code	**SBS MP E 70**					
Names of Academic Staff & Qualifications	**Dr J G RAY**					
Course Summary & Justification	The course is introduce the significance of medicinal plants of ethno-medicine in modern research, familiarize highly valuable medicinal plants for diverse medicinal uses and help biotechnology students to learn more technological applications of plants					
Semester	Fourth					
Total Student Learning Time (SLT)	Learning Approach	Lecture	Tutorial	Practical	Others	Total Learning Hours
	E.g., Authentic learning Collaborative learning Independent learning	54	18	0	28	100
Pre-requisite	**Basic knowledge in Plant Science**					

No.	**Expected Course Outcome**	**Learning Domains**	**PSO No.**
	Develop a critical knowledge on the significance of ethno-medicinal knowledge	R/U	
	Analyse modern applications of ethnomedicines	An	
	Understand the important indigenous medicinal plants of Kerala	U	
	Develop the skills to apply the ethno-medicinal knowledge in the modern way	C	

Remember (R), Understand (U), Apply (A), Analyse (An), Evaluate (E), Create (C), Skill (S), Interest (I) and Appreciation (Ap)

Module No	Module content	Credits offered
	Introduction to Herbal Medicines: Principles of identifying medicinal plants, Basics of the botanical description – basic principles of morphology and taxonomy – Plants as medicines in Ayurveda – Unani – Siddha and Homeopathy - Ethno botany Major Indian plants known as Antiseptic, Anti-allergic and Expectorants **Botanical descriptions** – cultivation, processing as crude remedies and basic knowledge of the phyto-chemistry: *Eclipta alba, Mentha piperita, Aloe vera, Melia azadirachta, Coscinium fenestratum, Syzigium aromaticum, Sesamum indicum, Aegle marmelos, Ruta graveloens, Curcuma longa, Curcuma aromatica, Curcuma celosia, Pterocarpus santanilus, Ricinus communis, Lawsonia inermis and Ophiorrhiza mungos; Expectorants: Adathoda beddomei, Tylophora indica, Terminalia chebula, Ocimum sanctum, Ocimum basilicum, Eucalyptus globulus, Clitoria ternatea, Glycorrhiza glabra, Kaempferia galanga, Piper longum and Piper nigrum*	1.0
	***Indian Hallucinogenic, toxic and perfume-yielding plants**: botanical descriptions – cultivation, processing as crude remedies and basic knowledge of the phyto-chemistry: Papaver somniferum, Datura alba, Nerium oleander, Strychnos nux-vomica, Cliestanthus colinus, Cannabis sativa, Gloriosa superba, Anamirta cocculus, Citrulus colocynthis, Abrus precatorius, Semecarpus anacardium, Excoecaria agallocha, Digitalis purpurea, Aconitum ferox, Croton triglium, Plumbago zeylanica, Jatropa gossypifolia, Euphorbia neerifolia, Parthenium hyssterophorus and Arisaema triphyllum*	0.5
	Indian plants known as Nerve tonics - botanical descriptions – cultivation, processing as crude remedies and basic knowledge of the phyto-chemistry: Nerve tonics: Centella asiatica, Coriandrum sativum, Acorus calamus, Cardiospermum halicacabum, Allium cepa, Allium sativum, Cymbopogon citratus, Moringa olefera, Crocus sativus, Sida cordifolia, Bacopa monnieri, Withania somnifera, Solanum nigrum, Plumbago zeylanica, Vitex negundo, Samadera indica, Cynodon dactylon	0.5
	Indian Medicinal plants for digestive problems and liver remedies (silagogues, carminatives, febrifuges, digestives, hepatoprotectives and laxatives): botanical descriptions – cultivation, processing as crude remedies and basic knowledge of the phyto-chemistry: *Tamarindus indica, Trigonella foenum-graceum, Solanum xanthocarpum, Coleus aromaticus, Abelmoschus moschatus, Syzigium cumini, Elettaria cardomomu, Cuminum cyminum, Punica granatumm, Curcuma amada, Ferula asafetida, Oxalis corniculata, Cinnamomum zeylanicum, Vernonia cinerea, Tinospora cordifolia, Andrographis paniculatus, Phyllanthus niruri, Phyllanthus emblica, Terminalia bellerica, Zingiber officianalis, Achyranthes aspera, Carica papaya, Casia alata and Boerahaavia diffusa.*	1.0
	Total Credits of the course	3
Books for References		

Compulsory Reading:

1. Tribal medicines by Pal DC and Jain SK, Naya Prakash Publishers, Calcutta
2. Hand book on herbal drugs and its plants sources, H Panda, National Institute of Industrial Research, Delhi

Further Reading:

3. Glossary of useful and economically important plants, Ashok K Panigrahi and Alaka Sahu, Central Book Agency, Calcutta.
4. Indian Medicinal Plants Vol ! and II, PS Warrier, Orient Longman
5. Medicinal Plants of India with special reference to Ayurveda, CKN Nair and N Mohanan, Nag Publishers, Delhi.
6. Indian Materia Medica Vol: 1 by, Dr. K.M. Nadkarani, Publisher: Popular Prakash, Mumbai

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